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Reportable Diseases, Emergency Illnesses and Health Conditions, and Reportable Laboratory Findings Changes for 2017

As required by Connecticut General Statutes Section 19a-2a and Section 19a-36-A2 of the Public Health Code, the lists of Reportable Diseases, Emergency Illnesses and Health Conditions, and Reportable Laboratory Findings are revised annually by the Department of Public Health (DPH). An advisory committee, consisting of public health officials, clinicians, and laboratorians, contribute to the process. There is 1 modification to both the healthcare provider list and laboratory list. There are 2 additions and 3 modifications to the laboratory list only. National case definitions can be found on the Centers for Disease Control and Prevention's (CDC), National Notifiable Diseases Surveillance System, Case Definitions webpage. Please select link to view the revised 2017 Reportable Disease Confidential Case Report form PD-23 and Laboratory Report of Significant Findings form OL-15C.

Changes to the List of Reportable Diseases, Emergency Illnesses and Health Conditions and Reportable Laboratory Findings

ABCs pathogens

Reporting requirements for invasive bacterial pathogens (Group A Streptococcus, Group B Streptococcus, Haemophilus influenzae, Neisseria meningitidis, Staphylococcus aureus, and Streptococcus pneumoniae) have been modified. Reporting is no longer limited to disease confirmed by isolation; this will allow surveillance to capture invasive disease identified by culture or culture-independent testing methods.

Changes to the List of Reportable Laboratory Findings

NOTE: Extensive changes have been made to the Reportable Laboratory Findings form OL-15C. Please review the form carefully. The form now lists the disease pathogen, and some footnotes have been altered and renumbered.

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Methicillin-sensitive Staphylococcus aureus (MSSA) - invasive disease

Laboratory reporting of invasive MSSA has been <u>added</u>. Nationally, the Emerging Infections Programs (EIP) are transitioning to surveillance for all invasive *S. aureus* isolates, regardless of methicillin-susceptibility status. A pilot study among 4 EIP sites showed MSSA accounted for the majority of *S. aureus* cases. MSSA case-patients had mortality rates similar to invasive methicillin resistant *S. aureus* case-patients, but were found to be younger and healthier. Due to the expected volume of invasive MSSA reports, laboratories will be contacted about reporting options.

Carbapenem-resistant Acinetobacter baumanni (CRAB)

Laboratory reporting of CRAB has been added. Report identification of A. baumannii collected from a clinical site (sterile sites, sputum, urine, and wounds, but not stool) resistant to any tested carbapenem excluding ertapenem (MIC of ≥ 8 mcg/ml) and/or that exhibit production of a carbapenemase demonstrated by a recognized test. Reported cases in individual patients only once every 30 days. Isolates should be forwarded to the DPH laboratory.

Carbapenem-resistant Enterobacteriacea (CRE)

Laboratory reporting of CRE has been <u>modified</u>. Isolates are now required to be submitted with case reports to permit state, regional antimicrobial resistance laboratories, and the CDC to validate and further test isolates for antibiotic susceptibly, to identify the mutations associated with this resistance

(Continued on page 4

REPORTABLE DISEASES, EMERGENCY ILLNESSES and HEALTH CONDITIONS - 2017

The Commissioner of the Department of Public Health (DPH) is required to declare an annual list of Reportable Diseases, Emergency Illnesses and Health Conditions. The Reportable Disease Confidential Case Report form (PD-23) or other disease specific form should be used to report the disease, illness, or condition, Reports (mailed, faxed, or telephoned into the DPH) should include the full name and address of the person reporting and attending physician, name of disease, illness or condition, and full name, address, date of birth, race/ethnicity, gender and occupation of the person affected. Forms can be found on the DPH website. See page 4 for a list of persons required to report Reportable Diseases, Emergency Illnesses and Health Conditions. Mailed reports must be sent in envelopes marked "CONFIDENTIAL." Changes for 2017 are noted in bold and with an asterisk (*).

Category 1 Diseases: Report immediately by telephone on the day of recognition or strong suspicion of disease for those

diseases marked with a telephone (22). Also mail a report within 12 hours.

Category 2 Diseases: Diseases not marked with a telephone are Category 2 diseases. Report by mail within 12 hours of

recognition or strong suspicion of disease.

