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Science of the Total Environment

journal homepage: www.elsevier.com/locate/scitotenv

Yale School of Public Health Symposium: An overview of the challenges and opportunities associated with *per*- and polyfluoroalkyl substances (PFAS)☆



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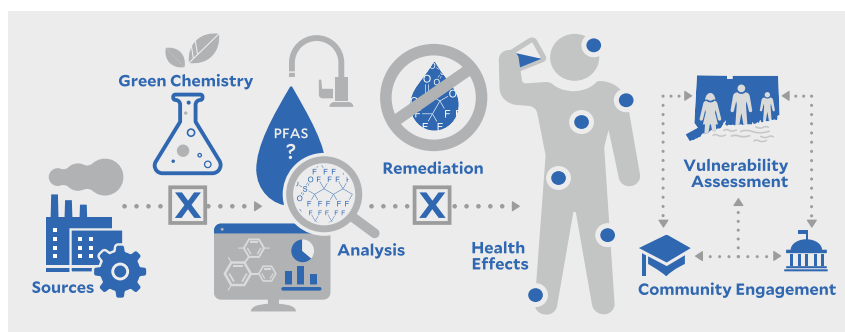
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GRAPHICAL ABSTRACT



☆ The views expressed herein are those of the authors and do not necessarily reflect those of the Connecticut Department of Energy and Environmental Protection or the Connecticut Department of Public Health.

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ARTICLE INFO

Article history:

Received 17 December 2020

Received in revised form 22 February 2021

Accepted 24 February 2021

Available online 3 March 2021

Editor: Damia Barcelo

Keywords:

Per- and polyfluoroalkyl substances (PFAS)

Exposure

Health effects

Environmental contamination

Drinking water

ABSTRACT

On December 13, 2019, the Yale School of Public Health hosted a symposium titled “Per- and Polyfluoroalkyl Substances (PFAS): Challenges and Opportunities” in New Haven, Connecticut. The meeting focused on the current state of the science on these chemicals, highlighted the challenges unique to PFAS, and explored promising opportunities for addressing them. It brought together participants from Yale University, the National Institute of Environmental Health Sciences, the University of Massachusetts Amherst, the University of Connecticut, the Connecticut Agricultural Experiment Station, the Connecticut Departments of Public Health and Energy and Environmental Protection, and the public and private sectors. Presentations during the symposium centered around several primary themes. The first reviewed the current state of the science on the health effects associated with PFAS exposure and noted key areas that warranted future research. As research in this field relies on specialized laboratory analyses, the second theme considered commercially available methods for PFAS analysis as well as several emerging analytical approaches that support health studies and facilitate the investigation of a broader range of PFAS. Since mitigation of PFAS exposure requires prevention and cleanup of contamination, the third theme highlighted new nanotechnology-enabled PFAS remediation technologies and explored the potential of green chemistry to develop safer alternatives to PFAS. The fourth theme covered collaborative efforts to assess the vulnerability of in-state private wells and small public water supplies to PFAS contamination by adjacent landfills, and the fifth focused on strategies that promote successful community engagement. This symposium supported a unique interdisciplinary coalition established during the development of Connecticut’s PFAS Action Plan, and discussions occurring throughout the symposium revealed opportunities for collaborations among Connecticut scientists, state and local officials, and community advocates. In doing so, it bolstered the State of Connecticut’s efforts to implement the ambitious initiatives that its PFAS Action Plan recommends.

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1. Introduction

Per- and polyfluoroalkyl substances (PFAS), a family of >9000 synthetic organic chemicals (EPAa), have captured the attention of people across the globe due to a growing concern about the health risks posed by widespread PFAS contamination in drinking water sources and other environmental media. While PFAS vary widely in their physical and chemical properties, they all contain at least one chain of carbon atoms in which one or more of the carbon atoms is perfluorinated, i.e., has fluorine atoms attached at all bonding sites not occupied by another carbon atom (Buck et al., 2011). Since their initial introduction in the 1940s, PFAS have become pervasive in consumer products and industrial processes because of the unique properties imparted by their chemical structures, including stability, heat resistance, friction reduction abilities, and oil and water repellence (Buck et al., 2012). However, as an unintended consequence of these same useful properties, many PFAS are now persistent pollutants that spread throughout the environment, contaminate food and drinking water sources, and ultimately bioaccumulate in animals and humans (Sunderland et al., 2019). Toxicological and epidemiological research has associated exposure to certain PFAS, particularly certain long-chain perfluoroalkyl acids, with a wide range of adverse health effects (reviewed in depth in Fenton et al., 2020), including thyroid disruption (Andersson et al., 2019; Ballesteros et al., 2017; Blake et al., 2018; Caron-Beaudoin et al., 2019), ulcerative colitis (Steenland et al., 2018; Steenland et al., 2013), high cholesterol (Lin et al., 2019; Nelson et al., 2010), pregnancy-induced hypertension (Starling et al., 2014), decreased immune responsiveness (DeWitt et al., 2019), and kidney and testicular cancer (Barry et al., 2013; Shearer et al., 2020). Measurable levels of some PFAS are present in the blood of over 95% of U.S. residents, and PFAS contamination has been discovered in drinking water nationwide (Hu et al., 2016). During the third Unregulated Contaminant Monitoring Rule (UCMR3; 2013–2015) water survey, the U.S. Environmental Protection Agency (EPA) found that 1.3% of the nation’s largest public water systems, which provide drinking water to an estimated 5.5 million people, contained at least one PFAS compound in concentrations exceeding its reference concentration of 70 ng/L (ppt) (EPA, 2016).

In the absence of timely federal action to regulate PFAS or set enforceable drinking water standards and in response to community concerns, many U.S. states have taken independent action to safeguard their residents against the health risks posed by PFAS (reviewed in

Blake and Fenton, 2020 and Post, 2020). In July 2019, Connecticut Governor Ned Lamont established the Interagency PFAS Task Force to advise his administration and formulate an action plan containing a comprehensive state strategy to address PFAS. The task force was led by the Commissioners of the Connecticut Departments of Public Health (CTDPH) and Energy and Environmental Protection (CTDEEP) and comprised representatives from 18 state agencies and entities. To enable all affected stakeholders to take part in the process, the task force established three subcommittees, each open to public participation, to discuss strategies to (1) minimize environmental exposure of Connecticut residents to PFAS, (2) minimize future releases of PFAS into the environment, and (3) identify, assess, and clean up historical releases of PFAS into the environment, respectively. This process brought together key academic, government, and private- and public-sector stakeholders from across the state. An action plan, developed by the Task Force and its subcommittees, was released in draft form in October 2019 for public comment. After revisions to reflect public input, the finalized PFAS Action Plan was delivered to Governor Lamont on November 1, 2019 and released to the public soon thereafter (The Connecticut Interagency PFAS Task Force, 2019). The plan laid out a series of actions that the State of Connecticut could take to protect public health, identify and remediate existing PFAS pollution, prevent future pollution, and enhance outreach and communication with the general public on the adverse health effects of PFAS exposure and strategies for mitigating exposure.

To foster continued collaboration between local scientists, government officials, public citizens, and other parties, the Yale School of Public Health hosted a daylong symposium on December 13, 2019 titled “Per- and Polyfluoroalkyl Substances (PFAS): Challenges and Opportunities.” This symposium drew participants from Yale, CTDEEP, CTDPH, the Connecticut Agricultural Experiment Station, the National Institute of Environmental Health Sciences (NIEHS), the University of Massachusetts Amherst, the University of Connecticut, and key stakeholders in the public and private sectors. Presentations during the symposium centered around several primary themes. The first reviewed the current state of the science on the health effects of PFAS and noted key research gaps that require further study. As research in this field relies on specialized laboratory analyses, the second theme considered commercially available methods for PFAS analysis as well as several emerging analytical approaches that support new health studies and facilitate the investigation of a broader range of PFAS. Since mitigation of PFAS exposure requires prevention and cleanup of contamination, the third

theme highlighted new nanotechnology-enabled PFAS remediation technologies and explored the potential of green chemistry to develop safer alternatives to PFAS. The fourth theme covered a collaboration between Yale researchers and CTDEEP to assess the vulnerability of private wells and small public water supplies to PFAS contamination by adjacent landfills, and the fifth focused on strategies that promote successful community engagement. Discussions occurring throughout the symposium revealed opportunities for collaborations that would support ongoing CTDEEP and CTDPH efforts to implement the initiatives recommended in the Connecticut PFAS Action Plan. The highlights of these sessions' presentations are summarized herein. This summary provides a valuable snapshot of early dialogue between researchers, government officials, and representatives of local health and nonprofit organizations convened to share cutting-edge scientific advances and coordinate future multisector collaborations to help the state confront an environmental health issue of national and global importance.

2. Health effects

Decades of widespread PFAS use have led to pervasive environmental contamination and human exposure. Comprehensive research on PFAS-related health effects is therefore necessary to inform the risk assessment process used by government officials to derive health-based guidelines and standards that protect the public from potentially unsafe levels of exposure. As exposure occurs through various pathways, including consumer product use and ingestion of contaminated food and water, these officials face a daunting task. Many regulatory and research efforts have initially focused on human exposure through drinking water. In recent years, a growing number of epidemiological studies investigating populations with different levels of PFAS exposure have identified probable links between certain PFAS and various human health effects, which are generally supported by findings in toxicological studies conducted in laboratory rodent models. These studies have collectively demonstrated that pregnant mothers and developing offspring are the sub-populations most sensitive to low levels of PFAS exposure, which highlights the need to study adverse effects on maternal and infant health as particularly essential for risk assessment purposes (Goeden et al., 2019). Recent research has significantly enhanced our understanding of the health implications of PFAS exposure. However, major remaining data gaps make it challenging for officials to effectively assess the risks posed by PFAS and safeguard public health (Fenton et al., 2020). Moving forward, it is important for scientists to fill in these data gaps by investigating additional PFAS, geographical regions, and health effects, considering the implications of exposure to mixtures of PFAS, and elucidating the mechanisms responsible for adverse health effects.

Environmental epidemiological studies using data-rich prospective birth cohorts have provided valuable information on how PFAS exposure influences the health of pregnant women and of children in the earliest and most sensitive stages of development. Developing human fetuses can be exposed to PFAS *in utero* through active or passive transplacental transfer (Eryasa et al., 2019; Mamsen et al., 2019); after birth, infants can be exposed through breastfeeding and/or formula made with contaminated water, and by PFAS in the home environment. Studies using the Danish National Birth Cohort have provided evidence linking PFAS exposure to a range of pregnancy complications and neurodevelopmental effects (Ernst et al., 2019; Liew et al., 2020; Liew et al., 2018; Liew et al., 2014; Liew et al., 2015; Meng et al., 2018). Specifically, these studies have linked prenatal PFAS exposure with altered maternal thyroid hormone function during early pregnancy, an increased risk of cerebral palsy in male offspring (Liew et al., 2014), increased risks of miscarriage and preterm birth (Liew et al., 2020; Meng et al., 2018), and sex-specific effects on the onset of puberty (Ernst et al., 2019). Studies using the Faroe Islands birth cohorts have provided further evidence of associations between PFAS exposure and a broad range of health effects in children, including decreased birth weight (Xiao et al., 2020), impaired neurodevelopment (Oulhote et al.,

2016), immunosuppressive effects (Grandjean et al., 2012), altered thyroid hormone levels (Xiao et al., 2020), childhood behavioral problems (Oulhote et al., 2016), and microbiome disruption (Oulhote et al., 2019). However, not all epidemiological studies examine the same set of PFAS, and the exposure levels may vary across populations. As such, it is challenging to corroborate findings across studies. To date, research efforts on the health risks of PFAS have predominantly focused on perfluorooctanoic acid (PFOA) and perfluorooctane sulfonic acid (PFOS), both of which have already been largely phased out of non-essential uses in several parts of the world but remain widely detected in waste streams and the environment (Boiteux et al., 2016; Clara et al., 2008; Mussabek et al., 2019). The health impacts of the many other types of PFAS currently in use (including the shorter-chain compounds that have been used as replacements for PFOA and PFOS) are still poorly understood. Moreover, most studies rely on single-pollutant models that might not adequately capture the cumulative effects of exposure to multiple PFAS, let alone other coexistent environmental toxicants.

Epidemiological studies have also provided evidence of geographical variations in PFAS exposure and the health effects associated with such exposure. Levels of exposure vary widely on both local and global scales, likely because the level of PFAS exposure in a given population depends on a myriad of factors ranging from its proximity to individual point sources of pollution to the effects of regulatory action and industry phaseouts on PFAS levels in the marketplace. For instance, unlike many western countries, China has yet to phase out industrial use of PFOA and PFOS. However, epidemiological studies of PFAS exposure in China are relatively scarce and primarily focused on birth or pregnancy-related outcomes (Cao et al., 2018; Chen et al., 2013b; Chen et al., 2012; Chen et al., 2018; Huang et al., 2019; Li et al., 2017; Lien et al., 2016; Shi et al., 2017; Wang et al., 2016a; Wang et al., 2019a; Wang et al., 2018a; Wang et al., 2016b; Wang et al., 2015a; Wang et al., 2018b; Wu et al., 2012; Yao et al., 2019). Studies measuring levels of PFAS in cord blood indicate that PFOA and PFOS account for the majority of total PFAS exposure in mainland China, whereas longer-chain ($C \geq 9$) compounds are the dominant PFAS in Taiwan (Shi et al., 2017; Wang et al., 2016a). These studies have also revealed levels of PFAS in mainland China that are lower than those found in many western countries (Shi et al., 2017); this could change in the future as PFAS manufacturing shifts from the western world to China and developing countries (Land et al., 2018). Birth cohort studies in China have shown relatively consistent results linking PFAS exposure to decreased birth weight (Chen et al., 2012; Chen et al., 2018; Li et al., 2017; Wang et al., 2016b; Wu et al., 2012) and demonstrated that the length and structure of PFAS affect their toxicity. Whereas longer-chain PFAS in maternal blood were associated with adverse effects on birth weight (Wang et al., 2015a), those in cord blood were not (Chen et al., 2012). This may reflect size-dependent differences in the compounds' transplacental transfer efficiency (Wang et al., 2015a). In addition, branched PFAS were associated with stronger adverse effects than their linear counterparts (Li et al., 2017). Other results of these studies have proven less consistent, such as the effect of PFAS on birth length (Cao et al., 2018; Chen et al., 2018; Shi et al., 2017; Wang et al., 2019a; Wang et al., 2015a; Wu et al., 2012) and the influence of the sex of the infant on PFAS-related health effects (Cao et al., 2018; Li et al., 2017; Shi et al., 2017; Wang et al., 2016a). A few studies have also shown PFAS to have adverse effects on gestational hypertension disorders (Huang et al., 2019), some neurobehavioral development endpoints (Chen et al., 2013b; Wang et al., 2015a), and blood glucose levels (Wang et al., 2018b). Additional studies are necessary to reliably characterize the levels of PFAS exposure and resulting health impacts in China as well as understudied countries and regions around the world. Information on health effects in countries that manufacture and export PFAS-containing goods is particularly crucial for understanding the PFAS-related health risks of consumer products derived from globalized supply chains.

