

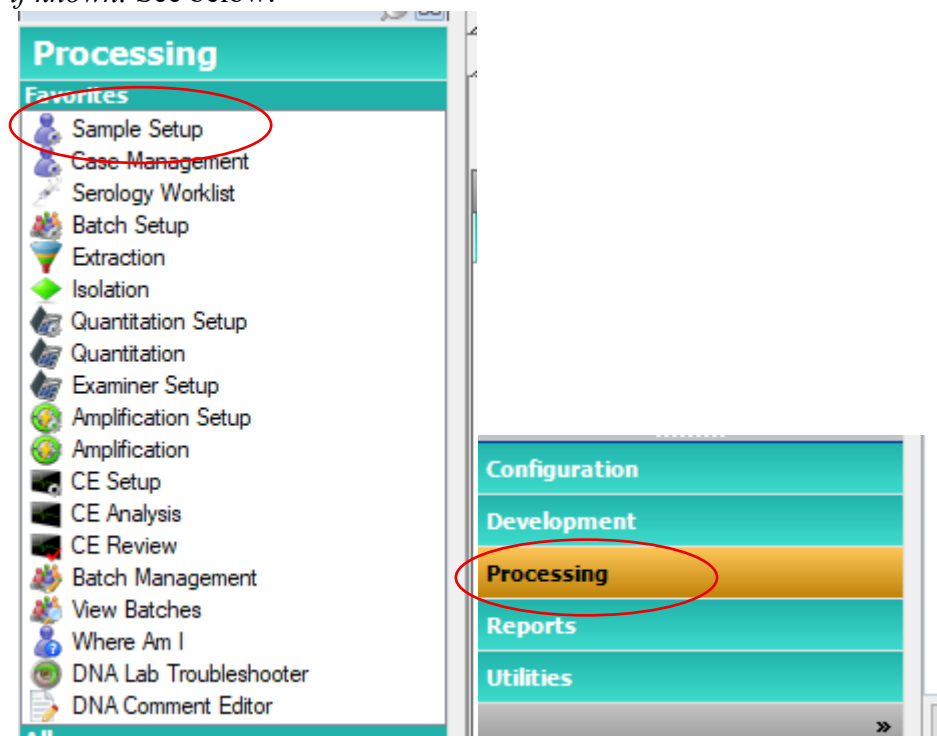
DNA Evidence Processing

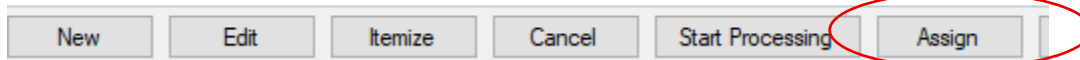
I. Batch Setup - Differentials

The Batch Setup module will be used to create an extraction batch and allocate samples/controls to the worksheet.

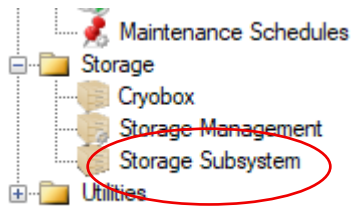
If the batch contains non-differential + differential samples, follow the *STACS Casework Non-Differential* instructions for the applicable samples. The non-differential samples may be added to the A-fraction isolation batch.

1. Upon batch assignment, assign the Justice-Trax STACS request to the processing examiner.
2. In STACS, assign the appropriate samples to the processing DNA examiner. This can be done in **Sample Setup**, under the **Sample Status** dropdown → **Processing**. Select the appropriate samples and choose **Assign**. Until this is done the sample will be assigned to the FB analyst who imported the sample. *The Reporting Analyst may be selected at this if known.* See below.