Acquired Immunodeficiency Syndrome (1,2) Acute flaccid myelitis

Anthrax

Babesiosis

Botulism Brucellosis

California group arbovirus infection

Campylobacteriosis

Carbon monoxide poisoning (3)

Chancroid Chickenpox

Chickenpox-related death

Chikungunya

Chlamydia (C. trachomatis) (all sites)

Cholera

Cryptosporidiosis

Cyclosporiasis

Dengue

Diphtheria

Eastern equine encephalitis virus infection

Ehrlichia chaffeensis infection

Escherichia coli O157:H7 gastroenteritis Gonorrhea

Group A Streptococcal disease, invasive (4*)

Group B Streptococcal disease, invasive (4*)

Haemophilus influenzae disease, invasive (4*)

Hansen's disease (Leprosy)

Healthcare-associated Infections (5)

Hemolytic-uremic syndrome (6)

Hepatitis A

Hepatitis B:

acute infection (2)

HBsAg positive pregnant women

Hepatitis C:

acute infection (2)

positive rapid antibody test result

HIV-1 / HIV-2 infection in: (1)

persons with active tuberculosis disease

 persons with a latent tuberculous infection (history or tuberculin skin test ≥5mm induration by Mantoux technique)

persons of any age

pregnant women

HPV: biopsy proven CIN 2, CIN 3 or AIS or their equivalent (1)

Influenza-associated death (7)

Influenza-associated hospitalization (7)

Lead toxicity (blood level ≥ 15 µg/dL)

Legionellosis

Listeriosis

Lyme disease

Malaria

Measles Melioidosis

Meningococcal disease

Mercury poisoning

Mumps

Neonatal bacterial sepsis (8)

Neonatal herpes (≤ 60 days of age)

Occupational asthma

Foodborne (involving ≥ 2 persons)

Institutional

• Unusual disease or illness (9)

Pertussis

Plaque

Pneumococcal disease, invasive (4*)

Poliomyelitis

Q fever

Rabies

Ricin poisoning

Rocky Mountain spotted fever

Rotavirus

Rubella (including congenital)

Salmonellosis

SARS-CoV

Shiga toxin-related disease (gastroenteritis) Shigellosis

Silicosis

Smallpox

St. Louis encephalitis virus infection

Staphylococcal enterotoxin B pulmonary poisoning

Staphylococcus aureus disease, reduced or

resistant susceptibility to vancomycin (1) Staphylococcus aureus methicillin-

resistant disease, invasive, community acquired (4*,10)

Staphylococcus epidermidis disease, reduced or resistant susceptibility to vancomycin (1)

Syphilis

Tetanus

Trichinosis

Tuberculosis

Tularemia Typhoid fever

Vaccinia disease

Venezuelan equine encephalitis virus infection Vibrio infection (parahaemolyticus, vulnificus, other)

Tiral hemorrhagic fever West Nile virus infection

Yellow fever

Zika virus infection

FOOTNOTES:

- Report only to State.
- As described in the* CDC case definition.
- Includes persons being treated in hyperbaric chambers for suspected CO
- Invasive disease: from sterile fluid (blood, CSF, pericardial, pleural, peritoneal, joint, or vitreous) bone, internal body sites, or other normally sterile site including muscle. *
- 5. Report HAIs according to current CMS pay-for-reporting or pay-forperformance requirements. Detailed instructions on the types of HAIs, facility types and locations, and methods of reporting are available on the DPH website: www.ct.gov/dph/HAI.
- 6. On request from the DPH and if adequate serum is available, send serum from patients with HUS to the DPH Laboratory for antibody testing.
- Reporting requirements are satisfied by submitting the Hospitalized and Fatal Cases of Influenza-Case Report Form in a manner specified by the DPH.
- Clinical sepsis and blood or CSF isolate obtained from an infant ≤ 72 hours of age. Individual cases of "significant unusual illness" are also reportable.
- 10. Community-acquired: infection present on admission to hospital, and person has no previous hospitalizations or regular contact with the health-care setting.