Animal-based studies are still regarded as necessary for deriving the "toxicity values" that are foundational to government officials' risk

assessment processes (EPA, 2002). Such studies have directly linked PFAS exposure with numerous health effects, and their findings are generally concordant with those of epidemiological studies (Fenton et al., 2020). For instance, studies using the developing mouse model have shown PFOA exposure to decrease birth weight, increase excess weight gain in offspring and the pregnant mother, and impair lactation (Blake et al., 2020; Hines et al., 2009; Koustas et al., 2014; White et al., 2007). Similar effects in humans have been associated with PFOA exposure (Ashley-Martin et al., 2016; Halldorsson et al., 2012; Johnson et al., 2014; Karlsen et al., 2017; Romano et al., 2016; Timmermann et al., 2017; Xiao et al., 2020). Unlike epidemiological studies, toxicological studies in rodent laboratory models are able to provide evidence of causality, allow for investigation into modes of action, and enable scientists to test the dependence of a given health outcome on individual variables, including dose magnitude, dose timing, and specific PFAS compound(s). This has made it possible to establish, for example, that developing tissues are highly sensitive to PFAS, resulting in persistent effects (e.g., mammary gland development), and that since developing fetuses are more susceptible than adults, effective doses of PFAS are lower in developmental exposure situations (e.g., liver gene expression). For the few compounds whose toxicology has been extensively studied (such as PFOA), toxicologists at state and federal agencies have derived minimal risk levels (MRLs) using non-cancer reference doses based on a variety of different target effects, including increased liver weight, weakened immune response, delayed mammary gland development, delayed bone ossification, neurobehavioral effects, and accelerated male puberty (Post, 2020). It should be recognized that choice of target effect has major policy implications. Since some developmental effects may result from PFAS doses below those causing an increase in liver weight, selection of certain developmental target effects gives rise to lower MRLs. As reviewed by Post (2020), the range of different target effects that toxicologists at state agencies and the EPA have used to derive reference doses for PFOA contributes to the approximately four-fold range in the values of their PFOA drinking water guidelines. Recent toxicological studies have linked delayed mammary gland development (a particularly sensitive effect that persists into adulthood, affects lactation, and has a similar mode of action in mice and humans) to both PFOA and a short-chain replacement compound known as GenX (Fenton, personal communication). These results highlight the critical need for more studies focused on PFAS other than PFOA and PFOS.

Although many of the individual effects of PFAS exposure identified in toxicological and epidemiological studies reflect altered endocrine function, our understanding of how and the extent to which PFAS disrupt the endocrine system is far from complete (Braun, 2017). Endocrine-disrupting chemicals (EDCs) are inherently challenging to study due to their unique non-monotonic dose-response relationships, transgenerational effects, organ-type- and cell-type-specific responses, and potential for long latency periods between fetal exposure and disease onset (La Merrill et al., 2020). Studies that systematically evaluate the ability of PFAS to disrupt the various cell and organ types within the endocrine system are critically needed to more accurately characterize the health impacts of PFAS exposure and elucidate the various mechanisms by which endocrine disruption can occur (Burman et al., 2020; Gore et al., 2015; White et al., 2011). Certain EDC actions of PFAS have been well-described. For example, *in silico* analysis indicated the capacity of PFAS to disrupt thyroid hormone signaling by competing with thyroid hormone thyroxine for binding to thyroid transport protein transthyretin (Weiss et al., 2009). Consistent with this mechanism, PFAS exposure levels have been linked to changes in thyroid function and altered serum levels of thyroid-stimulating hormone (TSH) in affected communities, although the magnitude of this impact has varied by study population and sex (Blake et al., 2018; Byrne et al., 2018; Inoue et al., 2019; Kim et al., 2018; Preston et al., 2018; Xiao et al., 2020; Yang et al., 2016). PFAS exposure can also affect reproductive development and reproductive hormone production, giving rise to altered pubertal timing, impaired ovarian function, and infertility (Bach et al.,

2016; Ding et al., 2020; Rappazzo et al., 2017). Exposures to PFOA, PFOS, and perfluorohexane sulfonic acid (PFHxS) have been associated with premature ovarian insufficiency and altered serum levels of estradiol (E2) and follicle-stimulating hormone (FSH) in women (Zhang et al., 2018). In agreement with these findings in humans, rodent models of PFOS exposure exhibit reduced serum E2 and progesterone levels (Feng et al., 2015). Importantly, the use of *in vivo* rodent models enabled investigators to attribute these alterations in hormone synthesis to targeted changes in both hypothalamic neurons and chromatin remodeling factors in the ovary regulating the expression of key steroidogenic enzymes, ultimately leading to impaired follicular development and ovulation. Moving forward, it is crucial to expand our understanding of how PFAS exposure can impact the endocrine system and to employ various testing models to establish the mechanisms by which these effects occur (Alofe et al., 2019).

3. Analytical methods

The scope and power of studies assessing PFAS exposure are inherently constrained by the analytical methods they employ to detect PFAS. Methods that pair liquid chromatography (LC) with tandem mass spectrometry (MS/MS) are well established for PFAS analysis in simple liquid media such as drinking water. The EPA has published multiple validated LC-MS/MS methods (i.e., Methods 537, 537.1, and 533) for the analysis of PFAS in drinking water, but has yet to do so for more complex liquid and solid matrices that are likewise important to monitor, such as serum and soil. Accurate PFAS analysis in such matrices can nevertheless be achieved by augmenting established methods with isotope dilution, appropriately tailored extraction and sample preparation procedures, and rigorous quality assurance and control measures to account for matrix effects. Most studies involving PFAS rely on targeted methods that measure tens of individual PFAS at best. Collectively, EPA Methods 537 (Shoemaker et al., 2008), 537.1 (Shoemaker and Tettenhorst, 2020), and 533 (Rosenblum and Wendelken, 2019) measure fewer than 40 compounds. Of the thousands of compounds in the PFAS class, analytical reference standards exist for fewer than 200. While non-targeted methods can be used to study much broader sets of PFAS, the resulting measurements are highly challenging to interpret. Emerging approaches in PFAS sampling and analysis present opportunities to use new data sources, broaden the range of PFAS that can be reliably measured, and predict the toxicity of PFAS that have yet to be studied in order to direct future health effects research and inform decision-makers working to develop health-protective policies.

PFAS analysis in new matrices requires investigation of suitable sample preparation and extraction methods. Advances in this area have made it possible to analyze PFAS in dried blood spots and thereby tap into new sources of prenatal PFAS exposure data. In an initial study, customized sampling, extraction, and analysis procedures were used to measure PFAS levels in dried blood spots collected through a newborn screening program in New York State between 1997 and 2010 (Spliethoff et al., 2008; Ma et al., 2013). This analytical method successfully quantified PFOS and PFOA concentrations in all blood spot samples and achieved detection limits in the ng/mL (ppb) range. While this initial investigation focused on targeted analysis of two specific compounds, its sampling and extraction procedures have more recently been adapted for analysis of an expanded list of PFAS (Kato et al., 2018; Poothong et al., 2019; Vorkamp et al., 2021) and could likely be further adapted for future use in non-targeted analyses. Given the sensitivity of the developing fetus to PFAS exposure, the ability to use archived repositories of newborn blood spots provides the opportunity to quantify exposures at a critical window of development and effectively explore the temporal relationship between PFAS exposure and onset of disease (Bell et al., 2018). This approach is expected to provide new opportunities for population-based PFAS epidemiological studies.

As a consequence of the multitude of PFAS used in commerce and of the chemical transformations they undergo in the environment, humans

are exposed to a complex mixture of PFAS that extends far beyond the compounds measured by typical targeted analytical methods. Being able to study the full complement of PFAS present in humans and environmental media is crucial, particularly given that the EPA CompTox Chemicals Dashboard contains over 9000 PFAS (EPAa) and that the health and environmental effects of the vast majority of these compounds are completely unknown. Non-targeted analyses pairing LC or gas chromatography (GC) with high-resolution tandem mass spectrometry (HRMS/MS) analyses can help meet this need and allow for more comprehensive exploration of PFAS exposure. However, processing the large volumes of mass spectrometry data generated in these analyses to identify individual PFAS that have no analytical reference standards for mass spectral matching is a challenging task that requires extensive expertise and time. To facilitate interpretation of these data, Koelmel and colleagues developed FluoroMatch, the first automated open-source software for non-targeted assignment of PFAS structures (Koelmel et al., 2020; Nason et al., 2020). This software uses PFAS libraries with over 7000 *in silico* HRMS/MS spectra to process the mass spectral output of non-targeted analyses and automatically annotate and identify PFAS. In future updates, FluoroMatch will automate intelligent data acquisition (Koelmel et al., 2017; Koelmel et al., 2020), homologous series detection, fragment screening, and prediction of transformation products and HRMS/MS spectra from proposed PFAS structures (Innovative Omics). These advances will help users not only screen for the thousands of known PFAS, but also discover new PFAS previously uncharacterized.

A publicly available, NIEHS-developed online resource addresses, at least on a cursory level, the major challenge posed by scientists' limited knowledge of the biological effects of most PFAS. Borrel and colleagues developed PFASMap, a specialized application within ChemMaps.com (<https://sandbox.ntp.niehs.nih.gov/chemmaps/>), to plot and visualize >5000 PFAS from EPA databases in three-dimensional chemical space. Within PFASMap (Fig. 1a), spatial coordinates represent chemical structural properties of the compounds, making it possible to assess their structural similarity (Borrel et al., 2018). The platform also consolidates available information on the physicochemical properties, regulatory classifications, predicted activity against endocrine pathways, and acute oral systemic toxicity data of individual PFAS, along with links to more detailed information in the EPA CompTox Chemicals Dashboard (<https://comptox.epa.gov/dashboard>). While structural similarity does not automatically equate to similarity in toxicity, some correlations undoubtedly exist, and PFASMap provides the ability to visualize and explore the characteristics of chemicals clustered near PFAS of interest (Fig. 1b). In light of the relatively small number of PFAS characterized in toxicological and epidemiological studies and of the infeasibility of studying the health effects of every member of this ever-expanding class of compounds, PFASMap serves as a valuable tool that could be used to inform risk assessment and prioritize PFAS for further study.

In the future, it would be highly beneficial to supplement complex laboratory techniques with technologies capable of rapid, sensitive PFAS detection in the field. Chromatographic techniques coupled with mass spectrometry (e.g., LC-MS/MS, GC-MS/MS) are accurate, sensitive, and increasingly powerful due to advances such as those discussed above. However, the accessibility of these technologies is limited by their high cost, and laboratory turnaround times give rise to an inevitable delay between the collection of samples and receipt of results. Field-deployable PFAS sensors that provide real-time results would enable environmental and public health professionals to immediately inform at-risk residents and thereby prevent ongoing exposures. While such sensors may not match the precision of the rigorous laboratory analyses used to investigate samples from PFAS-contaminated sites, they could be used as a screening tool to determine whether sites warrant additional, more in-depth studies. This would provide both time and cost savings. While several preliminary studies have successfully employed optical and electrochemical techniques to detect PFAS without the use of LC-MS/MS or GC-MS/MS systems (Cennamo et al., 2018; Chen et al.,

2013a; Cheng et al., 2019; Li et al., 2019; Niu et al., 2014; Ranaweera et al., 2019), the utility of these techniques is constrained by their insufficiently low limits of detection and/or their inability to detect a wide range of PFAS, i.e., beyond PFOA and PFOS. Further research is necessary to advance such technologies for timely, in-field applications.

4. Remediation and pollution prevention

To minimize human exposure, it is necessary to remediate the PFAS-contaminated media that contribute to exposure (such as soil and drinking water sources) and concurrently act to prevent future pollution. Due to the energy input required to break their carbon-fluorine bonds (Sabater et al., 2013), PFAS are resistant to many traditional degradation treatments (Dickenson and Higgins, 2016b). As such, remediation of PFAS-contaminated water currently relies on PFAS removal using established filtration technologies, i.e., granular activated carbon, ion exchange resins, and reverse osmosis (RO; CDM Smith, Inc., 2018; Dickenson and Higgins, 2016a; Flores et al., 2013; Tang et al., 2006). These approaches, although effective, generate concentrated waste in the form of spent sorbent materials, RO concentrate, and the backwashing liquid used to clean RO membranes and regenerable ion exchange resins. Without further treatment, these PFAS-rich waste streams pose a potential threat to the environment surrounding their disposal sites. Responsible waste management poses a considerable challenge for environmental officials in states where PFAS contamination requires extensive drinking water treatment and/or environmental remediation. Connecticut officials have already begun grapple with this challenge, and their waste management needs will only increase as they carry out the widespread PFAS testing recommended in the PFAS Action Plan.

