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Centers for Disease Control and Prevention (CDC) National Institute for Occupational Safety and Health (NIOSH) 1095 Willowdale Road Morgantown, WV 26505-2888

March 12, 2008 HETA 2001-0445 Interim Letter IX

Mike Winkler, First Vice-President Administrative and Residual Employees Union Local 4200 705 North Mountain Road, Suite A211 Newington, Connecticut 06111-1411

Dear Mr. Winkler:

Included with this letter is a hard copy of a poster to be presented by NIOSH personnel at the Society of Toxicology meeting to be held in Seattle, Washington on March 16-20, 2008. This presentation will report inflammatory effects in mice of floor dusts collected from the 2002 survey at the 25 Sigourney Street Building and associations of levels of endotoxin and glucan in dust with the inflammatory markers. The animal study is still ongoing, and the presentation is based on preliminary results.

The findings included in the presentation are as follows:

- 1. The exposure to the building floor dust produced dose-dependent inflammatory responses in mouse lung.
- 2. Although the levels of endotoxin and glucan in floor dust do not appear to be associated well with the markers of lung inflammation (since the significant association of the 2002 analytical results of endotoxin in dust with the inflammatory markers was driven by a single outlier), the polymyxin B experiment (polymyxin B: antibiotics which deactivates endotoxin activity) suggests that endotoxin may be an important inflammatory substance in dust in this animal model.

If you have any questions regarding the information provided in this interim letter, please do not hesitate to contact us at 1-800-232-2114.

Sincerely,

Ju-Hyeong Park, ScD, MPH, CIH

Park, Tuhyeon

Environmental Health Scientist Respiratory Disease Hazard Evaluation

and Technical Assistance Program

Field Studies Branch

Division of Respiratory Disease Studies

### Use of a Mouse Model to Evaluate Pulmonary Inflammation Caused by Floor Dust from a Water-Damaged Building

### S-H. Young, J. M. Cox-Ganser, M. Wolfarth, J.M. Antonini, V. Castranova, J-H. Park

### National Institute for Occupational Safety and Health, Morgantown, WV

### Introduction

Although the causes of building-related respiratory illness are still unclear epidemiological research has indicated that fungi and endotoxin in floor dust are associated with such health risks. In the present study, we used a mouse model to evaluate pulmonary inflammation caused by floor dusts collected from the workstations of employees in a water-damaged office building. The dass were tested in an endotoxin-sensitive strain of mouse. C3HeBFel, and palmonery atflammation was determined. We examined the correlation among markers of atflammation and levels of endotoxin and (1+3)-β-D-glucan, a major cell wall component of fungi

### Aims

- To examine outmonary inflammatory responses to increasing doses of floor dust from a water
- damaged building. 2) To examine the correlations among markers of inflammation and levels of endotoxin and (1-+3)-B-D-slucan

### **Methods and Materials**

Dust sample: Dust samples were collected from carneted floors of workstations using ackpack vacuum samplers with erevice tools. For each sampling location, a 2 m2 floor area was vacuumed for S minutes. Each dust sample was analyzed for endotoxin and (1→3)-8-D-elucan

- Animals: Pathogen-free male endotoxin-sensitive strain of mouse C3HeB/FeJ (number of mice#4 to 6 per dust sample) was used.
- Exposure: Each mouse was treated with dust sample (1.1.5, 2.5 or 3.5 mg/kg of body weight suspended in 40 µL of saline) by pharyngeal aspiration: Control mice recei an equivalent volume of saline by aspiration.
- Polymyxin B treatment: Dust was suspended in polymyxin B (PMB) solution (Img/ml) and incubated at 37 °C for 30 min. PMB binds to and deactivates endotoxin Pulmonary Parameters:
- At 18 hrs post aspiration, bronchoalveolar lavage (BAL) was tione postmorten on lungs and the following inflammatory and lung injury markers were measured;
- 1. Lung inflammation Polymorphomoclear Isukoestes (PMN) infiltration. Differential sells counts from total BAL coils
- Lactate dehydrogenase (LDH) activity and albumin concentration from acellular BAL.
- 3. Cytokina production
- Cytokines production: IL-12p70, TNF-ix, IFN-y, MCP-1, IL-10, and IL-5 in first neelligian fraction of BAL fluid.