How to report: The PD-23 is the general disease reporting form and should be used if other specialized forms are not available. The PD-23 can be found on the DPH website (<u>www.ct.gov/dph/forms</u>). It can also ordered by writing the Department of Public Health, 410 Capitol Ave., MS#11EPI, P.O. Box 340308, Hartford, CT 06134-0308 or by calling the Epidemiology and Emerging Infections Program (260-509-7994). Specialized reporting forms are available on the DPH <u>website</u> or by calling the following programs: Epidemiology and Emerging Infections Program (860-509-7994) - Hospitalized and Fatal Cases of Influenza, Healthcare Associated Infections (860-509-7995) - National Healthcare Safety Network, HIV/AIDS Surveillance (860-509-7900) - Adult HIV Confidential Case Report form, Immunizations Program (860-509-7929) -Chickenpox Case Report (Varicella) form, Occupational Health Surveillance Program (860-509-7740) - Physician's Report of Occupational Disease, Sexually Transmitted Disease Program (860-509-7920), and Tuberculosis Control Program (860-509-7722).

Telephone reports of Category 1 disease should be made to the local Director of Health for the town in which the patient resides, and to the Epidemiology and Emerging Infections Program (860-509-7994). Tuberculosis cases should be directly reported to the Tuberculosis Control Program (860-509-7722). For the name, address, or telephone number of the local Director of Health for a specific town contact the Office of Local Health Administration (860-509-7660).

For public health emergencies on evenings, weekends, and holidays call 860-509-8000.

REPORTABLE LABORATORY FINDINGS—2017

The director of a clinical laboratory must report laboratory evidence suggestive of reportable diseases. The Laboratory Report of Significant Findings form (OL-15C) can be obtained from the Connecticut Department of Public Health (DPH), 410 Capitol Ave., MS#11EPI, P.O. Box 340308, Hartford, CT 06134-0308; telephone: 860-509-7994 or on the DPH website. The OL-15C is not a substitute for the physician report; it is a supplement to it for verification of diagnosis. Pathogens on the OL-15C are listed in alphabetic order; however, there is a separate section for possible disease indicators of bioterrorism. Changes for 2017 are extensive; review entirely. Those outlined in the article are bolded. Footnotes may have changed or renumbered.