Researchers have recently leveraged advances in materials science to develop new state-of-the-art technologies for PFAS removal and destruction (Duan et al., 2020; Zhang et al., 2020; Huang et al., 2020; Le et al., 2019). For example, surface-tuned nanoscale composites have shown high potential for targeted PFAS separations (Saleh et al., 2019). Their high sorption capacities, a function of their specific surface area and tunable surface chemistries, present a distinct advantage over the larger granular activated carbon and ion exchange resin sorbents currently in use. Integrating superparamagnetic properties allows for low-energy recovery from complicated environmental matrices using a magnetic field (Li et al., 2016a; Li and Fortner, 2020). Specifically, nanoscale ferrite particles (diameter 8–20 nm) can be precisely synthesized using thermal decomposition processes, which provides precise control over their size and composition and thus over their magnetic susceptibility and (super)paramagnetic properties (Li et al., 2016b). These magnetic core particles can then be surface-functionalized with specific organic surfactants that have high selectivity and sorption capacity for PFAS. In a different approach also reliant on nanoscale engineering, Huang et al. have developed a method for fabricating cost-effective single-atom catalysts designed for PFAS destruction (Huang et al., 2018). The resulting catalysts, composed of single platinum atoms anchored onto silicon carbide substrates, photocatalytically hydrodefluorinate PFOA by breaking its carbon-fluorine bonds and immobilizing the resulting fluorine through covalent bonding to the substrate. More recently, Huang et al. developed a palladium-single-atom-loaded titanium oxide (Ti₄O₇) electrode that anodically oxidizes PFOA through an electrocatalytic process (Huang et al., 2020). These new materials not only outperform benchmark performance nanomaterials, but also enable selective destruction of carbon-fluorine bonds, and could eventually provide a new treatment option for the concentrated liquid waste streams generated during PFAS filtration. Further research is required to test the effectiveness of this method for degrading additional PFAS.

Mitigation would not be necessary if PFAS were prevented from being released into the environment in the first place. Although many industrial users and manufacturers have voluntarily phased out PFOS and PFOA, most have simply replaced these compounds with shorter-chain PFAS (Wang et al., 2015b; Wang et al., 2013; Zhou et al., 2013;

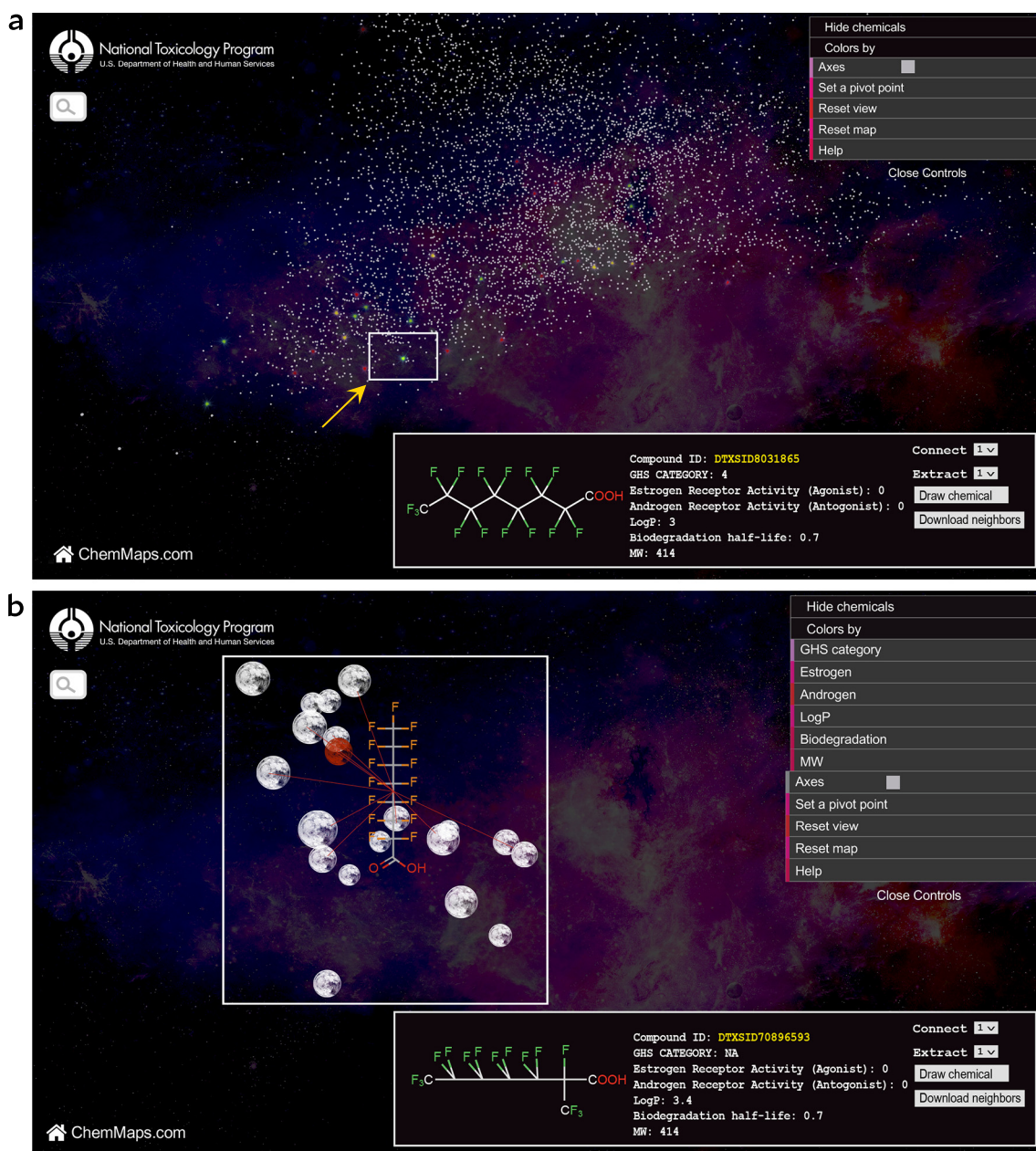


Fig. 1. PFASMap (<https://sandbox.ntp.niehs.nih.gov/chemmaps/PFASMap>). (a) PFASMap contains >5000 PFAS plotted in three-dimensional chemical space. For a selected compound (e.g., perfluorooctanoic acid, PFOA) in the map, the chemical information box (bottom right) provides a two-dimensional structure representation of the compound and user-selectable property/toxicity information. The local chemical neighborhood of the selected compound is shown in the white-outlined box (yellow arrow). (b) White-outlined box displays the 20 nearest neighbors of the selected compound, which can each be extracted and downloaded. Individual compounds within the box can be selected (e.g., red sphere, a structural analogue of PFOA) to display their basic information (bottom right). Further chemical-specific details are provided via the DTXSID link to the EPA CompTox Chemicals Dashboard (<https://comptox.epa.gov/dashboard/>). The navigation pane (upper right corner) and search bar (upper left) are also shown. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Sun et al., 2016; Wang et al., 2019b; Brendel et al., 2018; Hopkins et al., 2018) whose health risks have not been comprehensively studied (Cheng and Ng, 2018). These replacement compounds are generally less bioaccumulative than their long-chain legacy predecessors but just as persistent and even more difficult to remediate (Gagliano et al., 2020), making their continued release into the environment difficult to reverse. Green chemistry provides a framework for designing chemicals that fulfill the function and match the performance of PFAS while eliminating or minimizing hazards throughout their life cycle. This design approach aims to address the hazards associated with PFAS at each stage of their life cycle, i.e., from feedstocks and manufacturing through use and end-of-life disposal concerns (Fig. 2). The processes used to manufacture

PFAS typically rely on hydrofluoric acid, a highly hazardous chemical (Bertolini, 1992), to serve as either the direct fluorinating agent or the precursor to the fluorinating agent (Hekster et al., 2003). Hydrofluoric acid, in turn, is manufactured using sulfuric acid, another known hazard (Agency for Toxic Substances and Disease Registry, 1998). As such, its use as a feedstock poses serious risks to worker safety (Park, 2013). Moreover, PFAS themselves pose occupational safety risks. For example, in the early 2000s, the Centers for Disease Control and Prevention (CDC) found that the PFOS and PFOA blood levels of workers in PFAS manufacturing facilities were orders of magnitude higher than those of the general U.S. population (Agency for Toxic Substances and Disease Registry, 2017). These facilities have since shifted away from PFOA and

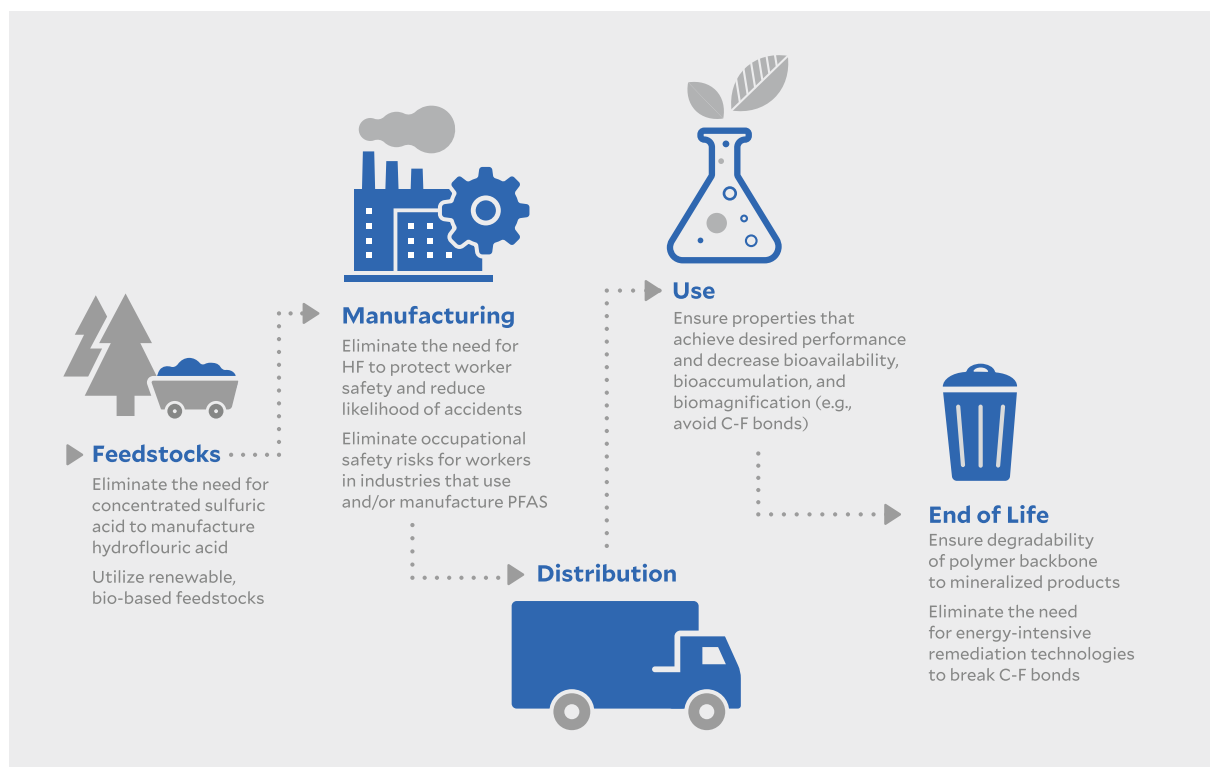


Fig. 2. Examples of the hazard reduction goals of green chemistry PFAS alternatives at each stage of the chemical life cycle. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

PFOS, but their workers (and the consumers of the products they manufacture) may now be exposed instead to shorter-chain replacement PFAS with potential health impacts that are poorly understood and persistence that is similarly problematic (Wang et al., 2019b).

Many green chemistry design approaches already show promise for the production of viable alternatives to PFAS. A variety of bio-based monomers, including stearic acid (Sharif et al., 2020), maleic acid (Yao and Tang, 2013), lactic acid (Zhang et al., 2017), and amines (Froidevaux et al., 2016), have been used as renewable feedstocks for fluorine-free polymers that could replace PFAS in flame-retardant coatings (Bourbigot and Fontaine, 2010) and in oil- and water-repellent coatings for fabric and paper (Hamdani et al., 2020; Kansal et al., 2020; Rabia et al., 2020). Ceramic-type coatings that create a barrier between a substrate and the surrounding environment (Lazar et al., 2020), typically based on silicates and alumina (Malucelli, 2016), have been demonstrated to be effective in similar coating applications (Colleoni et al., 2017; Hu et al., 2011; Liang et al., 2013; Shen et al., 2017). Biomimetic surface morphologies have been used to develop superhydrophobic surfaces that are non-fluorinated, inexpensive, mechanically strong, resistant to corrosion, and maintain water repellence despite weathering (Bhushan et al., 2009; Koch and Barthlott, 2009; Latthe et al., 2014; Lin et al., 2018; Sarkar and Saleema, 2010; Skoulas et al., 2017; Song et al., 2019; Xiu et al., 2010; Zorba et al., 2008). As many of the problems associated with PFAS stem from their persistence, degradable polymers such as biodegradable starch-based polymers are also being explored as potential replacements in water-repellence and flame-retardance applications (Albertsson and Hakkarainen, 2017; Chandra and Rustgi, 1998; Lu et al., 2009; Ma and Webster, 2018; Scott, 2002; Wu et al., 2009).