#### Disclaimer

The findings and conclusions in this report are those of the authors and do not necessarily represent the views of the National Institute for Occupational Safety and Health.

### Results

Dust-induced Pulmonary Inflammation- Dose-Response (table ), Pearson product moment conclutions between malyies in dast and markers of Decreased TNF-0/ After Polymyxin B Treatment

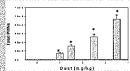
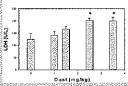


Figure 1. Total number of PMN after exposure to dust. One day post dust exposure PMN was harvested from BAL fluid. There is a dose-dependent increase of PMN counts: Values are means  $\pm S.E.*$  Significantly different than the mean value of

### Lung Cell Injury



uro 2, Lactate dehydrogenase (LDH) level in accillular HAL fluid after exposure to different doses of dust. There was a dose-dependent increase of LDH. Values are

#### Cytokine Responses

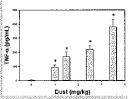


Figure 3, Pro-mflammatory cytokine TNP-to in the accilular BAL fluid of mice that were exposed to different doses of dust. Values are means ± SE. \* Significantly different than the mean value of saline control mice (p<0.05).

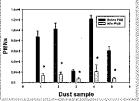
interpretation of the cell contents are (1 + 0)-rotation coefficient, p = P. Value, and in Number of Samples, Significant difference was set at p < 0.05 and was colored in sellow < 0.05 denotes that p-value in between 0.001 and 0.05.

		44	B44 cells	PMHs	TNF-4	IFP 2	H(P.)	R 5	Glucan	€#3007	E0:5005
	10.3	D627	6 477	0.538		9 723		3.45	-0.0978	9.3494	0.0967
LDH	10.	10001	<0.001	oc.	0.001	4005	42701	40.T1	0.136	5,097	9.2
	100	160	156	165	156	151	198	166	-22	172	15
W	14.5		0.508	0.057	0.4	9,25	3 683	0,549	-C.144	-9:00203	0.047
	E.		<0.001	-01UI	40(0)	40.06	<3(0)	40.01	0114	0,842	0/6
	177		155	156	198	156	164	168	122	122	159
	14.			0.047	0.573	0.126	0.48	0.542	4) 0007	0.0510	0.153
RAL Cells	e.			40.00L	40.E01	0.119	<1001	<0.001	U 277	USJ	<0.05
	10.		.,	155	155	155	155	155	177	172	155
	17	-			0539	D.192	158	5.842	5,130	0.3535	0,202
fális	120				0001	40(6	cirni	(C.X.)	8 129	0.73	100
	175	·····			155	155	155	155	123	127	154
TNF-	17					0 202	3619	C.757	-0.250	-0.3799	. 0 0300
	100					4006	<0.001	<0.00	4.35	E.352	0.64
	170					166	196	166	122	122	- 0
	77					-	0.304	0.427	0.0146	D.174	0 197
IFM a	10						<0.001	·C.X1	0.074	0.253	4.2
	10		1111111111		,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		150	160	122	122	150
	1.							6.860	-C.159	-0.0570	926
MCP-1	10.							4.11	0 '20	0.526	0.00
	10							166	727	122	150
	1.								-C.158	400600	0 105
11.6	180								40.05	0.347	-0.05
	10								122	122	100
	,									0.450	6.002
lucan	in.									<0.001	9,70
	100			******	,					122	122
	17										0.111
EU.	A.			******		1		1			0,225
7007	14.										122

Table 2. Descriptive statistic for endotoxin measured in 2007 (EU-2007), endotoxin measured in 2002 (EU-2002), and (1-3)- $\beta$ -D-glucan