Babesia	Anaplasma phagocytophilum by PCR only	Mercury poisoning
Blood smear		☐ Urine ≥ 35 μg/g creatinine μg/g
Culture	☐ Blood smear ☐ PCR ☐ Other	\square Blood \geq 15 μ g/L μ g/L
Culture	□ microti □ divergens □ duncani □ Unspeciated	Mumps virus (12) (titer)
□ Cluture (1)		Mycobacterium leprae
□PCA	☐ Culture (1) ☐ Non-pertussis Bordetella (1) (specify)	
Carbopnem-resistant Acinerobactericaee (CRE) (1,4) Carbapnem-resistant Acinerobactericaee (CRE) (1,4) Genus Spp		AFB Smear ☐ Positive ☐ Negative
Carbapenem-resistant Acinerobacter baumannii (CRAB) (1.4) Carbapenem-resistant Enterobacteriaceae (CRE) (1.4) Carbapenem-resistant Content Con	Borrelia burgdorferi (2)	If positive ☐ Rare ☐ Few ☐ Numerous
Carbapenem-resistant Acinetobacter baumannii (RAB) (1,4) Carbapenem-resistant Enterboacteriaceae (CRE) (1,4) Genus	California group virus (3) spp	NAAT □ Positive □ Negative □ Indeterminate
Carbapenem-resistant Enterobacteriaceae (CRE) (1,4) Genus Spp	Campylobacter (3)(spp/test type)	Culture Mycobacterium tuberculosis
Carbapenem-resistant Enterobacteriaceae (CRE) (1,4) Genus Spp	Carbapenem-resistant Acinetobacter baumannii (CRAB) (1,4)	☐ Non-TB mycobacterium. (specify <i>M</i>)
Couture Cortes Carboxytemoglobin 2.5% % COHb Couture Cotter	Carbapenem-resistant Enterobacteriaceae (CRE) (1,4)	Neisseria gonorrhoeae (test type)
Chisungunya virus Chismydia trachomatis (test type)	Genus Spp	Neisseria meningitidis, invasive (1,4)
Chisungunya virus Chismydia trachomatis (test type)	Carboxyhemoglobin ≥ 5%% COHb	☐ Culture ☐ Other
Clostridum difficite (5) Corynebacterium diphtheria (1) Cryptosporidium spp (test type) Dengue virus Eastern equine encephalitis virus Escherichia coli O157 (1) (test type) Scherichia coli O157 (Chikungunya virus	Neonatal bacterial sepsis (13) spp
Clostridum difficite (5) Corynebacterium diphtheria (1) Cryptosporidium spp (test type) Dengue virus Eastern equine encephalitis virus Escherichia coli O157 (1) (test type) Scherichia coli O157 (Chlamydia trachomatis (test type)	Plasmodium (1,3) spp
Cryptospordium spp (test type)		Poliovirus
Rotavirus Rotavirus Rotavirus Rubelia virus Ribelia virus Rubelia	Corynebacterium diphtheria (1)	Rabies virus
Rotavirus Rotavirus Rotavirus Rubelia virus Ribelia virus Rubelia	Cryptosporidium spp (test type)	Rickettsia rickettsii
Eastern equine encephalitis virus St. Louis encephalitis virus Staphylococcus aureus	Cyclospora spp (test type)	
St. Louis encephalitis virus Ig. All annotions St. Louis encephalitis virus Ig. All annotions St. Louis encephalitis virus Ig. All annotions Colliture Other St. All annotions Colliture Other Staphylococcus aureus, invasive (4) Culture Other Imperite Impe		Rubella virus (12) (titer)
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Giardia spp Group A Streptococcus, invasive (1,4)	Ehrlichia chaffeensis by PCR only	St. Louis encephalitis virus
Giardia spp Group A Streptococcus, invasive (1,4)	Escherichia coli O157(1) (test type)	Salmonella (1,3) (serogroup/serotype/test type)
Group B Streptococcus, invasive (4)	Giardia spp	SARS-CoV (1) □ IgM/IgG
Group B Streptococcus, invasive (4)	Group A <i>Streptococcus</i> , invasive (1,4) □ Culture □ Other	□ PCR (specimen) □ Other
Haemophilus ducreyi Haemophilus influenzae, invasive (1,4) □ Culture □ Other	Group B Streptococcus, invasive (4) ☐ Culture ☐ Other	Shiga toxin (1) ☐ Stx1 ☐ Stx2 ☐ Type Unknown
Hepatitis A virus (HAV) IgM anti-HAV (6)	Haemophilus ducreyi	Shigella (1,3) (serogroup/spp test type)
Hepatitis B anti-HBs (7) □ Positive (titer) □ Negative □ Indicillin-sensitive Staphylococcus aureus, vancomycin MIC ≥ 4 µg/mL (1) Hepatitis C virus (HCtV)□Rapid antibody□RNA (8) □ Genotype (8) Herpes simplex virus (infants ≤ 60 days of age) (specify type) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (9) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (1,9) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV Viral Load (all results) (9) □ HIV genotype (9) □ HIV genotype (9) □ CD4 count: □ cells/uL; □ // (9) HPV (report only to the State) (10) Biopsy proven □ CIN 2 □ CIN 3 □ AIS □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (11) □ Finger stick level □ Legionella pneumophila □ Culture □ DFA □ Ag positive □ Regional report only construction of the state of the pour-fold serologic change (titlers) □ Listeria monocytogenes (1) Hepatitis C virus (HCtV) □ Repid antibody □ RNA (8) □ Genotype (8) MIC to vancomycin □ MIC ≥ 4 µg/mL (1) MIC to vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella preusonycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin □ L	Haemophilus influenzae, invasive (1,4) ☐ Culture ☐ Other	Staphylococcus aureus, invasive (4) ☐ Culture ☐ Other