To mitigate current and future PFAS hazards, both approaches are clearly vital, i.e., to develop additional remediation technologies (especially destructive technologies) and to replace PFAS with safe and effective non-fluorinated green chemistry alternatives.

5. Fate and transport for local vulnerability assessment

In public health initiatives designed to minimize PFAS exposure, the testing of drinking water sources is a top priority. Detection of elevated PFAS concentrations in potable water enables further exposure to be prevented through remedial actions that remove PFAS or through provision of alternative water sources. However, the extent to which residents in Connecticut (and in many other states and nations) have been exposed to PFAS through drinking water ingestion is largely unknown. Public drinking water is regularly tested for numerous naturally occurring and anthropogenic contaminants, and this information is provided to CTDPH, the agency responsible for regulating the state's public drinking water systems. Between 2013 and 2015, as part of UCMR3 monitoring under the federal Safe Drinking Water Act, the EPA required large public water systems serving >10,000 people to test their finished drinking water for PFOA, PFOS, PFHxS, perfluorobutane sulfonic acid (PFBS), perfluoroheptanoic acid (PFHpA), and perfluorononanoic acid (PFNA). Of the 42 Connecticut public water systems in this category (which collectively provide drinking water to >2.3 million customers), none detected PFAS concentrations over the EPA reporting limits (EPAb). However, reporting limits at the time (20, 40, 30, 90, 10 and 20 ppt for PFOA, PFOS, PFHxS, PFBS, PFHpA, and PFNA, respectively) were higher than levels currently of concern to health officials in a number of states. In Connecticut, the current Action Level, an advisory level set by CTDPH in 2016 (CTDPHa), is 70 ppt for the summed concentrations of PFOA, PFOS, PFHxS, PFHpA, and PFNA. Continued monitoring of these large public water systems, as well as many smaller ones, is necessary to ensure that drinking water statewide is safe for human consumption. In 2018, CTDPH used its statutory authority to require (1) PFAS testing in all new sources of public drinking water and (2) public water systems serving >1000 people to evaluate their sources' vulnerability to PFAS contamination. CTDPH has also requested those water systems to test their finished drinking water for PFAS. While these initiatives made significant progress, many of the state's 2500

public drinking water systems (Fig. 3), which obtain their supplies from approximately 150 reservoir systems and 4000 groundwater sources, have not been assessed for PFAS vulnerability and have never been tested for PFAS. Moreover, nearly one quarter of Connecticut residents rely on water from an estimated 325,000 private wells (CTDPHb). This large quantity of wells presents a major challenge, and research assessing local PFAS sources and hydrogeology is necessary to prioritize the wells most vulnerable to PFAS contamination for initial rounds of testing.

Researchers at Yale are planning to study PFAS fate and transport near local landfills with the intention of developing vulnerability models for local drinking water resources and helping state officials assess the levels of PFAS exposure faced by residents. Investigation of landfills that pose potential threats to nearby residents was identified as a crucial need by regulators and researchers during the deliberations of the Inter-agency PFAS Task Force and highlighted in the resulting Action Plan. PFAS have already been detected in groundwater near two large landfills in Connecticut (Hladky, 2019), and although other landfills throughout the state are considered potential sources of PFAS contamination, few of these sites have been tested for groundwater contamination. Focusing on landfill sites prioritized by CTDEEP, Yale researchers intend to quantify the vulnerability of nearby wells using hydrologic models that simulate groundwater flow patterns and physicochemical factors governing PFAS migration away from a source. Drinking water from households with wells identified as vulnerable will be analyzed for concentrations of PFAS to enable evaluation of the models while providing immediate benefits to impacted residents through targeted interventions and focused monitoring. This work will provide a valuable model for further collaborations between university researchers and state officials to implement Action Plan recommendations and safeguard the health of Connecticut residents.

6. Community engagement

As testing for PFAS contamination in drinking water and the environment becomes more widespread, individual and community-based health concerns will require extensive research translation and risk communication by state and local leaders. Local health departments and municipal officials are often the first resources that concerned residents turn to with their questions about environmental health risks. During the development of the PFAS Action Plan, one of the primary

topics raised by stakeholders was the importance of effective communication on PFAS by state agencies. This requires officials to stay abreast of the ever-evolving science of PFAS health effects, exposure pathways, analytical methods, treatment technologies, and the impacts of these scientific advances on their communities. Because PFAS have only recently begun to gain public attention in Connecticut, residents often hear about these chemicals for the first time when they learn that contamination has been discovered locally and their households have potentially been exposed. When this occurs, it is crucial for state and local officials to be able to communicate the health risks of PFAS exposure quickly, accessibly (i.e., using culturally and linguistically appropriate formats and outreach platforms), and in a manner that does not cause unnecessary panic.

Officials' PFAS communication strategies should leverage the expertise of local health officials and community organizers to engage Connecticut communities in active partnerships that facilitate effective science translation and risk communication. These local experts understand the needs and backgrounds of their individual communities and will be able to draw upon their experience with a range of community organization techniques to actively engage residents. For example, community forums enable state and local officials to share information and learn about their constituents' understanding of the science, exposure, and risk of PFAS, which enables the officials to better tailor their science communication moving forward. Community advisory groups encourage continual education, engage a variety of stakeholders (e.g., community members, environmental advocates, research scientists, and state and local officials), and give participants a collective voice that allows their opinions to be heard. Identifying and utilizing opportunities to connect community members and environmental advocates to local scientists and to engage them in the design and implementation of research studies offers them the agency to help generate data and contribute to solutions in their own communities. Such efforts are underway in the handful of Connecticut communities that have already had to grapple with the discovery of PFAS contamination in their local environment and/or drinking water. As the State works to carry out the extensive PFAS testing recommended in the PFAS Action Plan, these isolated PFAS engagement efforts will need to be extended statewide. Implementing PFAS-focused community engagement initiatives will enhance residents' understanding of the health risks posed by PFAS and encourage their participation in the development and implementation of policies to address PFAS at the local and state levels.

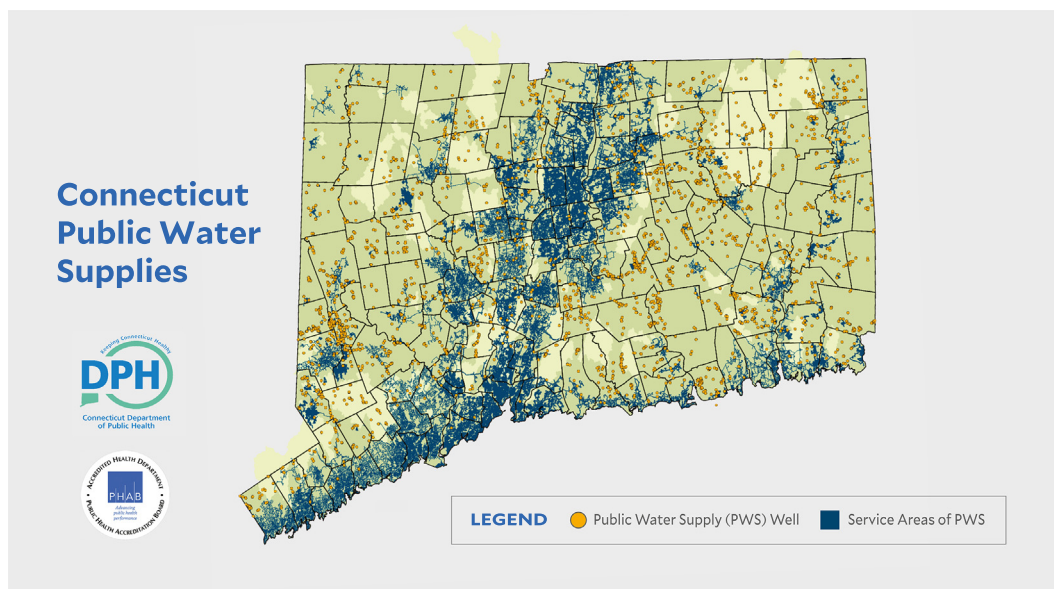


Fig. 3. Areas in Connecticut that are served by public drinking water supplies. It is presumed that outside of the service areas shown in dark blue, residents obtain drinking water from either private wells or the small public drinking water wells shown in gold. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Government researchers at the Connecticut Agricultural Experiment Station are currently collaborating with community member scientists to assess the potential of phytoremediation for removing PFAS from contaminated soil. The group is piloting this strategy at the former Loring Airforce Base, a former Superfund site in northern Maine where decades of firefighting drills and training with Class B firefighting foam (i.e., aqueous film-forming foam) contaminated the land in the vicinity of the burn house with high levels of PFAS, primarily PFOS. U.S. Air Force investigations between 2015 and 2017 measured PFOS concentrations up to 27 ng/g (ppb) in soil and 238 ng/L (ppt) in groundwater (Baker, 2018). The land is now owned by the Aroostook branch of the Micmac Nation, an indigenous people. Working with the Micmac Nation and a community organization called Upland Grassroots, a pilot test was conducted near the burn house using fiber hemp (*Cannabis sativa*), a crop that is suitable for phytoremediation due to its high water uptake, high biomass, and rapid growth. Hemp has previously been used for phytoremediation of both heavy metals (Ahmad et al., 2016) and organic contaminants (Campbell et al., 2002), although its use in the U.S. was legally restricted until recently (Smith-Heisters, 2008). Of the 19 PFAS quantified in soil at the test site, eight were taken up into hemp plants and four had significantly decreased soil concentrations at the end of the growing season. The group is currently exploring strategies to optimize hemp growth at the site and recently published their work on screening for additional PFAS in the soil (Nason et al., 2020). This work provides a model for citizen science initiatives in which members of PFAS-impacted communities can actively engage in research being conducted by scientists at local government laboratories and academic institutions.

7. Conclusion

As highlighted throughout this symposium, PFAS present unique challenges to scientists and policymakers alike. First and foremost, although the PFAS class comprises >9000 different chemicals, analytical

chemists have reference standards for fewer than 200, and extensive epidemiological and toxicological data exist for only a handful of these compounds; there are even fewer studies evaluating the EDC potential of PFAS. Many of the PFAS prevalent in the global marketplace are short-chain replacement compounds whose health effects have yet to be studied. Furthermore, while many products and industrial process employ complex mixtures of PFAS, existing studies provide scant information about the health impacts of mixed PFAS exposures. These data gaps make it challenging for health officials to accurately assess the risks posed by PFAS and to develop sufficiently protective policies. For environmental officials, management of PFAS waste presents an immense challenge due the concentrated waste generated during the remediation of contaminated water and the lack of commercially available PFAS destruction technologies and remediation methods for PFAS-contaminated environmental media, e.g., soil.

These challenges present opportunities for scientists to conduct research with meaningful real-world implications. Ongoing epidemiological and toxicological research can help fill data gaps by exploring the pathophysiological effects of exposure to PFAS mixtures and short-chain PFAS, and by characterizing the mechanisms by which adverse health effects manifest. Progress in the understanding of these mechanisms could, in turn, inform green chemistry research to produce safer alternatives to PFAS. New sampling, data acquisition, and data analysis approaches should serve as valuable resources for large-scale epidemiological studies, increase the number of PFAS that can be easily identified during testing, and facilitate the prioritization of PFAS for future health studies. New remediation technologies that efficiently degrade PFAS could help manage the waste streams generated during drinking water remediation.

Moving forward, sustained connections between researchers, government officials, and community leaders (such as those fostered by this symposium) will be invaluable as states and regions work to implement ambitious PFAS management initiatives, such as those laid out in the Connecticut PFAS Action Plan (Fig. 4). It is clear that extensive

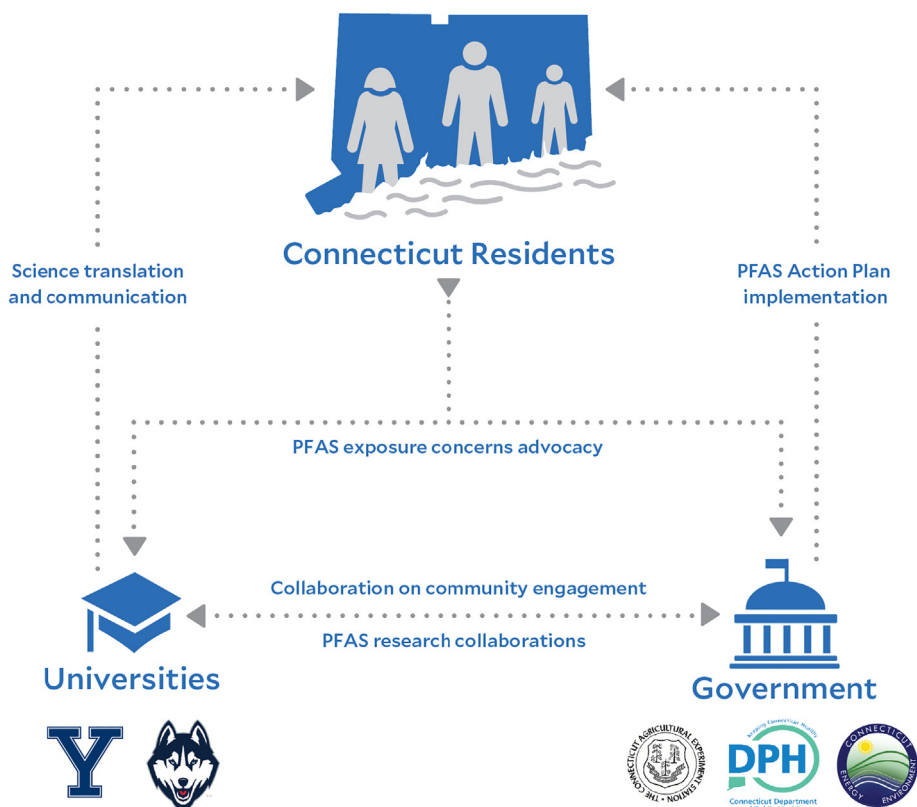


Fig. 4. Interactions between Connecticut residents, universities, and government entities that have taken place or could take place to address PFAS contamination and health concerns.

research will be necessary to assess the situation on the ground in Connecticut, i.e., to identify in-state PFAS sources and measure ambient PFAS concentrations in environmental media statewide. CTDPH efforts to identify public drinking water supplies potentially affected by PFAS contamination and CTDEEP efforts to inventory potential PFAS sources across the state provide an opportunity for a landfill-focused collaboration with Yale researchers to be broadened to apply a similar hydrological modeling approach on a larger scale. A statewide assessment of the vulnerability of community drinking water sources to PFAS contamination will help state government officials better identify and protect at-risk communities. In addition, connections with researchers will help ensure that messaging by state and local officials accurately represents the state of the science on PFAS. Collaborations between state officials, local officials, and community advocates will enhance existing communication channels and develop outreach practices that effectively meet the needs of Connecticut communities.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

The Connecticut Agricultural Experiment Station thanks the Aroostook Branch of the Micmac Nation for allowing them to study their land and Upland Grassroots for planting hemp and collecting soil samples. S.E.F. received NIEHS support (Z01E5102785). S.L.N. received support from USDA NIFA Hatch funds (CONH00789). X.M. received support from the China Scholarship Council (201906160169). S.W. received support from an Albert McKern Scholar Award.