Calera	Zie+	None	SIG. Zaw	Arms	Mex	Mis
etimet (Etimeti	в	50.220	3.944	267.290	223000	4210
SET STATES	ж	LEAST?	1.000	119.369	193100	4,0310
Discen (1921/14)	Te.	31.186	2.591	fia ten	137/260	4,790

### Polymyxin B Binds to Endotoxin and Decreases PMNs



dust to/without polymyxin B (PMB) freatment. The PMB-treated dust was suspended in PMB solution and incubated at 37 °C for 30 min. Values are means ± S.E. \* Significantly different that the mean value of without PMB treatment mice (p<0.05)

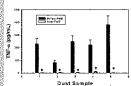


Figure 5, TNF-or production in BAL fluid of mice after exposure to 2.5 mg/kg bwt dust. sy/without polymyxin B (PMB) treatment. The PMB treated-dust was suspended in PMB solution and incubated at 37 °C for 30 min. Values are means ± S.E. \* Significantly different than the mean value of without PMB treatment mice (p<0.05)

#### Correlation Between EU2007 and EU2002

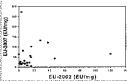


Figure 6, Seafter plot of EU/2007 vs EU/2002. These results were obtained by using the state LAL midhod but from 2 different labs. If we remove the two outliers from the graph, then EU/2007, was significantly correlated with EU/2002.

### Summary and Conclusions

- Endotoxin in the floor dust was associated with markers of inflamination in these experiments Endotoxin 2002 was significantly correlated with PMN, total BAL cells, IFN-y, MCP-1, and IL-However, even at a very low endotoxin level inflammation still occurred. This suggested that
  another component in the dust also contributed to the overall inflammatory potency.
- The animal experiment was done in 2007-8. The collected dust samples were stored for 6 years. in -80°C. EU-2002 data was 6 years old which may not reflect the setual endotoxin levels at the time of experiment. The correlations of EU-2002 with the markets were driven by an outlier. EU-2007 is a current endoloxia measurement, but was not correlated with markers of
- Polymyxin B experiments indicated that endotoxin was a contributing factor in pulmonary inflammation in mice exposed to dust from a water-damaged building.
- B-Glucan measurement was not correlated with markers of inflammation. Possible reasons may nelude: I. S. giucan is a weaker inflammatory agent than endotoxin, therefore some endotoxin contamination can mask the effect of S. giucan, 2. The current available S. giucan measurement method is for detecting water soluble β-glucen, which was much less inflammatory than insoluble 8. edican

## Introduction

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## Aims

- 1) To examine pulmonary inflammatory responses to increasing doses of floor dust from a water-damaged building.
- 2) To examine the correlations among markers of inflammation and levels of endotoxin and  $(1\rightarrow 3)$ - $\beta$ -D-glucan.

## Methods and Materials

Dust sample: Dust samples were collected from carpeted floors of workstations using backpack vacuum samplers with crevice tools. For each sampling location, a 2 m2 floor area was vacuumed for 5 minutes. Each dust sample was analyzed for endotoxin and  $(1\rightarrow 3)$ - $\beta$ -D-glucan.

Animals: Pathogen-free male endotoxin-sensitive strain of mouse - C3HeB/FeJ (number of mice=4 to 6 per dust sample) was used.

Exposure: Each mouse was treated with dust sample (1, 1.5, 2.5 or 3.5 mg/kg of body weight suspended in 40 µL of saline) by pharyngeal aspiration. Control mice received an equivalent volume of saline by aspiration.

Polymyxin B treatment: Dust was suspended in polymyxin B (PMB) solution (1mg/ml) and incubated at 37 °C for 30 min. PMB binds to and deactivates endotoxin.

### **Pulmonary Parameters:**

At 18 hrs post aspiration, bronchoalveolar lavage (BAL) was done postmortem on lungs and the following inflammatory and lung injury markers were measured:

- 1. Lung inflammation
  - Polymorphonuclear leukocytes (PMN) infiltration: Differential cells counts from total BAL cells.
- 2. Lung damage
  - Lactate dehydrogenase (LDH) activity and albumin concentration from acellular BAL fluid.
- 3. Cytokine production
  - Cytokines production: IL-12p70, TNF-a, IFN-g, MCP-1, IL-10, and IL-6 in first acellular fraction of BAL fluid.

