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Reportable Diseases, Emergency Illnesses and Health Conditions, and Reportable Laboratory Findings Changes for 2019

As required by Conn. Gen. Stat. §19a-2a and Conn. Agencies Regs. §19a-36-A2, the Commissioner of the Department of Public Health (DPH) is required to declare an annual list of Reportable Diseases, Emergency Illnesses and Health Conditions, and Reportable Laboratory Findings. The list of Reportable Diseases, Emergency Illnesses and Health Conditions has two parts: (A) reportable diseases; and (B) reportable emergency illnesses and conditions. An advisory committee, consisting of public health officials, clinicians, and laboratorians, contribute to the process. There is 1 modification to the healthcare provider list only; 2 additions and 5 modifications to the laboratory list only; and 2 additions to both the physician and laboratory lists. No changes have been made to emergency illnesses or health conditions.

Reportable disease forms can be found on the DPH "Forms" webpage at: https://portal.ct.gov/DPH/Communications/Forms/Forms.

Changes to the List of Reportable Diseases, Emergency Illnesses and Health Conditions

Part A: Reportable Diseases

HIV—acute cases

Reporting of acute HIV cases has been <u>modified</u>. Acute HIV cases will be reported as Category 1 diseases, which require a telephone call to the DPH immediately on the day of recognition or strong suspicion.

Changes to the List of Reportable Laboratory Findings

Candida spp, blood isolates only

Laboratory reporting of *Candida* species from blood specimens only has been <u>added</u>. Laboratories should submit all *Candida* spp. blood isolates to the State Public Health Laboratory for speciation and antifungal susceptibility testing. Reporting for *Candida auris* remains unchanged; *Candida auris*

In this issue	Page #
Reportable Diseases, Emergency Illnesses and Health Conditions, and Reportable Laboratory Findings - Changes for 2019	1
List of Reportable Diseases, Emergency Illnesses and Health Conditions - 2019	2
List of Reportable Laboratory Findings - 2019	3
Persons Required to Report Reportable Diseases, Emergency Illnesses and Health Conditions, and Reportable Laboratory Findings	4

from all sites and any potential *Candida auris* misidentifications should still be reported.

Enterotoxigenic Escherichia coli (ETEC)

Laboratory reporting of Enterotoxigenic *E. coli* (ETEC) has been <u>added</u>. This addition will allow DPH to estimate the number of ETEC positive tests typically identified by multiplex PCR GI panels that include ETEC and to conduct follow-up activities.

Group B Streptococcus (GBS)

Laboratory reporting of Group B *Streptococcus* has been <u>modified</u>. Laboratories should submit infant (<1 year of age) GBS isolates to the State Public Health Laboratory.

Hepatitis A

Laboratory reporting of Hepatitis A (HAV) has been modified. Laboratories should report (when available), nucleic acid/RNA test results and total bilirubin level conducted within 7 days of a positive test. These changes will align HAV surveillance with the national HAV case definition.

Hepatitis C

Laboratory reporting of Hepatitis C virus (HCV) has been <u>modified</u>. All labs are required to report HCV genotype results, either by paper or electronically. Laboratories are encouraged to develop policies consistent with CDC guidance for reflex HCV RNA testing following an initial reactive HCV antibody test.

Influenza

Laboratory reporting of influenza has been <u>modified</u>. Laboratories should report positive influenza results to the DPH only. Dual reporting to local health

(Continued on page 4)

REPORTABLE DISEASES, EMERGENCY ILLNESSES and HEALTH CONDITIONS - 2019 PART A: REPORTABLE DISEASES

Physicians, and other professionals are required to report using the Reportable Disease Confidential Case Report form (PD-23), other disease specific form or authorized method (see page 4 for additional information). Forms can be found on the DPH "Forms" webpage or by calling 860-509-7994. Mailed reports must be sent in envelopes marked "CONFIDENTIAL." Changes for 2019 are in bold font

Category 1 Diseases: Report immediately by telephone (860-509-7994) on the day of recognition or strong suspicion of disease

for those diseases marked with a telephone (28). On evenings, weekends, and holidays call 860-509-8000.