Hepatitis B anti-HBs (7) □ Positive (titer) □ Negative □ Indicillin-sensitive Staphylococcus aureus, vancomycin MIC ≥ 4 µg/mL (1) Hepatitis C virus (HCtV)□Rapid antibody□RNA (8) □ Genotype (8) Herpes simplex virus (infants ≤ 60 days of age) (specify type) □ Culture □ PCR □ IFA □ Ag detection HIV Related Testing (report only to the State) (9) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (1,9) HIV 1 □ Positive □ Neg/Ind □ HIV 2 □ Positive □ Neg/Ind □ HIV Viral Load (all results) (9) □ HIV genotype (9) □ HIV genotype (9) □ CD4 count: □ cells/uL; □ // (9) HPV (report only to the State) (10) Biopsy proven □ CIN 2 □ CIN 3 □ AIS □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (11) □ Finger stick level □ Legionella pneumophila □ Culture □ DFA □ Ag positive □ Regional report only construction of the state of the pour-fold serologic change (titlers) □ Listeria monocytogenes (1) Hepatitis C virus (HCtV) □ Repid antibody □ RNA (8) □ Genotype (8) MIC to vancomycin □ MIC ≥ 4 µg/mL (1) MIC to vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ MC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella preusonycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin MIC ≥ 4 µg/mL (1) MIC to vancomycin □ Legonella vancomycin □ L	Hepatitis A virus (HAV) IgM anti-HAV (6) ALT AST ☐ Not Done	☐ methicillin-resistant
Hepatitis C virus (HCV) □Rapid antibody □RNA (8) □ Genotype (8) Herpes simplex virus (infants ≤ 60 days of age) (specify type) □ Culture □ PCR □ □FA □ Ag detection HIV Related Testing (report only to the State) (9) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (1,9) HIV 1 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (9) □ □ CD4 count: □ cells/uL; □ % (9) HPV (report only to the State) (10) Stophylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin □ µg/mL Staphylococcus epidermidis, vancomycin □ µg/mL Staphylococus epidermidis (1) Staphylococus epidermidis (1) Staphylococus epidermidis (1) Staphylococus epidermidis (1) Stap	Hepatitis B anti-HBs (7) ☐ Positive (titer) ☐ Negative	☐ methicillin-sensitive
Herpes simplex virus (infants ≤ 60 days of age) (specify type) Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin µg/mL MIC to vancomycin		Staphylococcus aureus, vancomycin MIC ≥ 4 µg/mL (1)
Herpes simplex virus (infants ≤ 60 days of age) (specify type) Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1) MIC to vancomycin µg/mL MIC to vancomycin	Hepatitis C virus (HCV) □Rapid antibody □RNA (8) □ Genotype (8)	MIC to vancomycin μg/mL
Culture	Herpes simplex virus (infants < 60 days of age) (specify type)	Staphylococcus epidermidis, vancomycin MIC ≥ 32 µg/mL (1)
HIV Related Testing (report only to the State) (9) □ Detectable Screen (IA) Antibody Confirmation (WB/IFA/Type-diff) (1,9) HIV 1 □ Positive □ Neg/Ind □ HIV NAAT (or qualitative RNA) □ Detectable □ Not Detectable □ HIV Viral Load (all results) (9) □ HIV genotype (9) □ CD4 count: cells/uL; % (9) HPV (report only to the State) (10) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent (specify) Influenza virus: □ Rapid antigen (2) □ RT-PCR □ Culture-confirmed □ Type A □ Type B □ Type Unknown □ Subtype □ Lead poisoning (blood lead ≥10 µg/dL □ Venous level □ µg/dL Legionella pneumophila □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) □ Listeria monocytogenes (1) Streptococcus pneumoniae □ Culture (1,4) □ Urine antigen □ Other (4) □ Treponema pallidum □ RPR (titer) □ PTA □ EIA Trichinella □ Culture □ PCR □ DFA □ Other □ Vibrio (1,3) (spp/test virus, acute □ Culture □ PCR □ DFA □ Other □ Vibrio (1,3) (spp/test type) West Nile virus Yellow fever virus Yellow fever virus Yellow fever virus Yersinia, not pestis (3) (spp/test type) Zika virus BIOTERRORISM possible disease indicators (14) Bacillus anthracis (1) Burkholderia mallei (1) Clostridium botulinum Francisella tularensis Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Yersinia pestis (1)		MIC to vancomycin µg/mL
Antibody Confirmation (WB/IFA/Type-diff) (1,9) HIV 1 Positive Neg/Ind HIV 2 Positive Neg/Ind HIV 1 Positive Neg/Ind HIV NAAT (or qualitative RNA) Detectable Not Detectable HIV Viral Load (all results) (9) HIV genotype (9) CD4 count: cells/uL; % (9) Wibrio (1,3) (spp/test type) West Nile virus West Nile virus Yellow fever v	HIV Related Testing (report only to the State) (9)	Streptococcus pneumoniae
HIV 1 □ Positive □Neg/Ind	☐ Detectable Screen (IA)	☐ Culture (1,4) ☐ Urine antigen ☐ Other (4)
HIV 1 □ Positive □Neg/Ind	Antibody Confirmation (WB/IFA/Type-diff) (1,9)	Treponema pallidum □ RPR (titer) □ FTA □ EIA
HIV Viral Load (all results) (9)	HIV 1 ☐ Positive ☐ Neg/Ind HIV 2 ☐ Positive ☐ Neg/Ind	□ VDRL (titer) □ TPPA
□ HIV genotype (9) □ CD4 count: cells/uL;	☐ HIV NAAT (or qualitative RNA) ☐ Detectable ☐ Not Detectable	Trichinella
□ CD4 count: cells/uL;% (9) HPV (report only to the State) (10) Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent (specify) Influenza virus: □ Rapid antigen (2) □ RT-PCR □ Culture-confirmed □ Type A □ Type B □ Type Unknown □ Subtype Lead poisoning (blood lead ≥10 μg/dL <48 hrs; 0-9 μg/dL monthly) (11) □ Finger stick level □ μg/dL □ Venous level □ μg/dL Legionella pneumophila □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) □ Culture-confirmed □ Type B □ Type Unknown □ Subtype BIOTERORISM possible disease indicators (14) Bacillus anthracis (1) Burkholderia mallei (1) Clostridium botulinum Francisella tularensis Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Versinia pestis (1)	☐ HIV Viral Load (all results) (9)	Varicella-zoster virus, acute