## Results

## **Dust-induced Pulmonary Inflammation- Dose-Response**

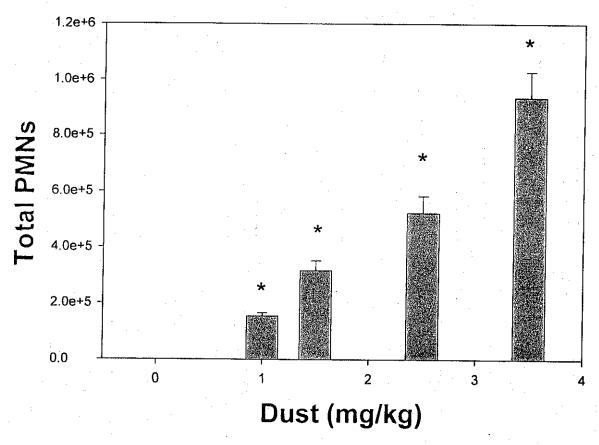


Figure 1, Total number of PMN after exposure to dust. One day post dust exposure, PMN was harvested from BAL fluid. There is a dose-dependent increase of PMN counts. Values are means  $\pm$  S.E. \* Significantly different than the mean value of saline control mice (p<0.05).

## **Lung Cell Injury**

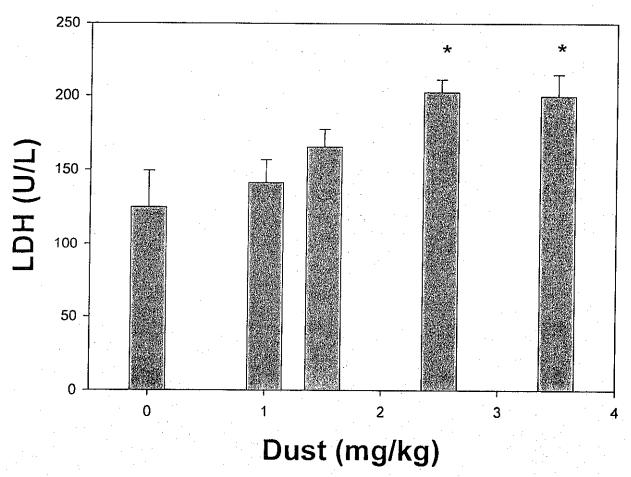


Figure 2, Lactate dehydrogenase (LDH) level in acellular BAL fluid after exposure to different doses of dust. There was a dose-dependent increase of LDH. Values are means  $\pm$  S.E. \* Significantly different than the mean value of saline control mice (p<0.05).

### **Cytokine Responses**

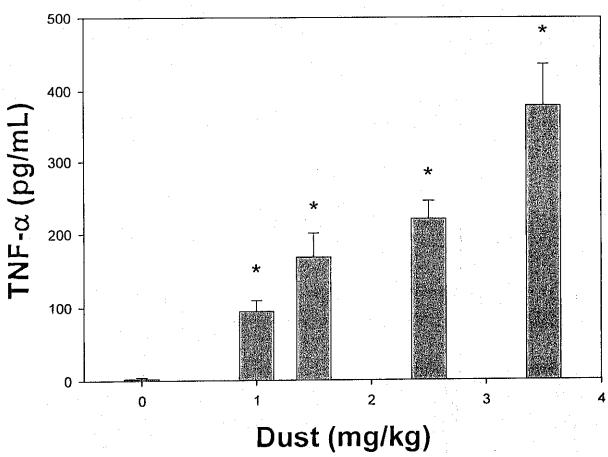


Figure 3, Pro-inflammatory cytokine TNF-a in the acellular BAL fluid of mice that were exposed to different doses of dust. Values are means  $\pm$  SE. \* Significantly different than the mean value of saline control mice (p<0.05).