These diseases must also be reported by mail within 12 hours.

Category 2 Diseases: All other diseases not marked with a telephone must be reported by mail within 12 hours of

recognition or strong suspicion of disease.

Acquired Immunodeficiency Syndrome (1,2) Acute flaccid myelitis

Acute HIV infection

Anthrax

Bahesiosis

Borrelia miyamotoi disease

Botulism

Brucellosis

California group arbovirus infection

Campylobacteriosis

Candida auris

Carbon monoxide poisoning (3)

Chancroid Chickenpox

Chickenpox-related death

Chikungunya

Chlamydia (C. trachomatis) (all sites)

Cholera

Cryptosporidiosis Cyclosporiasis Dengue

Diphtheria

Eastern equine encephalitis virus infection

Ehrlichia chaffeensis infection

Escherichia coli O157:H7 gastroenteritis

Gonorrhea

Group A Streptococcal disease, invasive (4) Group B Streptococcal disease, invasive (4) Haemophilus influenzae disease, invasive (4)

Hansen's disease (Leprosy)

Healthcare-associated Infections (5)

Hemolytic-uremic syndrome (6)

Hepatitis A Hepatitis B:

acute infection (2)

HBsAg positive pregnant women

Hepatitis C:

acute infection (2)

positive rapid antibody test result

HIV-1 / HIV-2 infection in: (1)

persons with active tuberculosis disease

 persons with a latent tuberculous infection (history or tuberculin skin test ≥5mm induration by Mantoux technique)

persons of any age

pregnant women

HPV: biopsy proven CIN 2, CIN 3 or AIS or their equivalent (1)

Influenza-associated death (7)

Influenza-associated hospitalization (7)

Legionellosis

Listeriosis

Lyme disease

Malaria

Measles

Melioidosis

Meningococcal disease

Mercury poisoning

Mumps

Neonatal bacterial sepsis (8)

Neonatal herpes (≤ 60 days of age)

Occupational asthma

Outbreaks:

Foodborne (involving ≥ 2 persons)

Institutional

Unusual disease or illness (9)

Pertussis

Plague

Pneumococcal disease, invasive (4)

Poliomvelitis

Powassan virus infection

Q fever

Rabies

Ricin poisoning

Rocky Mountain spotted fever Rubella (including congenital)

Salmonellosis

SARS-CoV

Shiga toxin-related disease (gastroenteritis) Shigellosis

Silicosis

Smallpox

St. Louis encephalitis virus infection

Staphylococcal enterotoxin B pulmonary

poisoning

Staphylococcus aureus disease, reduced or resistant susceptibility to vancomycin (1) Staphylococcus aureus methicillin-

resistant disease, invasive, community acquired (4,10)

Staphylococcus epidermidis disease, reduced or resistant susceptibility

to vancomycin (1)

Syphilis

Tetanus

Trichinosis

Tuberculosis

Tularemia Typhoid fever

Vaccinia disease

Venezuelan equine encephalitis virus infection Vibrio infection (parahaemolyticus, vulnificus, other)

West Nile virus infection

Yellow fever Zika virus infection

FOOTNOTES:

Report only to State.

As described in the CDC case definition.

Includes persons being treated in hyperbaric chambers for suspected CO

Invasive disease: from sterile fluid (blood, CSF, pericardial, pleural, peritoneal, joint, or vitreous) bone, internal body sites, or other normally sterile site including muscle.