HPV (report only to the State) (10) Biopsy proven	☐ HIV genotype (9)	☐ Culture ☐ PCR ☐ DFA ☐ Other
Biopsy proven □ CIN 2 □ CIN 3 □ AIS or their equivalent (specify) □ CIN 2 □ Type B □ Type Unknown □ Type A □ Type B □ Type Unknown □ Subtype □ Lead poisoning (blood lead ≥10 μg/dL <48 hrs; 0-9 μg/dL monthly) (11) □ Finger stick level □ μg/dL □ Venous level □ μg/dL □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) □ Culture-confirmed □ Type B □ Type Unknown □ Subtype □ Type B □ Type Unknown □ Subtype □ Type B □ Type Unknown □ Subtype □ BIOTERRORISM possible disease indicators (14) Bacillus anthracis (1) Burkholderia mallei (1) Clostridium botulinum Coxiella burnetii Francisella tularensis Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Versinia, not pestis (3) (spp/test type) □ Type Unknown Like virus BIOTERRORISM possible disease indicators (14) Bacillus anthracis (1) Burkholderia mallei (1) Coxiella burnetii Francisella tularensis Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Versinia pestis (1)	☐ CD4 count: cells/uL;% (9)	Vibrio (1,3) (spp/test type)
or their equivalent (specify)	HPV (report only to the State) (10)	West Nile virus
Influenza virus: ☐ Rapid antigen (2) ☐ RT-PCR ☐ Culture-confirmed ☐ Type A ☐ Type B ☐ Type Unknown ☐ Subtype ☐ Type B ☐ Type Unknown ☐ Subtype ☐ Type Unknown ☐ Subtype ☐ Type B ☐ Type Unknown ☐ Subtype ☐ Subtype ☐ Subtype ☐ Subtype ☐ Subtype ☐ Subtype ☐ Pringer stick level ☐ pringer	Biopsy proven ☐ CIN 2 ☐ CIN 3 ☐ AIS	
□ Type A □ Type B □ Type Unknown □ Subtype □	or their equivalent (specify)	Yersinia, not pestis (3) (spp/test type)
□ Subtype Bacillus anthracis (1) Brucella spp (1) Lead poisoning (blood lead ≥10 μg/dL <48 hrs; 0-9 μg/dL monthly) (11) □ Finger stick level μg/dL □ Venous level μg/dL Legionella pneumophila □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Listeria monocytogenes (1) Bacillus anthracis (1) Brucella spp (1) Burkholderia mallei (1) Costridium botulinum Coxiella burnetii Francisella tularensis Ricin Variola virus (1) Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Yersinia pestis (1)	Influenza virus: ☐ Rapid antigen (2) ☐ RT-PCR ☐ Culture-confirmed	Zika virus
□ Subtype Bacillus anthracis (1) Brucella spp (1) Lead poisoning (blood lead ≥10 μg/dL <48 hrs; 0-9 μg/dL monthly) (11) □ Finger stick level μg/dL □ Venous level μg/dL Legionella pneumophila □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Listeria monocytogenes (1) Bacillus anthracis (1) Brucella spp (1) Burkholderia mallei (1) Costridium botulinum Coxiella burnetii Francisella tularensis Ricin Variola virus (1) Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Yersinia pestis (1)	☐ Type A ☐ Type B ☐ Type Unknown	BIOTERRORISM possible disease indicators (14)
Lead poisoning (blood lead ≥10 µg/dL <48 hrs; 0-9 µg/dL monthly) (11) □ Finger stick level µg/dL □ Venous level µg/dL Legionella pneumophila □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) Listeria monocytogenes (1) □ Finger stick level µg/dL □ Venous level µg/dL Clostridium botulinum Francisella tularensis Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Yersinia pestis (1)	☐ Subtype	
□ Finger stick level □ μg/dL □ Venous level □ μg/dL □ Clostridium botulinum □ Coxiella burnetii □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) □ Culture □ DFA □ Ag positive □ Four-fold serologic change (titers) □ Venezuelan equine encephalitis virus Listeria monocytogenes (1) Venous level □ μg/dL □ Clostridium botulinum □ Coxiella burnetii Ricin □ Variola virus (1) Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Versinia pestis (1)	Lead poisoning (blood lead ≥10 μg/dL <48 hrs; 0-9 μg/dL monthly) (11)	
Legionella pneumophila ☐ Culture ☐ DFA ☐ Ag positive ☐ Four-fold serologic change (titers) ☐ Listeria monocytogenes (1) ☐ Francisella tularensis ☐ Staphylococcus aureus - enterotoxin B Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Versinia pestis (1)	□ Finger stick level µg/dL □ Venous level µg/dL	
□ Culture □ DFA □ Ag positive Staphylococcus aureus - enterotoxin B Variola virus (1) □ Four-fold serologic change (titers) Venezuelan equine encephalitis virus Listeria monocytogenes (1) Viral agents of hemorrhagic fevers Versinia pestis (1)		
Listeria monocytogenes (1) Venezuelan equine encephalitis virus Viral agents of hemorrhagic fevers Yersinia pestis (1)		
Listeria monocytogenes (1) Viral agents of hemorrhagic fevers Yersinia pestis (1)	0 0 \ / 	
	Listeria monocytogenes (1)	· · · · · · · · · · · · · · · · · · ·
	4. Condinates guitage or clide to the DDIII - hands and include the condinates of th	, , ,