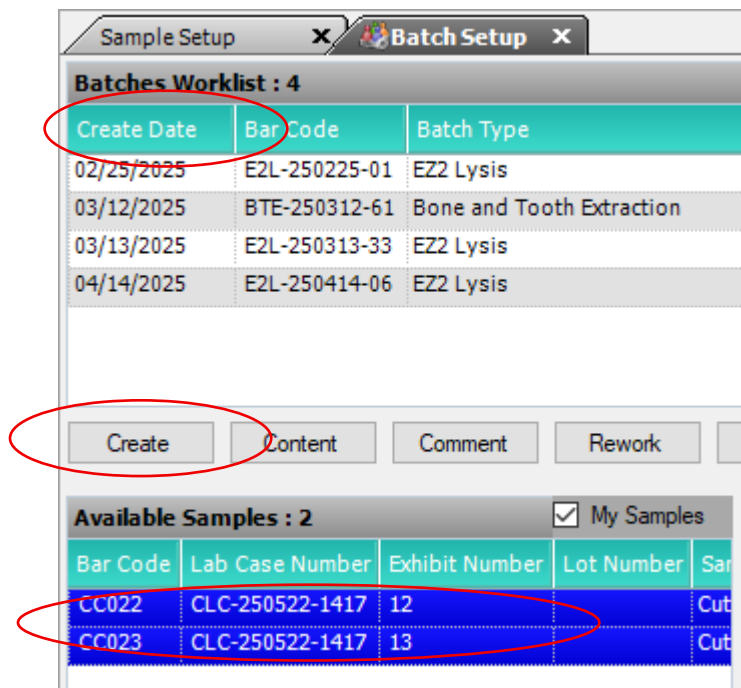


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3. Take custody of the sample tube(s) to be processed. In **Storage Subsystem (Utilities → Storage → Storage Subsystem)**, select the **Retrieve** tab, scan the item bar code(s) to be put into your custody. Select **Save**.



4. Create a lysis worksheet: Open **Processing → DNA Processing → Batch Setup**. Click **Create**.



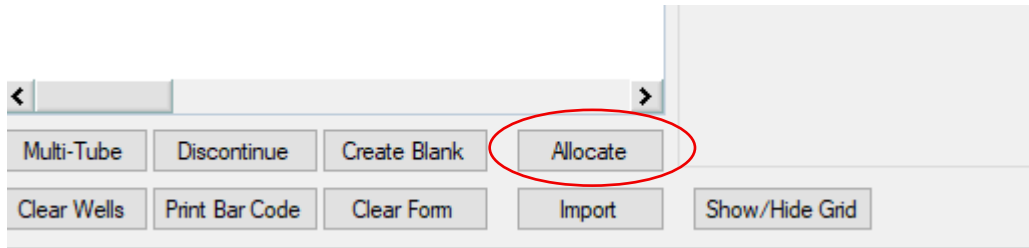
5. Your samples should be in the **Available Samples** area above. Make sure to check the “My Samples” box to only show samples assigned to you. Click **Create**.

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6. A **Create Batch** screen will appear as shown below.
7. Select the appropriate extraction under **Batch Type** (QIAcube Differential Lysis: **XX** samples), and the corresponding **Evidence Classification** (Question).
 - a) Select the **total** number of questioned samples, EP2 and reagent blanks as the total sample number scenario.
For example: 1 Q + EP2 + 2 RBs = 04 Samples
(QIAcube Differential Lysis: 04 Samples)

	1	2	3	4
A	DNA	BIS	Blank	Blank
B	DNA	DNA	DNA	DNA

8. Click **Create** (see above). A unique barcode associated with the extraction will be generated and printed. Moving forward, this barcode may be used to transfer the samples as a whole set. **Close** the Create Batch window at the bottom right corner.
9. To begin sample allocation, the batch should automatically populate into the “Sample Allocation” window. If it does not, scan the extraction barcode or double-click the batch in the upper window.
10. Select samples to be extracted from the available samples window on the left. Click **Allocate** to add selected samples to the extraction worksheet. You can also allocate samples by scanning the barcode on the sample tube.

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11. To add the BIS to the worksheet, select the designated BIS well and scan the barcode label on the EP2 tube.
12. If more samples need to be added, click Save.
13. If no changes/additions are needed, click **Complete** in the **Batch Setup** window.
14. By completing the batch, a **Blank** sample will automatically be generated. If multiple RBs are used for the differential batch, each RB will have a unique barcode.
15. In the Lot field, you may distinguish RB1, RB2 with your initials/date – but it is not necessary as the barcodes are affixed to the tubes.
16. In the pop-up window, click Save.

II. Extraction (Lysis & QIAcube Processing)

1. Open **Processing** → **DNA Processing** → **Extraction**.
2. Select/scan the batch to be processed from the worklist and click **Select Scenario** (on the right side).

Batch Setup X Extraction X Storage Subsystem X Sample Setup X						
Batches Worklist : 3						
	Selected	Create Date	Bar Code	Batch Type	Kit	Custodi
	<input type="checkbox"/>	04/11/2025	BTE-250411-01	Bone and Tooth Extraction	Not Defined	In Lab
	<input type="checkbox"/>	05/01/2025	E2L-250501-01	EZ2 Lysis	Not Defined	Adrianni
	<input checked="" type="checkbox"/>	05/21/2025	Q04-250521-06	QIAcube Differential Lysis: 04 Samples	Not Defined	Cheryl C

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3. Differential consumables are split into 3 groups.