Report HAIs according to current CMS pay-for-reporting or pay-forperformance requirements. Detailed instructions on the types of HAIs, facility types and locations, and methods of reporting are available on the DPH website: https://portal.ct.gov/DPH/Infectious-Diseases/HAI/Healthcare-Associated-Infections-and-Antimicrobial-Resistance

- 6. On request from the DPH and if adequate serum is available, send serum from patients with HUS to the DPH Laboratory for antibody testing.
- Reporting requirements are satisfied by submitting the Hospitalized and Fatal Cases of Influenza-Case Report Form in a manner specified by the DPH.
- Clinical sepsis and blood or CSF isolate obtained from an infant ≤ 72 hours of age. Individual cases of "significant unusual illness" are also reportable.

10. Community-acquired: infection present on admission to hospital, and person has no previous hospitalizations or regular contact with the health-care setting.

How to report: The PD-23 is the general disease reporting form and should be used if other specialized forms are not available. The PD-23 can be found on the DPH "Forms" webpage (https://portal.ct.gov/DPH/Communications/Forms/Forms). It can also be ordered by writing the Department of Public Health, 410 Capitol Ave., MS#11FDS, P.O. Box 340308, Hartford, CT 06134-0308 or by calling the Epidemiology and Emerging Infections Program (860-509-7994). Specialized reporting forms are available on the DPH "Forms" webpage or by calling the following programs: Epidemiology and Emerging Infections Program (860-509-7994) - Hospitalized and Fatal Cases of Influenza, Healthcare Associated Infections (860-509-7995) - National Healthcare Safety Network, HIV/AIDS Surveillance (860-509-7900) - Adult HIV Confidential Case Report form, Immunizations Program (860-509-7929) - Chickenpox Case Report (Varicella) form, Occupational Health Surveillance Program (860-509-7740) -Physician's Report of Occupational Disease, Sexually Transmitted Disease Program (860-509-7920), and Tuberculosis Control Program (860-509-7722). National notifiable disease case definitions are found on the CDC website.

Telephone reports of Category 1 disease should be made to the local Director of Health for the town in which the patient resides, and to the Epidemiology and Emerging Infections Program (860-509-7994). Tuberculosis cases should be directly reported to the Tuberculosis Control Program (860-509-7722). For the name, address, or telephone number of the local Director of Health for a specific town contact the Office of Local Health Administration (860-509-7660).

For public health emergencies on evenings, weekends, and holidays call 860-509-8000.

REPORTABLE LABORATORY FINDINGS—2019

The director of a clinical laboratory must report laboratory evidence suggestive of reportable diseases (see page 4 for additional information. The Laboratory Report of Significant Findings form (OL-15C) can be found on the DPH "Forms" webpage or by calling 860-509-7994. Changes for 2019 are in **bold font**.