- confirmation. For Salmonella, Shigella, and Vibrio tested by non-culture methods, send the isolate from reflex testing or if positive by CIDT and no isolate or culture results send stool specimen. For Shiga toxin-related disease, send positive broth or stool in transport media. For positive HIV, send ≥ 0.5mL residual serum.
- Only laboratories with electronic file reporting are required to report positive results.
- Specify species/serogroup/serotype.
- 4. Sterile site: defined as sterile fluids (blood, CSF, pericardial, pleural, peritoneal, joint, or vitreous), bone, internal body site (lymph node, brain, heart, liver, spleen, kidney, pancreas, or ovary), or other normally sterile site including muscle. For CRE and CRAB, also
- include urine or sputum, but not stool; and for CRAB also include wounds.
- 5. Submit reports of all *C. difficile* positive stool samples according to DPH instructions.
- Report the peak liver function tests (ALT, AST) conducted within one week of patient's HAV IgM positive test, if available. Check "Not Done" when appropriate.
- 7. Negative HBsAg and all anti-HBs results are reportable only for children ≤ 2 years old.
- Report all RNA results. Genotypes and Negative RNA results only reportable by electronic file reporting.
- Report all HIV antibody, antigen, viral load, and qualitative NAAT results. HIV genotype (DNA sequence) and all CD4 results are reportable by electronic file.
- If adequate tissue is available, send fixed tissue from the specimen used to diagnose CIN 2, 3 or cervical AIS or their equivalent for HPV typing according to DPH instructions.
- Report lead results ≥ 10 μg/dL within 48 hours to the Local Health Director and the DPH; submit ALL lead results at least monthly to the DPH only.
- Report all IgM positive titers, but only IgG titers that are considered significant by the laboratory performing the test.
- Report all bacterial isolates from blood or CSF from an infant ≤ 72 hours of age.
- Report by telephone to the DPH, weekdays 860-509-7994; evenings, weekends, and holidays 860-509-8000.