References

Agency for Toxic Substances and Disease Registry. Toxicological profile for sulfur trioxide and sulfuric acid, Atlanta, GA, 1998.

Agency for Toxic Substances and Disease Registry, 2017. Perfluoroalkyl and Polyfluoroalkyl Substances (PFAS) in the U.S. population. https://www.atsdr.cdc.gov/pfas/docs/PFAS_in_People.pdf (accessed 8 March 2021).

Ahmad, R., Tehsin, Z., Malik, S.T., Asad, S.A., Shahzad, M., Bilal, M., et al., 2016. Phytoremediation potential of hemp (*Cannabis sativa* L.): identification and characterization of heavy metals responsive genes. *Clean – Soil, Air, Water* 44, 195–201. <https://doi.org/10.1002/clen.201500117>.

Albertsson, A.-C., Hakkarainen, M., 2017. Designed to degrade. *Science* 358, 872–873. <https://doi.org/10.1126/science.aap8115>.

Alofe, O., Kisanga, E., Inayat-Hussain, S.H., Fukumura, M., Garcia-Milian, R., Perera, L., et al., 2019. Determining the endocrine disruption potential of industrial chemicals using an integrative approach: public databases, in vitro exposure, and modeling receptor interactions. *Environ. Int.* 131, 104969. <https://doi.org/10.1016/j.envint.2019.104969>.

Andersson, E.M., Scott, K., Xu, Y., Li, Y., Olsson, D.S., Fletcher, T., et al., 2019. High exposure to perfluorinated compounds in drinking water and thyroid disease. A cohort study from Ronneby, Sweden. *Environ. Res.* 176, 108540. doi:<https://doi.org/10.1016/j.envres.2019.108540>.

Ashley-Martin, J., Dodds, L., Arbuckle, T.E., Morisset, A.-S., Fisher, M., Bouchard, M.F., et al., 2016. Maternal and neonatal levels of perfluoroalkyl substances in relation to gestational weight gain. *Int. J. Environ. Res. Public Health* 13, 146. <https://doi.org/10.3390/ijerph13010146>.

Bach, C.C., Vested, A., Jørgensen, K.T., Bonde, J.P.E., Henriksen, T.B., Toft, G., 2016. Perfluoroalkyl and polyfluoroalkyl substances and measures of human fertility: a systematic review. *Crit. Rev. Toxicol.* 46, 735–755. <https://doi.org/10.1080/10408444.2016.1182117>.

Baker, P., 2018. *Site Inspection Report for Aqueous Film Forming Foam (AFFF) Areas at Former Loring Air Force Base (Maine)*.

Ballesteros, V., Costa, O., Iniguez, C., Fletcher, T., Ballester, F., Lopez-Espinosa, M.J., 2017. Exposure to perfluoroalkyl substances and thyroid function in pregnant women and children: a systematic review of epidemiologic studies. *Environ. Int.* 99, 15–28. <https://doi.org/10.1016/j.envint.2016.10.015>.

Barry, V., Winquist, A., Steenland, K., 2013. Perfluorooctanoic acid (PFOA) exposures and incident cancers among adults living near a chemical plant. *Environ. Health Perspect.* 121, 1313–1318. <https://doi.org/10.1289/ehp.1306615>.

Bell, E.M., Yeung, E.H., Ma, W., Kannan, K., Sundaram, R., Smarr, M.M., et al., 2018. Concentrations of endocrine disrupting chemicals in newborn blood spots and infant outcomes in the upstate KIDS study. *Environ. Int.* 121, 232–239. <https://doi.org/10.1016/j.envint.2018.09.005>.

Bertolini, J.C., 1992. Hydrofluoric acid: a review of toxicity. *J. Emerg. Med.* 10, 163–168. [https://doi.org/10.1016/0736-4679\(92\)90211-B](https://doi.org/10.1016/0736-4679(92)90211-B).

Bhushan, B., Jung, Y.C., Koch, K., 2009. Micro-, nano- and hierarchical structures for superhydrophobicity, self-cleaning and low adhesion. *Philos. Trans. R. Soc. A* 367, 1631–1672. <https://doi.org/10.1098/rsta.2009.0014>.

Blake, B.E., Fenton, S.E., 2020. Early life exposure to per- and polyfluoroalkyl substances (PFAS) and latent health outcomes: a review including the placenta as a target tissue and possible driver of peri- and postnatal effects. *Toxicology* 443, 152565. <https://doi.org/10.1016/j.tox.2020.152565>.

Blake, B.E., Pinney, S.M., Hines, E.P., Fenton, S.E., Ferguson, K.K., 2018. Associations between longitudinal serum perfluoroalkyl substance (PFAS) levels and measures of thyroid hormone, kidney function, and body mass index in the Fernald Community Cohort. *Environ. Pollut.* 242, 894–904. <https://doi.org/10.1016/j.envpol.2018.07.042>.

Blake, B.E., Cope, H.A., Hall, S.M., Keys, R.D., Mahler, B.W., McCord, J., et al., 2020. Evaluation of maternal, embryo, and placental effects in CD-1 mice following gestational exposure to perfluorooctanoic acid (PFOA) or hexafluoropropylene oxide dimer acid (HFPO-DA or GenX). *Environ. Health Perspect.* 128, 027006. <https://doi.org/10.1289/EHP6233>.

Boiteux, V., Bach, C., Sagres, V., Hemard, J., Colin, A., Rosin, C., et al., 2016. Analysis of 29 per- and polyfluorinated compounds in water, sediment, soil and sludge by liquid chromatography–tandem mass spectrometry. *Int. J. Environ. An. Ch.* 96, 705–728. <https://doi.org/10.1080/03067319.2016.1196683>.

Borrel, A., Kleinstreuer, N.C., Fourches, D., 2018. Exploring drug space with ChemMaps. *com. Bioinformatics* 34, 3773–3775. <https://doi.org/10.1093/bioinformatics/bty412>.

Bourbigot, S., Fontaine, G., 2010. Flame retardancy of polylactide: an overview. *Polym. Chem.* 1, 1413–1422. <https://doi.org/10.1039/C0PY00106F>.

Braun, J.M., 2017. Early-life exposure to EDCs: role in childhood obesity and neurodevelopment. *Nat. Rev. Endocrinol.* 13, 161–173. <https://doi.org/10.1038/nrendo.2016.186>.

Brendel, S., Fetter, E., Staude, C., Vierke, L., Biegel-Engler, A., 2018. Short-chain perfluoroalkyl acids: environmental concerns and a regulatory strategy under REACH. *Environ. Sci. Eur.* 30, 9. <https://doi.org/10.1186/s12302-018-0134-4>.

Buck, R.C., Franklin, J., Berger, U., Conder, J.M., Cousins, I.T., de Voogt, P., et al., 2011. Perfluoroalkyl and polyfluoroalkyl substances in the environment: terminology, classification, and origins. *Integr. Environ. Assess. Manag.* 7, 513–541. <https://doi.org/10.1002/ieam.258>.

Buck, R.C., Murphy, P.M., Pabon, M., 2012. Chemistry, properties, and uses of commercial fluorinated surfactants. In: Knepper, T.P., Lange, F.T. (Eds.), *Polyfluorinated Chemicals and Transformation Products. The Handbook of Environmental Chemistry* vol. 17. Springer, Berlin, Heidelberg, pp. 1–24. https://doi.org/10.1007/978-3-642-21872-9_1.

Burman, A., Garcia-Milian, R., Whirlledge, S., 2020. Gene X environment: the cellular environment governs the transcriptional response to environmental chemicals. *Hum. Genomics* 14, 19. <https://doi.org/10.1186/s40246-020-00269-1>.

Byrne, S.C., Miller, P., Seguinot-Medina, S., Waghiyi, V., Buck, C.L., von Hippel, F.A., et al., 2018. Exposure to perfluoroalkyl substances and associations with serum thyroid hormones in a remote population of Alaska Natives. *Environ. Res.* 166, 537–543. <https://doi.org/10.1016/j.envres.2018.06.014>.

Campbell, S., Paquin, D., Awaya, J.D., Li, Q.X., 2002. Remediation of benzo[a]pyrene and chrysene-contaminated soil with industrial hemp (*Cannabis sativa*). *Int. J. Phytoremediation* 4, 157–168. <https://doi.org/10.1080/15226510208500080>.

Cao, W., Liu, X., Liu, X., Zhou, Y., Zhang, X., Tian, H., et al., 2018. Perfluoroalkyl substances in umbilical cord serum and gestational and postnatal growth in a Chinese birth cohort. *Environ. Int.* 116, 197–205. <https://doi.org/10.1016/j.envint.2018.04.015>.

Caron-Beaudoin, E., Ayyot, P., Laouan Sidi, E.A., Community of Lac, S., Community of Winneway - Long Point First, N., Nutashkuan, C.T.K.o., et al., 2019. Exposure to perfluoroalkyl substances (PFAS) and associations with thyroid parameters in First Nation children and youth from Quebec. *Environ. Int.* 128, 13–23. <https://doi.org/10.1016/j.envint.2019.04.029>.

CDM Smith, Inc. Advanced Treatment Options for the Northwest Water Treatment Plant. Final Report Prepared for Brunswick County Public Utilities, 2018.

Cennamo, N., D'Agostino, G., Porto, G., Biasiolo, A., Perri, C., Arcadio, F., et al., 2018. A molecularly imprinted polymer on a plasmonic plastic optical fiber to detect perfluorinated compounds in water. *Sensors* 18, 1836. <https://doi.org/10.3390/s18061836>.

Chandra, R., Rustgi, R., 1998. Biodegradable polymers. *Prog. Polym. Sci.* 23, 1273–1335. [https://doi.org/10.1016/S0079-6700\(97\)00039-7](https://doi.org/10.1016/S0079-6700(97)00039-7).

Chen, M.-H., Ha, E.-H., Wen, T.-W., Su, Y.-N., Lien, G.-W., Chen, C.-Y., et al., 2012. Perfluorinated compounds in umbilical cord blood and adverse birth outcomes. *PLoS One* 7, e42474. <https://doi.org/10.1371/journal.pone.0042474>.

Chen, L.D., Lai, C.-Z., Granda, L.P., Fierke, M.A., Mandal, D., Stein, A., et al., 2013a. Fluorous membrane ion-selective electrodes for perfluorinated surfactants: trace-level detection and in situ monitoring of adsorption. *Anal. Chem.* 85, 7471–7477. <https://doi.org/10.1021/ac401424j>.

Chen, M.-H., Ha, E.-H., Liao, H.-F., Jeng, S.-F., Su, Y.-N., Wen, T.-W., et al., 2013b. Perfluorinated compound levels in cord blood and neurodevelopment at 2 years of age. *Epidemiology* 24, 800–808. <https://doi.org/10.1097/EDE.0b013e3182a6dd46>.

Chen, W.-L., Bai, F.-Y., Chang, Y.-C., Chen, P.-C., Chen, C.-Y., 2018. Concentrations of perfluoroalkyl substances in foods and the dietary exposure among Taiwan general population and pregnant women. *J. Food Drug Anal.* 26, 994–1004. <https://doi.org/10.1016/j.jfda.2017.12.011>.

Cheng, W., Ng, C.A., 2018. Predicting relative protein affinity of novel per- and polyfluoroalkyl substances (PFASs) by an efficient molecular dynamics approach. *Environ. Sci. Technol.* 52, 7972–7980. <https://doi.org/10.1021/acs.est.8b01268>.

Cheng, Z., Dong, H., Liang, J., Zhang, F., Chen, X., Du, L., et al., 2019. Highly selective fluorescent visual detection of perfluorooctane sulfonate via blue fluorescent carbon dots and berberine chloride hydrate. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* 207, 262–269. <https://doi.org/10.1016/j.saa.2018.09.028>.

Clara, M., Scharf, S., Weiss, S., Gans, O., Scheffknecht, C., 2008. Emissions of perfluorinated alkylated substances (PFAS) from point sources—identification of relevant branches. *Water Sci. Technol.* 58, 59–66. <https://doi.org/10.2166/wst.2008.641>.