Anaplasma phagocytophilum by PCR only	Legionella spp
Babesia: ☐ IFA IgM (titer) IgG (titer)	☐ Culture ☐ DFA ☐ Ag positive
□ Blood smear □ PCR □ Other	☐ Four-fold serologic change (titers)
□ microti □ divergens □ duncani □ Unspeciated	Listeria monocytogenes (1) □ Culture □ PCR
Bordetella pertussis (titer) Culture (1) ☐ Non-pertussis Bordetella (1) (specify)	Mercury poisoning
☐ Culture (1) ☐ Non-pertussis Bordetella (1) (specify)	□ Urine ≥ 35 μg/g creatinine μg/g
Borrelia burgdorferi (2)	□ Blood ≥ 15 μg/L μg/L Mumps virus (13) (titer) □ PCR
Borrelia miyamotoi	Mycobacterium leprae
California group virus (3) spp	Mysophostorium tuboroulosis Polated Testing (1)
Campylobacter (3) spp	AFB Smear
Candida auris (1,4)	If positive ☐ Rare ☐ Few ☐ Numerous
Candida spp. [blood isolates only]:(1,3)	NAAT □ Positive □ Negative □ Indeterminate
Carbapenem-resistant Acinetobacter baumannii (CRAB) (1,5)	Culture Mycobacterium tuberculosis
Carbapenem-resistant Enterobacteriaceae (CRE) (1,3,5)	☐ Non-TB mycobacterium. (specify <i>M</i>)
Genus spp Carboxyhemoglobin ≥ 5%% COHb	Neisseria gonorrhoeae (test type)
Carboxyhemoglobin ≥ 5%% COHb	Neisseria meningitidis, invasive (1,5)
Chikungunya virus	Culture Other
Chlamydia trachomatis (test type) Clostridium difficile (6)	Neonatal bacterial sepsis (3,14) spp
	Plasmodium (1,3) spp Poliovirus
Corynebacterium diphtheria (1) Cryptosporidium spp (3)	Powassan virus
□ Microscopy □ Other:	Rabies virus
Cyclospora spp (3) □ PCR □ Microscopy □ Other:	Rickettsia rickettsii
Dengue virus	
Eastern equine encephalitis virus	Rubella virus (13) (titer) Rubeola virus (Measles) (13) (titer) PCR
Ehrlichia chaffeensis by PCR only	St. Louis encephalitis virus
Enterotoxigenic Escherichia coli (ETEC) ☐ Culture ☐ PCR	Salmonella (1,3)(serogroup & type) ☐ Culture ☐ PCR
Escherichia coli O157(1) ☐ Culture ☐ PCR	SARS-CoV (1) ☐ IgM/IgG
Giardia spp (3)	□ PCR (specimen) □ Other
Group A <i>Streptococcus</i> , invasive (1,5) ☐ Culture ☐ Other	
Group B Streptococcus, invasive (1,5) ☐ Culture ☐ Other	□ PCR □ EIA
Haemophilus ducreyi	Shigella (1,3) (serogroup/spp) ☐ Culture ☐ PCR
Haemophilus influenzae, invasive (1,5) ☐ Culture ☐ Other	Staphylococcus aureus, invasive (5) ☐ Culture ☐ Other
Hepatitis A virus (HAV): ☐ IgM anti-HAV (7) ☐ NAAT Positive (7)	☐ methicillin-resistant ☐ methicillin-sensitive
ALT Total Bilirubin	Staphylococcus aureus, vancomycin MIC ≥ 4 μg/mL (1)
Hepatitis B HBsAg ☐ Positive ☐ Negative (8)	MIC to vancomycin μg/mL
☐ IgM anti-HBc ☐ HBeAg (2) ☐ HBV DNA (2)	Staphylococcus epidermidis, vancomycin MIC ≥ 32 μg/mL (1)
anti-HBs (8) ☐ Positive (titer) ☐ Negative	MIC to vancomycin µg/mL
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody	MIC to vancomycin µg/mL Streptococcus pneumoniae
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ Genotype specify ☐	MIC to vancomycin µg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5)
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ ☐ Genotype specify ☐ Herpes simplex virus (infants ≤ 60 days of age)	MIC to vancomycin µg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5) Treponema pallidum □ RPR (titer) □ FTA □ EIA
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ ☐ Genotype specify ☐ Herpes simplex virus (infants ≤ 60 days of age) ☐ Culture ☐ PCR ☐ IFA ☐ Ag detection	MIC to vancomycin µg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5) Treponema pallidum □ RPR (titer) □ FTA □ EIA □ VDRL (titer) □ TPPA
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ ☐ Genotype specify ☐ Herpes simplex virus (infants ≤ 60 days of age)	MIC to vancomycin µg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5) Treponema pallidum □ RPR (titer) □ FTA □ EIA □ VDRL (titer) □ TPPA Trichinella
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ Genotype specify ☐ Herpes simplex virus (infants ≤ 60 days of age) ☐ Culture ☐ PCR ☐ IFA ☐ Ag detection HIV Related Testing (report only to the State) (10) ☐ Detectable Screen (IA)	MIC to vancomycin µg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5) Treponema pallidum □ RPR (titer) □ FTA □ EIA □ VDRL (titer) □ TPPA Trichinella Varicella-zoster virus, acute