and to establish and maintain an isolate bank of novel and emerging CRE strains in conjunction with the CDC.

Hepatitis B

Laboratory reporting of Hepatitis B has been $\underline{\text{modified}}$. Laboratories are now required to report negative hepatitis B surface antigen results and all surface antibody results for children aged ≤ 2 years. This will facilitate follow up of children born to hepatitis B positive women.

Salmonella, Shigella, and Vibrio

Laboratory reporting of *Salmonella*, *Shigella*, and *Vibrio* has been <u>modified</u>. Laboratories are now required to send stool specimens for these pathogens to the DPH Public Health laboratory if no isolate is recovered after reflex culture for a positive culture-independent testing result. Isolate recovery (at the clinical and/or public health laboratory) is necessary for subsequent isolate-based molecular testing to aid in outbreak detection and monitoring of antibiotic resistance.

Persons Required to Report Reportable Diseases, Emergency Illnesses and Health Conditions

- 1. Every health care provider who treats or examines any person who has or is suspected to have a reportable disease, emergency illness or health condition shall report the case to the local director of health or other health authority within whose jurisdiction the patient resides and to the Department of Public Health.
- 2. If the case or suspected case of reportable disease, emergency illness or health condition is in a health care facility, the person in charge of such facility shall ensure that reports are made to the local director of health and Department of Public Health. The person in charge shall designate appropriate infection control or record keeping personnel for this purpose.
- 3. If the case or suspected case of reportable disease, emergency illness or health condition is not in a health care facility, and if a health care provider is not in attendance or is not known to have made a report within the appropriate time, such report of reportable disease, emergency illness or health condition shall be made to the local director of health or other health authority within whose jurisdiction the patient lives and the Department of Public Health by:
 - A. the administrator serving a public or private school or day care center attended by any person affected or apparently affected with such disease, emergency illness or health condition;
 - B. The person in charge of any camp;
 - C. The master or any other person in charge of any vessel lying within the jurisdiction of the state;
 - D. The master or any other person in charge of any aircraft landing within the jurisdiction of the state;
 - E. The owner or person in charge of any establishment producing, handling, or processing dairy products, other food or non-alcoholic beverages for sale or distribution:
 - F. Morticians and funeral directors.

Persons Required to Report Reportable Laboratory Findings

The director of a laboratory that receives a primary specimen or sample, which yields a reportable laboratory finding, shall be responsible for reporting such findings within 48 hours to the local director of health of the town in which the affected person normally resides. In the absence of such information, the reports should go to the town from which the specimen originated and to the Department of Public Health.

IMPORTANT NOTICE

Reporting forms are available on the Connecticut Department of Public Health (DPH) website. Persons required to report must use the Reportable Disease Confidential Case Report Form PD-23 to report reportable diseases, emergency illnesses and health conditions on the current list unless there is a specialized reporting form available. The director of a clinical laboratory must report laboratory evidence suggestive of reportable diseases using the Laboratory Report of Significant Findings Form OL-15C or other approved format by the DPH. Reporting forms can be obtained by contacting the Connecticut Department of Public Health, 410 Capitol Ave., MS#11EPI, P.O. Box 340308, Hartford, CT 06134-0308; telephone: 860-509-7994, fax: 860-509-7910 or from the website (www.ct.gov/dph/forms). Please follow these guidelines when submitting reports:

- Any mailed documents must have "CONFIDENTIAL" marked on the envelope.
- Complete all required information including name, address, and phone number of person reporting and healthcare provider, infectious agent, test method, date of onset of illness, and name, address, date of birth, race/ethnicity, gender, and occupation of the person affected.
- Send one copy of completed report to the DPH via fax, or mail to Connecticut Department of Public Health, 410 Capitol Ave., MS#11EPI, P.O. Box 340308, Hartford, CT 06134-0308.
- Send one copy to the Director of Health of the patient's town of residence.
- Keep a copy for the patient's medical record.

Raul Pino, MD, MPH Commissioner of Public Health

Matthew L. Cartter, MD, MPH State Epidemiologist

Lynn Sosa, MD Deputy State Epidemiologist Epidemiology and Emerging Infections 860-509-7995 Healthcare Associated Infections 860-509-7995 HIV & Viral Hepatitis 860-509-7900 Immunizations 860-509-7929 Sexually Transmitted Diseases (STD) 860-509-7920 Tuberculosis Control 860-509-7722

Connecticut Epidemiologist

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