- Colleoni, C., Esposito, F., Guido, E., Migani, V., Trovato, V., Rosace, G., 2017. Ceramic coatings for water-repellent textiles. *IOP Conf. Ser. Mater. Sci. Eng.* 254, 122002. <https://doi.org/10.1088/1757-899x/254/12/122002>.
- CTDPHa. Per- and Polyfluoroalkyl Substances. <https://portal.ct.gov/DPH/Drinking-Water/DWS/Per-and-Polyfluoroalkyl-Substances> (accessed 15 February 2021).
- CTDPHb. Private Wells. <https://portal.ct.gov/DPH/Environmental-Health/Private-Well-Water-Program/Private-Wells> (accessed 29 March 2016).
- DeWitt, J.C., Blossom, S.J., Schaidler, L.A., 2019. Exposure to per-fluoroalkyl and polyfluoroalkyl substances leads to immunotoxicity: epidemiological and toxicological evidence. *J. Expo. Sci. Environ. Epidemiol.* 29, 148–156. <https://doi.org/10.1038/s41370-018-0097-y>.
- Dickenson, E.R.V., Higgins, C., 2016a. Treatment Mitigation Strategies for Poly-and Perfluoroalkyl Substances [Project # 4322].
- Dickenson, E.R.V., Higgins, C., 2016b. Treatment mitigation strategies for poly-and perfluoroalkyl substances. WRF Report 4322. Denver, CO, Water Research Foundation.
- Ding, N., Harlow, S.D., Randolph Jr., J.F., Loch-Carus, R., Park, S.K., 2020. Perfluoroalkyl and polyfluoroalkyl substances (PFAS) and their effects on the ovary. *Hum. Reprod. Update* 26, 724–752. <https://doi.org/10.1093/humupd/dmaa018>.
- Duan, L., Wang, B., Heck, K., Guo, S., Clark, C.A., Arredondo, J., et al., 2020. Efficient PFOA degradation over boron nitride. *Environ. Sci. Technol. Lett.* 7, 613–619. <https://doi.org/10.1021/acs.estlett.0c00434>.
- EPA. A review of the reference dose and reference concentration processes; Risk Assessment Forum, EPA/630/P-02/002F, 2002.
- EPA. Lifetime health advisories and health effects support documents for perfluorooctanoic acid and perfluorooctane sulfonate, EPA-HQ-OW2014-0138; FRL-9946-91-OW. Federal Register. 81, 2016, pp. 33250–33251.
- EPAa. PFAS Master List of PFAS Substances. https://comptox.epa.gov/dashboard/chemical_lists/pfasmaster (accessed 20 November 2020).
- EPAb. Occurrence Data for the Unregulated Contaminant Monitoring Rule <https://www.epa.gov/dwucmr/occurrence-data-unregulated-contaminant-monitoring-rule> (accessed 30 November 2020).
- Ernst, A., Brix, N., Lauridsen, L.L.B., Olsen, J., Parner, E.T., Liew, Z., et al., 2019. Exposure to perfluoroalkyl substances during fetal life and pubertal development in boys and girls from the Danish National Birth Cohort. *Environ. Health Perspect.* 127, 17004. <https://doi.org/10.1289/EHP3567>.
- Eryasa, B., Grandjean, P., Nielsen, F., Valvi, D., Zmirou-Navier, D., Sunderland, E., et al., 2019. Physico-chemical properties and gestational diabetes predict transplacental transfer and partitioning of perfluoroalkyl substances. *Environ. Int.* 130, 104874. <https://doi.org/10.1016/j.envint.2019.05.068>.
- Feng, X., Wang, X., Cao, X., Xia, Y., Zhou, R., Chen, L., 2015. Chronic exposure of female mice to an environmental level of perfluorooctane sulfonate suppresses estrogen synthesis through reduced histone H3K14 acetylation of the StAR promoter leading to deficits in follicular development and ovulation. *Toxicol. Sci.* 148, 368–379. <https://doi.org/10.1093/toxsci/kfv197>.
- Fenton, S.E., Ducatman, A., Boobis, A., DeWitt, J.C., Lau, C., Ng, C., et al., 2020. Per- and polyfluoroalkyl substance toxicity and human health review: current state of knowledge and strategies for informing future research. *Environ. Toxicol. Chem.* <https://doi.org/10.1002/etc.4890>.
- Flores, C., Ventura, F., Martin-Alonso, J., Caixach, J., 2013. Occurrence of perfluorooctane sulfonate (PFOS) and perfluorooctanoate (PFOA) in N.E. Spanish surface waters and their removal in a drinking water treatment plant that combines conventional and advanced treatments in parallel lines. *Sci. Total Environ.* 461–462, 618–626. <https://doi.org/10.1016/j.scitotenv.2013.05.026>.
- Froidevaux, V., Negrelli, C., Caillois, S., Pascault, J.-P., Boutevin, B., 2016. Biobased amines: from synthesis to polymers; present and future. *Chem. Rev.* 116, 14181–14224. <https://doi.org/10.1021/acs.chemrev.6b00486>.
- Gagliano, E., Sgroi, M., Falciglia, P.P., Vagliasindi, F.G.A., Roccaro, P., 2020. Removal of poly- and perfluoroalkyl substances (PFAS) from water by adsorption: role of PFAS chain length, effect of organic matter and challenges in adsorbent regeneration. *Water Res.* 171, 115381. <https://doi.org/10.1016/j.watres.2019.115381>.
- Goeden, H.M., Greene, C.W., Jacobus, J.A., 2019. A transgenerational toxicokinetic model and its use in derivation of Minnesota PFOA water guidance. *J. Expo. Sci. Environ. Epidemiol.* 29, 183–195. <https://doi.org/10.1038/s41370-018-0110-5>.
- Gore, A.C., Chappell, V.A., Fenton, S.E., Flaws, J.A., Nadal, A., Prins, G.S., et al., 2015. EDC-2: the Endocrine Society's second scientific statement on endocrine-disrupting chemicals. *Endocr. Rev.* 36, E1–E150. <https://doi.org/10.1210/er.2015-1010>.
- Grandjean, P., Andersen, E.W., Budtz-Jørgensen, E., Nielsen, F., Mølbak, K., Weihe, P., et al., 2012. Serum vaccine antibody concentrations in children exposed to perfluorinated compounds. *JAMA* 307, 391–397. <https://doi.org/10.1001/jama.2011.2034>.
- Halldorsson, T.I., Rytter, D., Haug Line, S., Bech Bodil, H., Danielsen, I., Becher, G., et al., 2012. Prenatal exposure to perfluorooctanoate and risk of overweight at 20 years of age: a prospective cohort study. *Environ. Health Perspect.* 120, 668–673. <https://doi.org/10.1289/ehp.1104034>.
- Hamdani, S.S., Li, Z., Rabnawaz, M., Kamdem, D.P., Khan, B.A., 2020. Chitosan-graft-poly (dimethylsiloxane)/zein coatings for the fabrication of environmentally friendly oil- and water-resistant paper. *ACS Sustain. Chem. Eng.* 8, 5147–5155. <https://doi.org/10.1021/acscuschemeng.9b07397>.
- Hekster, F.M., Laane, R.W.P.M., de Voogt, P., 2003. Environmental and toxicity effects of perfluoroalkylated substances. In: Ware, G. (Ed.), *Reviews of Environmental Contamination and Toxicology*. Springer vol. 179. New York, NY, pp. 99–121. https://doi.org/10.1007/0-387-21731-2_4.
- Hines, E.P., White, S.S., Stanko, J.P., Gibbs-Flournoy, E.A., Lau, C., Fenton, S.E., 2009. Phenotypic dichotomy following developmental exposure to perfluorooctanoic acid (PFOA) in female CD-1 mice: low doses induce elevated serum leptin and insulin, and overweight in mid-life. *Mol. Cell. Endocrinol.* 304, 97–105. <https://doi.org/10.1016/j.mce.2009.02.021>.
- Hladky, G.B., 2019. New tests show PFAS pollution leaking from Hartford and Ellington landfills. *Hartford Courant*, 15 August. <https://www.courant.com/news/connecticut/hc-news-pfas-landfill-worries-20190815-ozvng6kdfhhu5apowusupmsusy-story.html> (accessed 30 November 2020).
- Hopkins, Z.R., Sun, M., DeWitt, J.C., Knappe, D.R.U., 2018. Recently detected drinking water contaminants: GenX and other per- and polyfluoroalkyl ether acids. *J. AWWA* 110, 13–28. <https://doi.org/10.1002/awwa.1073>.
- Hu, Z., Chen, L., Zhao, B., Luo, Y., Wang, D.-Y., Wang, Y.-Z., 2011. A novel efficient halogen-free flame retardant system for polycarbonate. *Polym. Degrad. Stab.* 96, 320–327. <https://doi.org/10.1016/j.polymdegradstab.2010.03.005>.
- Hu, X.C., Andrews, D.Q., Lindstrom, A.B., Bruton, T.A., Schaidler, L.A., Grandjean, P., et al., 2016. Detection of poly- and perfluoroalkyl substances (PFASs) in U.S. drinking water linked to industrial sites, military fire training areas, and wastewater treatment plants. *Environ. Sci. Technol. Lett.* 3, 344–350. <https://doi.org/10.1021/acs.estlett.6b00260>.
- Huang, D., de Vera, G.A., Chu, C., Zhu, Q., Stavitski, E., Mao, J., et al., 2018. Single-atom Pt catalyst for effective C–F bond activation via hydrodefluorination. *ACS Catal.* 8, 9353–9358. <https://doi.org/10.1021/acscatal.8b02660>.
- Huang, R., Chen, Q., Zhang, L., Luo, K., Chen, L., Zhao, S., et al., 2019. Prenatal exposure to perfluoroalkyl and polyfluoroalkyl substances and the risk of hypertensive disorders of pregnancy. *Environ. Health* 18, 5. <https://doi.org/10.1186/s12940-018-0445-3>.
- Huang, D., Wang, K., Niu, J., Chu, C., Weon, S., Zhu, Q., et al., 2020. Amorphous Pd-loaded Ti4O7 electrode for direct anodic destruction of perfluorooctanoic acid. *Environ. Sci. Technol.* 54, 10954–10963. <https://doi.org/10.1021/acs.est.0c03800>.
- Innovative Omics. FluoroMatch – Covers entire non-targeted PFAS workflow. <https://innovativeomics.com/software/fluoromatch-flow-covers-entire-pfas-workflow/> (accessed 4 December 2020).
- Inoue, K., Ritz, B., Andersen Stine, L., Ramlau-Hansen Cecilia, H., Høyer Birgit, B., Bech Bodil, H., et al., 2019. Perfluoroalkyl substances and maternal thyroid hormones in early pregnancy; findings in the Danish National Birth Cohort. *Environ. Health Perspect.* 127, 117002. <https://doi.org/10.1289/EHP5482>.
- Johnson, P.I., Sutton, P., Atchley Dylan, S., Koustas, E., Lam, J., Sen, S., et al., 2014. The navigation guide—evidence-based medicine meets environmental health: systematic review of human evidence for PFOA effects on fetal growth. *Environ. Health Perspect.* 122, 1028–1039. <https://doi.org/10.1289/ehp.1307893>.
- Kansal, D., Hamdani, S.S., Ping, R., Rabnawaz, M., 2020. Starch and zein biopolymers as a sustainable replacement for PFAS, silicone oil, and plastic-coated paper. *Ind. Eng. Chem. Res.* 59, 12075–12084. <https://doi.org/10.1021/acs.iecr.0c01291>.
- Karlsen, M., Grandjean, P., Weihe, P., Steuerwald, U., Oulhote, Y., Valvi, D., 2017. Early-life exposures to persistent organic pollutants in relation to overweight in preschool children. *Reprod. Toxicol.* 68, 145–153. <https://doi.org/10.1016/j.reprotox.2016.08.002>.
- Kato, K., Kalathil, A.A., Patel, A.M., Ye, X., Calafat, A.M., 2018. Per- and polyfluoroalkyl substances and fluorinated alternatives in urine and serum by on-line solid phase extraction–liquid chromatography–tandem mass spectrometry. *Chemosphere* 209, 338–345. <https://doi.org/10.1016/j.chemosphere.2018.06.085>.
- Kim, M.J., Moon, S., Oh, B.C., Jung, D., Ji, K., Choi, K., et al., 2018. Association between perfluoroalkyl substances exposure and thyroid function in adults: a meta-analysis. *PLoS One* 13, e0197244. <https://doi.org/10.1371/journal.pone.0197244>.
- Koch, K., Barthlott, W., 2009. Superhydrophobic and superhydrophilic plant surfaces: an inspiration for biomimetic materials. *Philos. Trans. R. Soc. A* 367, 1487–1509. <https://doi.org/10.1098/rsta.2009.0022>.
- Koelmel, J.P., Kroeger, N.M., Gill, E.L., Ulmer, C.Z., Bowden, J.A., Patterson, R.E., et al., 2017. Expanding lipidome coverage using LC-MS/MS data-dependent acquisition with automated acquisition list generation. *J. Am. Soc. Mass Spectrom.* 28, 908–917. <https://doi.org/10.1007/s13361-017-1608-0>.
- Koelmel, J.P., Paige, M.K., Aristizabal-Henao, J.J., Robey, N.M., Nason, S.L., Stelben, P.J., et al., 2020. Toward comprehensive per- and polyfluoroalkyl substances snnotation using FluoroMatch software and intelligent high-resolution tandem mass spectrometry acquisition. *Anal. Chem.* 92, 11186–11194. <https://doi.org/10.1021/acs.analchem.0c01591>.
- Koustas, E., Lam, J., Sutton, P., Johnson Paula, I., Atchley Dylan, S., Sen, S., et al., 2014. The navigation guide—evidence-based medicine meets environmental health: systematic review of nonhuman evidence for PFOA effects on fetal growth. *Environ. Health Perspect.* 122, 1015–1027. <https://doi.org/10.1289/ehp.1307177>.
- La Merrill, M.A., Vandenberg, L.N., Smith, M.T., Goodson, W., Browne, P., Patisaul, H.B., et al., 2020. Consensus on the key characteristics of endocrine-disrupting chemicals as a basis for hazard identification. *Nat. Rev. Endocrinol.* 16, 45–57. <https://doi.org/10.1038/s41574-019-0273-8>.
- Land, M., de Wit, C.A., Bignert, A., Cousins, I.T., Herzke, D., Johansson, J.H., et al., 2018. What is the effect of phasing out long-chain per- and polyfluoroalkyl substances on the concentrations of perfluoroalkyl acids and their precursors in the environment? A systematic review. *Environ. Evid.* 7, 4. <https://doi.org/10.1186/s13750-017-0114-y>.
- Lathe, S.S., Terashima, C., Nakata, K., Fujishima, A., 2014. Superhydrophobic surfaces developed by mimicking hierarchical surface morphology of lotus leaf. *Molecules* 19, 4256–4283. <https://doi.org/10.3390/molecules19044256>.
- Lazar, S.T., Kolibaba, T.J., Grunlan, J.C., 2020. Flame-retardant surface treatments. *Nat. Rev. Mater.* 5, 259–275. <https://doi.org/10.1038/s41578-019-0164-6>.
- Le, T.X.H., Haflich, H., Shah, A.D., Chaplin, B.P., 2019. Energy-efficient electrochemical oxidation of perfluoroalkyl substances using a Ti4O7 reactive electrochemical membrane anode. *Environ. Sci. Technol. Lett.* 6, 504–510. <https://doi.org/10.1021/acs.estlett.9b00397>.
- Li, W., Fortner, J., 2020. (Super)paramagnetic nanoparticles as platform materials for environmental applications: From synthesis to demonstration. *Front. Env. Sci. Eng.* 14, 77. <https://doi.org/10.1007/s11783-020-1256-7>.
- Li, W., Mayo, J., Benoit, D., Troyer, L., Lewicka, Z., Lafferty, B., et al., 2016a. Engineered superparamagnetic iron oxide nanoparticles for ultra-enhanced uranium separation and sensing. *J. Mater. Chem. A* 4, 15022–15029. <https://doi.org/10.1039/c6ta04709b>.