Table 1, Pearson product moment correlations between analytes in dust and markers of inflammation. The cell contents are : r = Correlation Coefficient, p = P Value, and n = Number of Samples. Significant difference was set at <math>p < 0.05 and was colored in yellow. < 0.05 denotes that p-value is between 0.001 and 0.05.

		Ab	BAL cells	PMNs	TNF-a	IFN-y	MCP-1	IL-6	Glucan	EU-2007	EU-2002
	r	0.637	0.477	0.538	0.384	0.223	0.42	0.45	-0.0878	0.0484	0.0967
LDH	p	<0.001	<0.001	<0.001	<0.001	<0.05	<0.001	<0.001	0.336	0.597	0.23
	n	156	155	155	156	156	156	156	122	122	156
Δb	r		0.396	0.487	0.4	0.25	0.688	0.549	-0.144	0.00209	0.0473
	$P_{-1}$		<0.001	<0.001	<0.001	<0.05	<0.001	<0.001	0.114	0.982	0.558
	tt		155	155	156	156	156	156	122	122	156
BAL	r			0.947	0.573	0.126	0.48	0.542	-0.0807	0.0519	0.162
cells	$ p_{\perp} $			<0.001	<0.001	0.119	<0.001	<0.001	0.377	0.57	<0.05
	n.			155	155	155	155	155	122	122	155
	r				0.639	0.182	0.532	0.642	-0.138	0,0605	0.207
PMNs	$ p_{\perp} $				<0.001	<0.05	<0.001	_<0.001	0.129	0.508	<0.05
	n				155	155	155	155	-122	122	155
TNF-02	r					0.202	0.519	0.757	-0.236	-0.0799	-0.0368
	$p_{\perp}$					<0.05	<0.001	<0.001	<0.05	0.382	0.648
	13					156	156	156	122	122	156
	r						0.304	0.427	-0.0145	0.174	0.197
IFN-y	$ p_{\perp} $						<0.001	<0.001	0.874	0.0553	<0.05
	n						156	156	122	122	156
	ŗ							0.699	-0,138	-0.0579	0.216
MCP-1	$p_{\perp}$							<0.001	0.129	0.526	<0.05
·	Ĥ							156	122	122	156
	r								-0.198	-0.00609	0.198
IL-6	p_								<0.05	0.947	<0.05
	n								122	122	156
	r									0.498	0.092
Glucan	p									<0.001	0.314
	n								-	122	122
	r		_								0.111
EU.	$ \vec{p} $										0.222
2007	n										122

Table 2, Descriptive statistic for endotoxin measured in 2007 (EU-2007), endotoxin measured in 2002 (EU-2002), and  $(1\rightarrow 3)$ - $\beta$ -D-glucan.

Column	Size	Mean	Std. Error	Range	Max	Min
EU-2007 (EU/mg)	25	56.260	5.544	267.790	272.000	4.210
EU-2002 (EU/mg)	36	16.807	2.003	119.969	120.000	0.0310
Glucan (ng/mg)	25	51.186	3.015	118.980	123.760	4.780

### Polymyxin B Binds to Endotoxin and Decreases PMNs

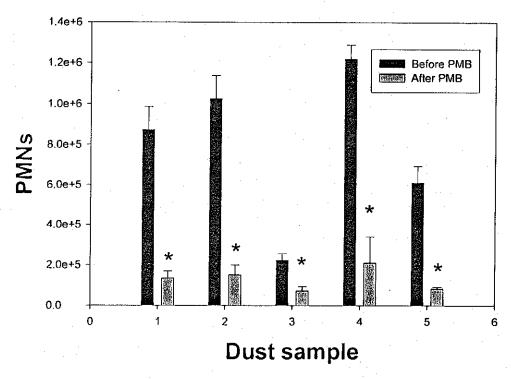


Figure 4, Total number of PMNs in BAL fluid of mice after exposure to 2.5 mg/kg bwt dust w/without polymyxin B (PMB) treatment. The PMB-treated dust was suspended in PMB solution and incubated at 37 °C for 30 min. Values are means  $\pm$  S.E.