Consumables : 3							
Group Type	Group Name		Extra		Extra Number		
All	Sample Pretreatment		# Extra Reactions		1		
Optional	Fixed	Consumable Name 1	Format 1	Bar Code 1	Reagent Amount	Measurement	Kit
<input type="checkbox"/>	<input type="checkbox"/>	Buffer ATL	EI#####		1760.00	ul	
<input type="checkbox"/>	<input type="checkbox"/>	Buffer ATE	EH#####		3520.00	ul	
<input type="checkbox"/>	<input type="checkbox"/>	Qiagen Proteinase K	PK#####		220.00	ul	
<input type="checkbox"/>	<input type="checkbox"/>	EZ1 cRNA	RN#####		11.00	ul	
Group Type	Group Name		Extra		Extra Number		
Non-Sperm	Separation & Washes		No Extra				
Optional	Fixed	Consumable Name 1	Format 1	Bar Code 1	Reagent Amount	Measurement	Kit
<input type="checkbox"/>	<input type="checkbox"/>	EZ1 Buffer G2	EF#####				
Group Type	Group Name		Extra		Extra Number		
Sperm	Sperm Lysis Buffer		No Extra				
Optional	Fixed	Consumable Name 1	Format 1	Bar Code 1	Reagent Amount	Measurement	Kit
<input type="checkbox"/>	<input checked="" type="checkbox"/>	EZ1 Buffer G2	EF#####		623.00	ul	
<input type="checkbox"/>	<input checked="" type="checkbox"/>	Qiagen Proteinase K	PK#####		42.00	ul	
<input type="checkbox"/>	<input checked="" type="checkbox"/>	Prepared DTT	WW#####		166.00	ul	

4. Highlight the **Samples Pretreatment** header and scan the consumable barcodes in the **Sample Pretreatment** portion of the consumable window. This is for the incubation of your samples with the master mix.

Consumables : 3							
Group Type	Group Name		Extra		Extra Number		
All	Sample Pretreatment		# Extra Reactions		1		
Optional	Fixed	Consumable Name 1	Format 1	Bar Code 1	Reagent Amount	Measurement	Kit
<input type="checkbox"/>	<input type="checkbox"/>	Buffer ATL	EI#####	EI000008	1440.00	ul	
<input type="checkbox"/>	<input type="checkbox"/>	Buffer ATE	EH#####	EH000021	2880.00	ul	
<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	Qiagen Proteinase K	PK#####	PK000013	180.00	ul	
<input type="checkbox"/>	<input type="checkbox"/>	EZ1 cRNA	RN#####	RN000012	9.00	ul	
Group Type	Group Name		Extra		Extra Number		
Non-Sperm	Separation & Washes		No Extra				
Sperm	Sperm Lysis Buffer		No Extra				

5. Scan the batch and click **Start Process** on the bottom right-hand corner.

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6. Complete the tube check by scanning the tube labels and proceed with lysis. Click **OK**

Tube Check - Q04-250522-02 (TubeCheckForm)

		# of Checks: 1		Check Order: Horizontal	
	1	2	3	4	
A	CC022	BI000193	240018	240019	
B					

Performed By: OK Cancel

7. A window with a .jpg picture will populate depicting the QIAcube Loading Scheme based on the sample quantity chosen.
8. Labels are needed for subsequent QIAcube loading and can be printed from the **Print Labels** button on top right. Print 2 copies. The A frac labels will go on the tubes in the shaker and your EZ2 elution tubes. An additional copy of the B frac labels can be printed from the “content” button. Highlight just the samples you want to print and press “print selected bar code” at the bottom. The B frac labels will go on the rotor adaptor, the tube inside the rotor adaptor and your elution tubes for your EZ2.

Bar Code / Label Printing (BarcodeLabelPrintForm)

Batch Bar Code: Q04-250522-03

Batch Bar Code Printing

Copies:

Sample Bar Code Printing

Copies:

9. Once lysis is completed, highlight the batch and click **Load Scenario** on the right-hand side.

Consumables : 3							
Group Type	Group Name	Extra	Extra Number				
All	Sample Pretreatment	# Extra Reactions	1				
Non-Sperm	Separation & Washes	No Extra					
Optional	Fixed	Consumable Name 1	Format 1	Bar Code 1	Reagent Amount	Measurement	Kit
<input type="checkbox"/>	<input type="checkbox"/>	EZ1 Buffer G2	EF#####				

10. Highlight the **Separation and Washes** header and scan the Buffer G2 lot. Scan the QIAcube instrument barcode. Click **Continue** on right hand side.