- Li, W., Lee, S., Wu, J., Hinton, C., Fortner, J., 2016b. Shape and size controlled synthesis of uniform iron oxide nanocrystals through new non-hydrolytic routes. *Nanotechnology* 27, 324002. <https://doi.org/10.1088/0957-4484/27/32/324002>.
- Li, M., Zeng, X.-W., Qian, X., Vaughn, M.G., Sauv e, S., Paul, G., et al., 2017. Isomers of perfluorooctanesulfonate (PFOS) in cord serum and birth outcomes in China: Guangzhou Birth Cohort Study. *Environ. Int.* 102, 1–8. <https://doi.org/10.1016/j.envint.2017.03.006>.
- Li, J., Zhang, C., Yin, M., Zhang, Z., Chen, Y., Deng, Q., et al., 2019. Surfactant-sensitized covalent organic frameworks-functionalized lanthanide-doped nanocrystals: an ultrasensitive sensing platform for perfluorooctane sulfonate. *ACS Omega* 4, 15947–15955. <https://doi.org/10.1021/acsomega.9b01996>.
- Liang, S., Neisius, N.M., Gaan, S., 2013. Recent developments in flame retardant polymeric coatings. *Prog. Org. Coat.* 76, 1642–1665. <https://doi.org/10.1016/j.porgcoat.2013.07.014>.
- Lien, G.-W., Huang, C.-C., Shiu, J.-S., Chen, M.-H., Hsieh, W.-S., Guo, Y.-L., et al., 2016. Perfluoroalkyl substances in cord blood and attention deficit/hyperactivity disorder symptoms in seven-year-old children. *Chemosphere* 156, 118–127. <https://doi.org/10.1016/j.chemosphere.2016.04.102>.
- Liew, Z., Ritz, B., Bonfeld-J rgensen, E.C., Henriksen, T.B., Nohr, E.A., Bech, B.H., et al., 2014. Prenatal exposure to perfluoroalkyl substances and the risk of congenital cerebral palsy in children. *Am. J. Epidemiol.* 180, 574–581. <https://doi.org/10.1093/aje/kwu179>.
- Liew, Z., Ritz, B., von Ehrenstein, O.S., Bech, B.H., Nohr, E.A., Fei, C., et al., 2015. Attention deficit/hyperactivity disorder and childhood autism in association with prenatal exposure to perfluoroalkyl substances: a nested case-control study in the Danish National Birth Cohort. *Environ. Health Perspect.* 123, 367–373. <https://doi.org/10.1289/ehp.1408412>.
- Liew, Z., Ritz, B., Bach, C.C., Asarnow, R.F., Bech, B.H., Nohr, E.A., et al., 2018. Prenatal exposure to perfluoroalkyl substances and IQ scores at age 5; a study in the Danish National Birth Cohort. *Environ. Health Perspect.* 126, 067004. <https://doi.org/10.1289/EHP2754>.
- Liew, Z., Luo, J., Nohr, E.A., Bech, B.H., Bossi, R., Arah, O.A., et al., 2020. Maternal plasma perfluoroalkyl substances and miscarriage: a nested case-control study in the Danish National Birth Cohort. *Environ. Health Perspect.* 128, 47007. <https://doi.org/10.1289/EHP6202>.
- Lin, Y., Han, J., Cai, M., Liu, W., Luo, X., Zhang, H., et al., 2018. Durable and robust transparent superhydrophobic glass surfaces fabricated by a femtosecond laser with exceptional water repellency and thermostability. *J. Mater. Chem. A* 6, 9049–9056. <https://doi.org/10.1039/C8TA01965G>.
- Lin, P.D., Cardenas, A., Hauser, R., Gold, D.R., Kleinman, K.P., Hivert, M.F., et al., 2019. Per- and polyfluoroalkyl substances and blood lipid levels in pre-diabetic adults—longitudinal analysis of the diabetes prevention program outcomes study. *Environ. Int.* 129, 343–353. <https://doi.org/10.1016/j.envint.2019.05.027>.
- Lu, D., Xiao, C., Xu, S., 2009. Starch-based completely biodegradable polymer materials. *Express Polym Lett* 3, 366–375. <https://doi.org/10.3144/expresspolymlett.2009.46>.
- Ma, S., Webster, D.C., 2018. Degradable thermosets based on labile bonds or linkages: a review. *Prog. Polym. Sci.* 76, 65–110. <https://doi.org/10.1016/j.progpolymsci.2017.07.008>.
- Ma, W., Kannan, K., Wu, Q., Bell, E.M., Druschel, C.M., Caggana, M., et al., 2013. Analysis of polyfluoroalkyl substances and bisphenol A in dried blood spots by liquid chromatography tandem mass spectrometry. *Anal. Bioanal. Chem.* 405, 4127–4138. <https://doi.org/10.1007/s00216-013-6787-3>.
- Malucelli, G., 2016. Surface-engineered fire protective coatings for fabrics through sol-gel and layer-by-layer methods: an overview. *Coatings* 6, 33. <https://doi.org/10.3390/coatings6030033>.
- Mamsen, L.S., Bj rsvang, R.D., Mucs, D., Vinnars, M.T., Papadogiannakis, N., Lindh, C.H., et al., 2019. Concentrations of perfluoroalkyl substances (PFASs) in human embryonic and fetal organs from first, second, and third trimesters. *Environ. Int.* 124, 482–492. <https://doi.org/10.1016/j.envint.2019.01.010>.
- Meng, Q., Inoue, K., Ritz, B., Olsen, J., Liew, Z., 2018. Prenatal exposure to perfluoroalkyl substances and birth outcomes; an updated analysis from the Danish National Birth Cohort. *Int. J. Environ. Res. Public Health* 15, 1832. <https://doi.org/10.3390/ijerph15091832>.
- Mussabek, D., Ahrens, L., Persson, K.M., Berndtsson, R., 2019. Temporal trends and sediment-water partitioning of per- and polyfluoroalkyl substances (PFAS) in lake sediment. *Chemosphere* 227, 624–629. <https://doi.org/10.1016/j.chemosphere.2019.04.074>.
- Nason, S.L., Koelmel, J., Zuverza-Mena, N., Stanley, C., Tamez, C., Bowden, J.A., et al., 2020. Software comparison for non-targeted analysis of PFAS in AFFF-contaminated soil. *J. Am. Soc. Mass Spectrom.* <https://pubs.acs.org/doi/10.1021/jasms.0c00261>
- Nelson, J.W., Hatch, E.E., Webster, T.F., 2010. Exposure to polyfluoroalkyl chemicals and cholesterol, body weight, and insulin resistance in the general U.S. population. *Environ. Health Perspect.* 118, 197–202. <https://doi.org/10.1289/ehp.0901165>.
- Niu, H., Wang, S., Zhou, Z., Ma, Y., Ma, X., Cai, Y., 2014. Sensitive colorimetric visualization of perfluorinated compounds using poly(ethylene glycol) and perfluorinated thiols modified gold nanoparticles. *Anal. Chem.* 86, 4170–4177. <https://doi.org/10.1021/ac403406d>.
- Oulhote, Y., Steuerwald, U., Debes, F., Weihe, P., Grandjean, P., 2016. Behavioral difficulties in 7-year old children in relation to developmental exposure to perfluorinated alkyl substances. *Environ. Int.* 97, 237–245. <https://doi.org/10.1016/j.envint.2016.09.015>.
- Oulhote, Y., Huttenhower, C., Valvi, D., Mallick, H., Lloyd-Price, J., Rahnavard, G., et al., 2019. Human microbiome and lifetime exposure to environmental chemicals in healthy young adults. *Environ. Epidemiol.* 3, 298–299. <https://doi.org/10.1097/01.EE9.0000609236.60775.5e>.
- Park, S.B., 2013. Alert over South Korea toxic leaks: government moves to tighten oversight after string of hydrogen fluoride accidents. *Nature* 494, 15–17. <https://doi.org/10.1038/494015a>.
- Poonthong, S., Papadopoulou, E., Lundanes, E., Padilla-S nchez, J.A., Thomsen, C., Haug, L.S., 2019. Dried blood spots for reliable biomonitoring of poly- and perfluoroalkyl substances (PFASs). *Sci. Total Environ.* 655, 1420–1426. <https://doi.org/10.1016/j.scitotenv.2018.11.214>.
- Post, G.B., 2020. Recent US state and federal drinking water guidelines for per- and polyfluoroalkyl substances. *Environ. Toxicol. Chem.* <https://doi.org/10.1002/etc.4863>.
- Preston, E.V., Webster, T.F., Oken, E., Claus Henn, B., McClean, M.D., Rifas-Shiman, S.L., et al., 2018. Maternal plasma per- and polyfluoroalkyl substance concentrations in early pregnancy and maternal and neonatal thyroid function in a prospective birth cohort: Project Viva (USA). *Environ. Health Perspect.* 126, 027013. <https://doi.org/10.1289/EHP2534>.
- Rabia, S., Muhammad, M., Naveed, R., Waqas, A.S., Qutab, H.G., 2020. Development of free fluorine and formaldehyde oil and water repellent finishes for cotton fabrics through polymerization of bio-based stearic acid with carboxylic acids. *Ind. Textila* 71, 145–155. <https://doi.org/10.35530/IT.071.02.1731>.
- Ranaweera, R., Ghafari, C., Luo, L., 2019. Bubble-nucleation-based method for the selective and sensitive electrochemical detection of surfactants. *Anal. Chem.* 91, 7744–7748. <https://doi.org/10.1021/acs.analchem.9b01060>.
- Rappazzo, K.M., Coffman, E., Hines, E.P., 2017. Exposure to perfluorinated alkyl substances and health outcomes in children: a systematic review of the epidemiologic literature. *Int. J. Environ. Res. Public Health* 14, 691. <https://doi.org/10.3390/ijerph14070691>.
- Romano, M.E., Xu, Y., Calafat, A.M., Yolton, K., Chen, A., Webster, G.M., et al., 2016. Maternal serum perfluoroalkyl substances during pregnancy and duration of breastfeeding. *Environ. Res.* 149, 239–246. <https://doi.org/10.1016/j.envres.2016.04.034>.
- Rosenblum, L., Wendelken, S.C., 2019. Method 533: Determination of Per- and Polyfluoroalkyl Substances in Drinking Water by Isotope Dilution Anion Exchange Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry. U.S. Environmental Protection Agency, Washington, D.C.
- Sabater, S., Mata, J.A., Peris, E., 2013. Hydrodefluorination of carbon-fluorine bonds by the synergistic action of a ruthenium-palladium catalyst. *Nat. Commun.* 4, 2553. <https://doi.org/10.1038/ncomms3553>.
- Saleh, N., Khalid, A., Tian, Y., Ayres, C., Sabaraya, I., Pietari, J., et al., 2019. Removal of poly- and per-fluoroalkyl substances from aqueous systems by nano-enabled water treatment strategies. *Environ. Sci.-Wat. Res.* 5, 198–208. <https://doi.org/10.1039/c8ew00621k>.
- Sarkar, D.K., Saleema, N., 2010. One-step fabrication process of superhydrophobic green coatings. *Surf. Coat. Technol.* 204, 2483–2486. <https://doi.org/10.1016/j.surfcoat.2010.01.033>.
- Scott, G., 2002. Why degradable polymers?, in: Scott, G. (Ed.), *Degradable Polymers: Principles and Applications*. Springer, Dordrecht, pp. 1–15. https://doi.org/10.1007/978-94-017-1217-0_1.
- Sharif, R., Mohsin, M., Ramzan, N., Ahmad, S.W., Qutab, H.G., 2020. Synthesis and application of fluorine-free environment-friendly stearic acid-based oil and water repellent for cotton fabric. *J. Nat. Fibers*, 1–16. <https://doi.org/10.1080/15440478.2020.1787918>.
- Shearer, J.J., Callahan, C.L., Calafat, A.M., Huang, W.Y., Jones, R.R., Sabbiseti, V.S., et al., 2020. Serum concentrations of per- and polyfluoroalkyl substances and risk of renal cell carcinoma. *J. Natl. Cancer Inst.* <https://doi.org/10.1093/jnci/djaa143>.
- Shen, K., Yu, M., Li, Q., Sun, W., Zhang, X., Quan, M., et al., 2017. Synthesis of a fluorine-free polymeric water-repellent agent for creation of superhydrophobic fabrics. *Appl. Surf. Sci.* 426, 694–703. <https://doi.org/10.1016/j.apsusc.2017.07.245>.
- Shi, Y., Yang, L., Li, J., Lai, J., Wang, Y., Zhao, Y., et al., 2017. Occurrence of perfluoroalkyl substances in cord serum and association with growth indicators in newborns from Beijing. *Chemosphere* 169, 396–402. <https://doi.org/10.1016/j.chemosphere.2016.11.050>.
- Shoemaker, J., Tettenhorst, D. Method 537.1: Determination of selected per- and polyfluorinated alkyl substances in drinking water by solid phase extraction and liquid chromatography/tandem mass spectrometry (LC/MS/MS). U.S. Environmental Protection Agency, Washington, D.C., 2020.
- Shoemaker, J.A., Grimmett, P., Boutin, B., 2008. Determination of Selected Perfluorinated Alkyl Acids in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS). U.S. Environmental Protection Agency, Washington, D.C.
- Skoulas, E., Manousaki, A., Fotakis, C., Stratakis, E., 2017. Biomimetic surface structuring using cylindrical vector femtosecond laser beams. *Sci. Rep.* 7, 45114. <https://doi.org/10.1038/srep45114>.
- Smith-Heisters, S., 2008. Environmental costs of hemp prohibition in the United States. *Journal of Industrial Hemp* 13, 157–170. <https://doi.org/10.1080/15377880802391308>.
- Song, J., Li, Y., Xu, W., Liu, H., Lu, Y., 2019. Inexpensive and non-fluorinated superhydrophobic concrete coating for anti-icing and anti-corrosion. *J. Colloid Interf. Sci.* 541, 86–92. <https://doi.org/10.1016/j.jcis.2019.01.014>.
- Splithoff, H.M., Tao, L., Shaver, S.M., Aldous, K.M., Pass, K.A., Kannan, K., et al., 2008. Use of newborn screening program blood spots for exposure assessment: declining levels of perfluorinated compounds in New York State infants. *Environ. Sci. Technol.* 42, 5361–5367. <https://doi.org/10.1021/es8006244>.
- Starling, A.P., Engel, S.M., Richardson, D.B., Baird, D.D., Haug, L.S., Stuebe, A.M., et al., 2014. Perfluoroalkyl substances during pregnancy and validated preeclampsia among nulliparous women in the Norwegian Mother and Child Cohort Study. *Am. J. Epidemiol.* 179, 824–833. <https://doi.org/10.1093/aje/kwt432>.
- Steenland, K., Zhao, L., Winquist, A., Parks, C., 2013. Ulcerative colitis and perfluorooctanoic acid (PFOA) in a highly exposed population of community residents and workers in the mid-Ohio valley. *Environ. Health Perspect.* 121, 900–905. <https://doi.org/10.1289/ehp.1206449>.
- Steenland, K., Kugathasan, S., Barr, D.B., 2018. PFOA and ulcerative colitis. *Environ. Res.* 165, 317–321. <https://doi.org/10.1016/j.envres.2018.05.007>.
- Sun, M., Arevalo, E., Strynar, M., Lindstrom, A., Richardson, M., Kearns, B., et al., 2016. Legacy and emerging perfluoroalkyl substances are important drinking water contaminants in the Cape Fear River watershed of North Carolina. *Environ. Sci. Technol. Lett.* 3, 415–419. <https://doi.org/10.1021/acs.estlett.6b00398>.
- Sunderland, E.M., Hu, X.C., Dassuncao, C., Tokranov, A.K., Wagner, C.C., Allen, J.G., 2019. A review of the pathways of human exposure to poly- and perfluoroalkyl substances