<sup>\*</sup> Significantly different than the mean value of without PMB treatment mice (p<0.05).

### Decreased TNF-a After Polymyxin B Treatment

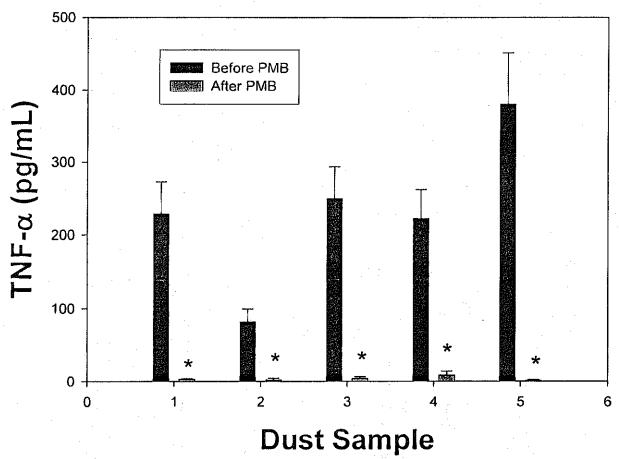


Figure 5, TNF-a production in BAL fluid of mice after exposure to 2.5 mg/kg bwt dust w/without polymyxin B (PMB) treatment. The PMB treated-dust was suspended in PMB solution and incubated at 37 °C for 30 min. Values are means  $\pm$  S.E. \* Significantly different than the mean value of without PMB treatment mice (p<0.05).

## Correlation Between EU2007 and EU2002

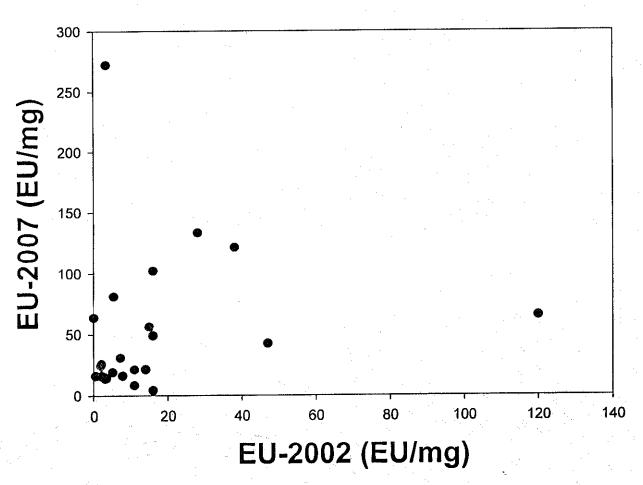


Figure 6, Scatter plot of EU2007 vs EU2002. These results were obtained by using the same LAL method but from 2 different labs. If we remove the two outliers from the graph, then EU2007 was significantly correlated with EU2002.

# Summary and Conclusions

- Endotoxin in the floor dust was associated with markers of inflammation in these experiments. Endotoxin 2002 was significantly correlated with PMN, total BAL cells, IFN-g, MCP-1, and IL-6. However, even at a very low endotoxin level inflammation still occurred. This suggested that another component in the dust also contributed to the overall inflammatory potency.
- The animal experiment was done in 2007-8. The collected dust samples were stored for 6 years in -80°C. EU-2002 data was 6 years old which may not reflect the actual endotoxin levels at the time of experiment. The correlations of EU-2002 with the markers were driven by an outlier. EU-2007 is a current endotoxin measurement, but was not correlated with markers of inflammation.
- \*Polymyxin B experiments indicated that endotoxin was a contributing factor in pulmonary inflammation in mice exposed to dust from a water-damaged building.
- $\dot{\beta}$ -Glucan measurement was not correlated with markers of inflammation. Possible reasons may include: 1. β-glucan is a weaker inflammatory agent than endotoxin, therefore some endotoxin contamination can mask the effect of β-glucan; 2. The current available β-glucan measurement method is for detecting water soluble β-glucan, which was much less inflammatory than insoluble β-glucan.