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ ☐ Genotype specify ☐ Herpes simplex virus (infants ≤ 60 days of age) ☐ Culture ☐ PCR ☐ IFA ☐ Ag detection HIV Related Testing (report only to the State) (10)	MIC to vancomycin µg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5) Treponema pallidum □ RPR (titer) □ FTA □ EIA □ VDRL (titer) □ TPPA Trichinella Varicella-zoster virus, acute
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anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) copies/mL	MIC to vancomycin µg/mL Streptococcus pneumoniae Culture (1,5)
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anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; % (10)	MIC to vancomycinµg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5) Treponema pallidum □ RPR (titer) □ □ FTA □ EIA □ VDRL (titer) □ □ TPPA Trichinella Varicella-zoster virus, acute □ Culture □ PCR □ DFA □ Other Vibrio (1,3) spp □ Culture □ PCR West Nile virus Yellow fever virus Yersinia, not pestis (1,3) spp □ Culture □ PCR Zika virus
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anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) ☐ Culture ☐ PCR ☐ IFA ☐ Ag detection HIV Related Testing (report only to the State) (10) ☐ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 ☐ Positive ☐ Neg/Ind ☐ HIV 2 ☐ Positive ☐ Neg/Ind ☐ HIV NAAT (or qualitative RNA) ☐ Detectable ☐ Not Detectable ☐ HIV Viral Load (all results) (10) ☐ copies/mL ☐ HIV genotype (10) ☐ CD4 count: cells/uL;% (10) HPV (report only to the State) (11) Biopsy proven ☐ CIN 2 ☐ CIN 3 ☐ AIS or their equivalent, (specify)	MIC to vancomycinµg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5) Treponema pallidum □ RPR (titer) □ FTA □ EIA □ VDRL (titer) □ TPPA Trichinella Varicella-zoster virus, acute □ Culture □ PCR □ DFA □ Other Vibrio (1,3) spp □ Culture □ PCR West Nile virus Yellow fever virus Yersinia, not pestis (1,3) spp □ Culture □ PCR Zika virus BIOTERRORISM possible disease indicators (15) Bacillus anthracis (1) Brucella spp (1) Burkholderia mallei (1)
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anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown	MIC to vancomycinµg/mL Streptococcus pneumoniae Culture (1,5)
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype □	MIC to vancomycinµg/mL Streptococcus pneumoniae Culture (1,5)
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12)	MIC to vancomycinµg/mL Streptococcus pneumoniae Culture (1,5)
anti-HBs (8) ☐ Positive (titer) ☐ Negative Hepatitis C virus (HCV) (9) ☐ Antibody ☐ PCR/NAAT/RNA ☐ ☐ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) ☐ Culture ☐ PCR ☐ IFA ☐ Ag detection HIV Related Testing (report only to the State) (10) ☐ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 ☐ Positive ☐ Neg/Ind ☐ HIV 2 ☐ Positive ☐ Neg/Ind ☐ HIV NAAT (or qualitative RNA) ☐ Detectable ☐ Not Detectable ☐ HIV Viral Load (all results) (10) ☐ copies/mL ☐ HIV genotype (10) ☐ CD4 count: ☐ cells/uL; ☐ (10) HPV (report only to the State) (11) Biopsy proven ☐ CIN 2 ☐ CIN 3 ☐ AIS or their equivalent, (specify) Influenza virus: (report only to State) ☐ Rapid antigen (2) ☐ RT-PCR ☐ Type A ☐ Type B ☐ Type Unknown ☐ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) ☐ Finger stick level ☐ µg/dL ☐ Venous level ☐ µg/dL	MIC to vancomycinµg/mL Streptococcus pneumoniae Culture (1,5)
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL;% (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level µg/dL □ Venous level µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send 6. Upon request from	MIC to vancomycinµg/mL Streptococcus pneumoniae Culture (1,5)