- (PFASs) and present understanding of health effects. *J. Expo. Sci. Environ. Epidemiol.* 29, 131–147. <https://doi.org/10.1038/s41370-018-0094-1>.
- Tang, C.Y., Fu, Q.S., Robertson, A.P., Criddle, C.S., Leckie, J.O., 2006. Use of reverse osmosis membranes to remove perfluorooctane sulfonate (PFOS) from semiconductor wastewater. *Environ. Sci. Technol.* 40, 7343–7349. <https://doi.org/10.1021/es060831q>.
- The Connecticut Interagency PFAS Task Force, 2019. PFAS Action Plan. <https://portal.ct.gov/-/media/Office-of-the-Governor/News/20191101-CT-Interagency-PFAS-Task-Force-Action-Plan.pdf>.
- Timmermann, C.A.G., Budtz-Jørgensen, E., Petersen, M.S., Weihe, P., Steuerwald, U., Nielsen, F., et al., 2017. Shorter duration of breastfeeding at elevated exposures to perfluoroalkyl substances. *Reprod. Toxicol.* 68, 164–170. <https://doi.org/10.1016/j.reprotox.2016.07.010>.
- Vorkamp, K., Castaño, A., Antignac, J.P., Boada, L.D., Cequier, E., Covaci, A., et al., 2021. Biomarkers, matrices and analytical methods targeting human exposure to chemicals selected for a European human biomonitoring initiative. *Environ. Int.* 146, 106082. <https://doi.org/10.1016/j.envint.2020.106082>.
- Wang, Z., Cousins, I.T., Scheringer, M., Hungerbühler, K., 2013. Fluorinated alternatives to long-chain perfluoroalkyl carboxylic acids (PFCAs), perfluoroalkane sulfonic acids (PFASs) and their potential precursors. *Environ. Int.* 60, 242–248. <https://doi.org/10.1016/j.envint.2013.08.021>.
- Wang, Y., Rogan, W.J., Chen, H.-Y., Chen, P.-C., Su, P.-H., Chen, H.-Y., et al., 2015a. Prenatal exposure to perfluoroalkyl substances and children's IQ: the Taiwan Maternal and Infant Cohort Study. *Int. J. Hyg. Environ. Health* 218, 639–644. <https://doi.org/10.1016/j.ijheh.2015.07.002>.
- Wang, Z., Cousins, I.T., Scheringer, M., Hungerbühler, K., 2015b. Hazard assessment of fluorinated alternatives to long-chain perfluoroalkyl acids (PFAAs) and their precursors: status quo, ongoing challenges and possible solutions. *Environ. Int.* 75, 172–179. <https://doi.org/10.1016/j.envint.2014.11.013>.
- Wang, B., Chen, Q., Shen, L., Zhao, S., Pang, W., Zhang, J., 2016a. Perfluoroalkyl and polyfluoroalkyl substances in cord blood of newborns in Shanghai, China: implications for risk assessment. *Environ. Int.* 97, 7–14. <https://doi.org/10.1016/j.envint.2016.10.008>.
- Wang, Y., Adgent, M., Su, P.-H., Chen, H.-Y., Chen, P.-C., Hsiung Chao, A., et al., 2016b. Prenatal exposure to perfluorocarboxylic acids (PFCAs) and fetal and postnatal growth in the Taiwan Maternal and Infant Cohort Study. *Environ. Health Perspect.* 124, 1794–1800. <https://doi.org/10.1289/ehp.1509998>.
- Wang, H., Yang, J., Du, H., Xu, L., Liu, S., Yi, J., et al., 2018a. Perfluoroalkyl substances, glucose homeostasis, and gestational diabetes mellitus in Chinese pregnant women: a repeat measurement-based prospective study. *Environ. Int.* 114, 12–20. <https://doi.org/10.1016/j.envint.2018.01.027>.
- Wang, Y., Zhang, L., Teng, Y., Zhang, J., Yang, L., Li, J., et al., 2018b. Association of serum levels of perfluoroalkyl substances with gestational diabetes mellitus and postpartum blood glucose. *J. Environ. Sci. (China)* 69, 5–11. <https://doi.org/10.1016/j.jes.2018.03.016>.
- Wang, H., Du, H., Yang, J., Jiang, H., 2019a. PFOS, PFOA, estrogen homeostasis, and birth size in Chinese infants. *Chemosphere* 221, 349–355. <https://doi.org/10.1016/j.chemosphere.2019.01.061>.
- Wang, Y., Chang, W., Wang, L., Zhang, Y., Zhang, Y., Wang, M., et al., 2019b. A review of sources, multimedia distribution and health risks of novel fluorinated alternatives. *Ecotoxicol. Environ. Saf.* 182, 109402. <https://doi.org/10.1016/j.ecoenv.2019.109402>.
- Weiss, J.M., Andersson, P.L., Lamoree, M.H., Leonards, P.E., van Leeuwen, S.P., Hamers, T., 2009. Competitive binding of poly- and perfluorinated compounds to the thyroid hormone transport protein transthyretin. *Toxicol. Sci.* 109, 206–216. <https://doi.org/10.1093/toxsci/kfp055>.
- White, S.S., Calafat, A.M., Kuklenyik, Z., Villanueva, L., Zehr, R.D., Helfant, L., et al., 2007. Gestational PFOA exposure of mice is associated with altered mammary gland development in dams and female offspring. *Toxicol. Sci.* 96, 133–144. <https://doi.org/10.1093/toxsci/kfl177>.
- White, S.S., Fenton, S.E., Hines, E.P., 2011. Endocrine disrupting properties of perfluorooctanoic acid. *J. Steroid Biochem. Mol. Biol.* 127, 16–26. <https://doi.org/10.1016/j.jsbmb.2011.03.011>.
- Wu, K., Hu, Y., Song, L., Lu, H., Wang, Z., 2009. Flame retardancy and thermal degradation of intumescent flame retardant starch-based biodegradable composites. *Ind. Eng. Chem. Res.* 48, 3150–3157. <https://doi.org/10.1021/ie801230h>.
- Wu, K., Xu, X., Peng, L., Liu, J., Guo, Y., Huo, X., 2012. Association between maternal exposure to perfluorooctanoic acid (PFOA) from electronic waste recycling and neonatal health outcomes. *Environ. Int.* 48, 1–8. <https://doi.org/10.1016/j.envint.2012.06.018>.
- Xiao, C., Grandjean, P., Valvi, D., Nielsen, F., Jensen, T.K., Weihe, P., et al., 2020. Associations of exposure to perfluoroalkyl substances with thyroid hormone concentrations and birth size. *J. Clin. Endocrinol. Metab.* 105, 735–745. <https://doi.org/10.1210/clinem/dgz147>.
- Xiu, Y., Liu, Y., Hess, D.W., Wong, C.P., 2010. Mechanically robust superhydrophobicity on hierarchically structured Si surfaces. *Nanotechnology* 21, 155705. <https://doi.org/10.1088/0957-4484/21/15/155705>.
- Yang, L., Li, J., Lai, J., Luan, H., Cai, Z., Wang, Y., et al., 2016. Placental transfer of perfluoroalkyl substances and associations with thyroid hormones: Beijing prenatal exposure study. *Sci. Rep.* 6, 21699. <https://doi.org/10.1038/srep21699>.
- Yao, K., Tang, C., 2013. Controlled polymerization of next-generation renewable monomers and beyond. *Macromolecules* 46, 1689–1712. <https://doi.org/10.1021/ma3019574>.
- Yao, Q., Shi, R., Wang, C., Han, W., Gao, Y., Zhang, Y., et al., 2019. Cord blood per- and polyfluoroalkyl substances, placental steroidogenic enzyme, and cord blood reproductive hormone. *Environ. Int.* 129, 573–582. <https://doi.org/10.1016/j.envint.2019.03.047>.
- Zhang, C., Garrison, T.F., Madbouly, S.A., Kessler, M.R., 2017. Recent advances in vegetable oil-based polymers and their composites. *Prog. Polym. Sci.* 71, 91–143. <https://doi.org/10.1016/j.progpolymsci.2016.12.009>.
- Zhang, S., Tan, R., Pan, R., Xiong, J., Tian, Y., Wu, J., et al., 2018. Association of perfluoroalkyl and polyfluoroalkyl substances with premature ovarian insufficiency in Chinese women. *J. Clin. Endocrinol. Metab.* 103, 2543–2551. <https://doi.org/10.1210/jc.2017-02783>.
- Zhang, W., Efstathiadis, H., Li, L., Liang, Y., 2020. Environmental factors affecting degradation of perfluorooctanoic acid (PFOA) by In₂O₃ nanoparticles. *J. Environ. Sci.* 93, 48–56. <https://doi.org/10.1016/j.jes.2020.02.028>.
- Zhou, Z., Liang, Y., Shi, Y., Xu, L., Cai, Y., 2013. Occurrence and transport of perfluoroalkyl acids (PFAAs), including short-chain PFAAs in Tangxun Lake, China. *Environ. Sci. Technol.* 47, 9249–9257. <https://doi.org/10.1021/es402120y>.
- Zorba, V., Stratakis, E., Barberoglou, M., Spanakis, E., Tzanetakis, P., Anastasiadis, S.H., et al., 2008. Biomimetic artificial surfaces quantitatively reproduce the water repellency of a lotus leaf. *Adv. Mater.* 20, 4049–4054. <https://doi.org/10.1002/adma.200800651>.