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11. Complete the tube check by scanning the tube labels and proceed with instrument setup.
12. Highlight the batch and click **Load Scenario**.
13. Highlight the **Sperm Lysis Buffer** header. Scan the appropriate consumables to prepare the Sperm Lysis Buffer.

Group Type	Group Name	Extra	Extra Number
Sperm	Sperm Lysis Buffer	No Extra	

Optional	Fixed	Consumable Name 1	Format 1	Bar Code 1	Reagent Amount	Me
<input type="checkbox"/>	<input checked="" type="checkbox"/>	EZ1 Buffer G2	EF#####	EF000016	506.00	ul
<input type="checkbox"/>	<input checked="" type="checkbox"/>	Qiagen Proteinase K	PK#####	PK000013	34.00	ul
<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	Prepared DTT	WW#####	WW000020	135.00	ul

14. Click **Complete Process** (bottom right) and record the results using the Complete Batch Activity screen.
 - **Process Successful:** the batch advances to the next processing step.
 - **Process Aborted:** the batch remains on the Extraction Batches worklist. A **Batch Comment** is required with this option.
 - **Process Failed:** the batch is abandoned and all samples return to **Batch Setup**. A **Batch Comment** is required with this option. This option should only be used if the instrument crashes or something occurs that affects all samples.
15. Click **Save**.

III. Isolation

1. Create an Isolation (purification) worksheet: Open **Processing → DNA Processing → Isolation**. Click **Create Batch**.

Bar Code	Batch Type	Kit
Q04-250522-03	QIAcube Differential Lysis: 04 Samples	Not Defi

2. Click the **Create** button under the New Batches worklist.

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3. Select a **Batch Type** and click **Create** to create the unique barcode associated with isolation.

- a. **EZ1/EZ2 Purification** = “A” fractions + any non-differentials lysed separately.

The screenshot shows the 'Create Batch (CreateBatchForm)' window. At the top, there are four dropdown menus: 'Batch Type' (set to 'EZ1/2 Purification'), 'Kit' (set to 'Not Defined'), 'Fill Order' (set to 'Horizontal'), and 'Batch Bar Code' (empty). To the right of these is a 'Create' button. Below the dropdowns is a grid of 24 columns, numbered 1 to 24. Each column has a circular icon with the text 'DNA' inside. The first column is labeled 'A' on the left.

- b. **EZ2 Trace – Sperm** = “B” fractions

The screenshot shows the 'Create Batch (CreateBatchForm)' window. At the top, there are four dropdown menus: 'Batch Type' (set to 'EZ2 Trace - Sperm'), 'Kit' (set to 'Not Defined'), 'Fill Order' (set to 'Horizontal'), and 'Batch Bar Code' (set to 'E2S-250523-08'). To the right of these is a 'Create' button. Below the dropdowns is a grid of 24 columns, numbered 1 to 24. Each column has a circular icon with the text 'DNA' inside. The first column is labeled 'A' on the left.

4. Scan the newly created barcode(s) into each **Destination Batch** section (top and bottom on right hand side).
5. Scan the extraction barcode into the **Source Batch** section. If necessary, non-differential samples can be merged with the A fractions here.

STACS SOP-4 Differential

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Page 9 of 11

The screenshot displays the STACS SOP-4 Differential software interface. It features several panels:

- New Batches: 7**: A table listing various sample batches with columns for Bar Code, Batch Type, Kit, Custodian/Location, and Sample Type. Examples include EZ2-250523-06 (EZ2 Trace - Sperm) and EZ2-250523-07 (EZ1/2 Purification).
- Source Batch**: A dropdown menu showing 'Q04-250522-03'.
- Available Batches: 1**: A table showing available batches, including 'Q04-250522-03' (QAcube Differential Lysis; 04 Samples).
- Samples for Batch 'Q04-250522-03': 8**: A table listing samples with columns for Action, Well, Bar Code, Lab Case Number, and Exhibit Number. Samples include A01, B01, A02, B02, A03, B03, A04, and B04.
- Destination Batch**: A section for 'EZ2-250523-08 (Not Defined)' with a 'Complete' button and a grid for sample allocation (A, B, C, D) across four lanes (1, 2, 3, 4). Lane 1 is highlighted in red.
- Destination Batch 2**: A section for 'EZ2-250523-07 (Not Defined)' with a 'Complete' button and a similar allocation grid.