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send 6. Upon request from isolate for cases <1 year of age. For Salmonella,	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, positive stool sam, Shigella, Vibrio, and Yersinia (not pestis) tested by non- 7. Report peak ALT a	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen. 8. Negative HBsAg a	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available. For Shiga toxin-related disease, send positive broth or stool specimen. 8. Negative HBsAg a reportable for child.	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen. 2. Only laboratories with electronic file reporting are required to report positive results.	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen. 3. Specify species/serogroup/serotype. □ RT-PCR □ Type B □ Type Unknown positive stool sam solate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen. 2. Only laboratories with electronic file reporting are required to report positive For Chicken.	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ Genotype specify □ Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype □ Subtype □ Also □ Type B □ Type Unknown □ Subtype □ Subtype □ Venous level □ μg/dL □ Venous level □ μg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen. 8. Negative HBsAg a reportable for child to report positive results. 8. Specify species/serogroup/serotype. 8. Negative ABSAg a reportable for child to report positive results. 9. Report positive A Genotype results reportable by ele test result reports.	MIC to vancomycinµg/mL Streptococcus pneumoniae
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen. 2. Only laboratories with electronic file reporting are required to report positive results. 3. Specify species/serogroup/serotype. 4. Include samples from all sites. 5. Sterile site: sterile fluids (blood, CSF, pericardial, pleural, peritoneal, joint, or vitreous), bone, internal body site	MIC to vancomycinµg/mL Streptococcus pneumoniae
Anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) □ Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 μg/dL <48 hrs; 0-9 μg/dL monthly) (12) □ Finger stick level □ μg/dL □ Venous level □ μg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shígella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen. 2. Only laboratories with electronic file reporting are required to report positive results. 3. Specify species/serogroup/serotype. 4. Include samples from all sites. 5. Sterile site: sterile fluids (blood, CSF, pericardial, pleural, peritoneal, joint, or vitreous), bone, internal body site (lymph node, brain, heart, liver, spleen, kidney, pancreas, 10. Report all HIV anti	MIC to vancomycinµg/mL Streptococcus pneumoniae □ Culture (1,5) □ Urine antigen □ Other (5)
anti-HBs (8) □ Positive (titer) □ Negative Hepatitis C virus (HCV) (9) □ Antibody □ PCR/NAAT/RNA □ □ Genotype specify Herpes simplex virus (infants ≤ 60 days of age) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (10) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (10) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (10) □ copies/mL □ HIV genotype (10) □ CD4 count: □ cells/uL; □ % (10) HPV (report only to the State) (11) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent, (specify) Influenza virus: (report only to State) □ Rapid antigen (2) □ RT-PCR □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (12) □ Finger stick level □ µg/dL □ Venous level □ µg/dL 1. Send isolate/specimen to DPH Laboratory. For GBS, send isolate for cases <1 year of age. For Salmonella, Shigella, Vibrio, and Yersinia (not pestis) tested by non-culture methods, send isolate if available; send stool specimen if no isolate available. For Shiga toxin-related disease, send positive broth or stool specimen 2. Only laboratories with electronic file reporting are required to report positive results. 3. Specify species/serogroup/serotype. 4. Include samples from all sites. 5. Sterile site: sterile fluids (blood, CSF, pericardial, pleural, peritoneal, joint, or vitreous), bone, internal body site (lymph node, brain, heart, liver, spleen, kidney, pancreas, or ovary), or other normally sterile site including muscle. 10. Report pasitive NAAT report for specific form specific fo	MIC to vancomycinµg/mL Streptococcus pneumoniae