16. Click **Allocate** to add the samples to the isolation worksheet. “A” and “B” fractions will automatically separate on to the appropriate isolation batch. If sample(s) need to be moved, this can be done after the allocation is saved, followed by dragging the sample(s) as needed.
6. If more samples need to be added, click **Save**.
7. If finished, click **Complete** (next to Destination Batch Barcode) and then close.
8. In the Batches Worklist window, click **Select Scenario** for each Isolation worksheet.
 - a. EZ1/EZ2 Purification = EZ1/2 Large Volume Protocol
 - b. EZ2 Trace -Sperm
9. Scan the batch barcode and all necessary consumables and instrumentation.
10. Click **Start Process**.
11. Complete the tube check by scanning the lysis tube barcode labels.
12. Once the isolation is complete, click **Complete Process** and select the appropriate option.
 - a. **Process Successful:** the batch advances to the next processing step.
 - b. **Process Aborted:** the batch remains on the Isolation worklist. A **Batch Comment** is required with this option.

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- c. **Process Failed:** the batch is assigned a **Status** of '**Abandoned**' and its samples are returned to Isolation Batch Create to be allocated to a new batch. A **Batch Comment** is required with this option.

Complete Batch Activity (CompleteBatchActivityForm)

☒ Process Successful ☐ Process Routed ☐ Process Failed

Batches : 1

Bar Code	Process	Status	New Status	Batch Comment
EZZ-250523-07	Success	Isolation Batch Created	Isolation Batch Complete	

Samples : 1

Sample	Kit	Lab Case Number	Exhibit Number	Final Volume (uL)
Batch Bar Code : EZZ-250523-07 (4 items)				
CC023.A	Not Defined	CLC-250522-1417	13	30.0
B1000193.A	Not Defined			30.0
250086.A	Not Defined			30.0
250087.A	Not Defined			30.0

Required Batches : 2

Batch Type	Controllant's Location	Bar Code
Q/Acetic Differential Lysis: 04 Samples	Cheryl Carneiro	Q04-250522-03
EZZ/Q Purification	Cheryl Carneiro	EZZ-250523-07

13. If an elution volume discrepancy results in a manual volume check, this will be captured in the **Complete Process** window. Enter each sample's elution volume as measured off the instrument and select **Process Successful**. A note will be made (see red circle below) if water is added to an elution volume to bring up to RB volume, if necessary (the same water falcon tube is used throughout the entire process).

Batch Bar Code : EZZ-250523-07 (4 items)

CC023.A	Not Defined	CLC-250522-1417
B1000193.A	Not Defined	
250086.A	Not Defined	
250087.A	Not Defined	

Activity Comment

Batch Comment

14. Proceed to the **Batch Management** module to document subsequent adjustments. Scan the extraction batch or locate using the **Quantitation Batch Create** option in the **Processing Step** dropdown
- If an elution volume is significantly increased ($>40\mu\text{L}$), select the sample(s) and control(s) and proceed to the **Concentrate** tab.
 - If an elution volume is significantly reduced ($0-20\mu\text{L}$), select the sample(s) and control(s) and click on the **Rework** tab. Select **Isolation Batch Create** in the

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Rework Entry Point dropdown and document the **Rework Reason**. Click **Save**.

The sample(s)/control(s) can now be added to a new isolation batch.

- c. If an eluate needs a volume adjustment, this can be done through the **Dilute** tab.

Batch Management (BatchManagementTabForm)

Processing Step
Quantitation Batch Create

Batches : 4

Concentrate	Custodian/Location	Bar Code	Kit	Processing Method	Batch Type
<input type="checkbox"/>	In Lab	EZ2-250224-02	Not Defined	Manual	EZ1/2 Purification
<input type="checkbox"/>	In Lab	EZ2-250313-38	Not Defined	Manual	EZ1/2 Purification
<input type="checkbox"/>	Cheryl Carreiro	EZ2-250523-05	Not Defined	Manual	EZ1/2 Purification
<input type="checkbox"/>	Cheryl Carreiro	EZ2-250523-07	Not Defined	Manual	EZ1/2 Purification

Source Batch Bar Code Destination Batch Bar Code Sample/Control Bar Code

Add **Dilute** **Concentrate** **Discontinue** **Combine** **Purify** **Remove** **Rework** **Transfer**

Kit

Samples