departments will be facilitated through a shared database (CTEDSS) maintained by DPH.

Yersinia (not pestis)

Laboratory reporting of *Yersinia* (not pestis) has been <u>modified</u> to require submission of *Yersinia* (not pestis) isolates, or stool specimens if no isolate is recovered for positive culture-independent testing results, to the State Public Health Laboratory for confirmation/isolation of the organism.

Changes to Both Lists

Borrelia miyamotoi

Reporting of *Borrelia miyamotoi* has been <u>added</u>. *B. miyamotoi* is an emerging tick-borne pathogen,

which has been identified in *Ixodes scapularis* ticks in Connecticut. State surveillance will assist in characterizing the incidence, epidemiology, and clinical spectrum of *B. miyamotoi* disease in Connecticut.

Powassan virus

Reporting of Powassan virus has been <u>added</u>. Powassan virus is a tick-borne arbovirus, which has been identified in *Ixodes* species ticks and in humans in Connecticut. State surveillance will contribute to national surveillance to better understand the epidemiology of the infection.

Persons Required to Report Reportable Diseases, Emergency Illnesses and Health Conditions

- 1. Every health care provider who treats or examines any person who has or is suspected to have a reportable disease, emergency illness or health condition shall report the case to the local director of health or other health authority within whose jurisdiction the patient resides and to the Department of Public Health.
- 2. If the case or suspected case of reportable disease, emergency illness or health condition is in a health care facility, the person in charge of such facility shall ensure that reports are made to the local director of health and Department of Public Health. The person in charge shall designate appropriate infection control or record keeping personnel for this purpose.
- 3. If the case or suspected case of reportable disease, emergency illness or health condition is not in a health care facility, and if a health care provider is not in attendance or is not known to have made a report within the appropriate time, such report of reportable disease, emergency illness or health condition shall be made to the local director of health or other health authority within whose jurisdiction the patient lives and the Department of Public Health by:
 - A. the administrator serving a public or private school or day care center attended by any person affected or apparently affected with such disease, emergency illness or health condition;
 - B. the person in charge of any camp;
 - C. the master or any other person in charge of any vessel lying within the jurisdiction of the state;
 - D. the master or any other person in charge of any aircraft landing within the jurisdiction of the state;
 - E. the owner or person in charge of any establishment producing, handling, or processing dairy products, other food or non-alcoholic beverages for sale or distribution;
 - F. morticians and funeral directors

Persons Required to Report Reportable Laboratory Findings

The director of a laboratory that receives a primary specimen or sample, which yields a reportable laboratory finding, shall be responsible for reporting such findings within 48 hours to the local director of health of the town in which the affected person normally resides. In the absence of such information, the reports should go to the town from which the specimen originated and to the Department of Public Health.

IMPORTANT NOTICE

Persons required to report must use the Reportable Disease Confidential Case Report Form PD-23 to report Reportable Diseases, Emergency Illnesses and Health Conditions on the current list unless there is a specialized reporting form or other authorized method available. The director of a clinical laboratory must report laboratory evidence suggestive of reportable diseases using the Laboratory Report of Significant Findings Form OL-15C or other approved format by the DPH. Reporting forms can be found on the DPH "Forms" webpage: (https://portal.ct.gov/DPH/Communications/Forms/Forms) or by calling 860-509-7994. Please follow these guidelines when submitting reports:

- Mailed documents must have "CONFIDENTIAL" marked on the envelope.
- All required information on the form must be completed, including name, address, and phone number of person reporting and healthcare provider, infectious agent, test method, date of onset of illness, and name, address, date of birth, race, ethnicity, gender, and occupation of patient.
- Send one copy of completed report to the DPH via fax (860-509-7910), or mail to: Connecticut Department of Public Health, 410 Capitol Ave., MS#11FDS, P.O. Box 340308, Hartford, CT 06134-0308.
- Unless otherwise noted, send one copy of the completed report to the Director of Health of the patient's town of residence.
- Keep a copy in the patient's medical record.

Raul Pino, MD, MPH Commissioner of Public Health

Matthew L. Cartter, MD, MPH State Epidemiologist

Lynn Sosa, MD Deputy State Epidemiologist Epidemiology and Emerging Infections 860-509-7995 Healthcare Associated Infections 860-509-7995 HIV & Viral Hepatitis 860-509-7900 Immunizations 860-509-7929 Sexually Transmitted Diseases (STD) 860-509-7920 Tuberculosis Control 860-509-7722

Connecticut Epidemiologist

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